THE CHEMISTRY INSIDE SPICES & HERBS: RESEARCH AND DEVELOPMENT

Editors: **Pankaj Kumar Chaurasia Shashi Lata Bharati**

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The Chemistry inside Spices & Herbs: Research and Development

(Volume 1)

Edited by

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FOREWORD

The book titled "The Chemistry inside Spices and Herbs: Research and Developments (Volume-1)" Edited by: Dr. Pankaj Kumar Chaurasia and Dr. Shashi Lata Bharati has an excellent collection of 10 chapters written by the experts of their subjects from countries like India, Iran, and Egypt. Each chapter of the book, attractively written by the experts, is full of research as well as academically momentous information. This book brilliantly deals with biologically valuable spices, herbs, their related chemistry, biochemistry, structure-activity relationships, biologically as well as pharmaceutically valuable active natural compounds, roles in the natural treatment of various human problems, treatment of neurobiological disorders, roles as antifungal and antibacterial agents, naturally-derived analgesics and anti-inflammatory agents, phenolic compounds, flavonoids, curcumin, turmeric, natural therapy, and so on.

In the present time of pandemic and other problems, when the whole world is searching for various types of immunity boosters to fight this virus, this volume may be helpful in this direction in order to provide in-depth information because there are different types of spices, herbs and their constituents discussed in the book which are radiantly useful in the treatment of various human problems and enhancements of immunity. In my view, after giving a thorough look at the contents, this book may be very advantageous for academicians, researchers and scientists working in the field of spices, herbs, their related chemistry, natural medicinal therapy, and so on. I am congratulating the editors of the book for producing such a useful, academically as well as a scientifically relevant book by compiling the comprehensive chapters contributed by the experts of various countries. I strongly recommend this volume for UG and PG students of life sciences, natural chemistry, biochemistry, natural medicinal studies and scientists working in aforesaid areas.

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PREFACE

Plants are the boon of nature on the earth for us in many ways. They detoxify the environments and save the lives living on this earth. Out of several advantages of plants, their different parts and/or substances are known for their noteworthy medicinal values. Spices and herbs which are involved in our daily routine life are the treasure of good health. Spices, a routine part of the kitchen, as well as herbs of our garden, are full of medicinal virtues and benefits and can be significantly used for the treatment of various disorders and diseases of humans. Spices are actually fruits, seeds, barks, roots and other parts of the plants widely used for enhancing the taste, color and quality of the foods (https://en.wikipedia.org/wiki/Spice) and are the source of various valuable chemical constituents of pharmaceutical significances while herbs are leafy green or flowering parts of the various plants with savory or aromatic properties (https://en.wikipedia.org/wiki/Herb). They are the major source of Ayurveda and other traditional culture of treatments and also have a great potential in the modern time. Spices and culinary herbs and their various chemical constituents involved in the treatment of various problems, diseases and wounds have been beautifully covered in this book.

In the present time of the serious pandemic COVID 19 period, demands of pharmaceutically valuable spices and herbs have been surprisingly enhanced all over the world because they have a substantial and valuable position as nutraceutical which doubtlessly are due to their significant healthy, nutritious and immunity boosting properties. Actually, the main objective of the construction of this book was to collect the more significant valuable researches and information on spices and herbs, which are being widely used in our daily life either in the form of taste enhancing savory materials or quality improving materials or beautiful home decoration and so on. Collection of weighty researches on biologically active pharmaceutically interesting chemical compounds and their compositions and structure activity relationships of these compounds was the second most interesting objective of this book.

This book is full of scientific knowledge on spices, herbs, associated internal chemistry and wide biological performances. It includes biochemistry and biotechnology of spices and herbs, antimicrobial properties, analgesics and anti-inflammatory agents, cure of neurobiological disorders, phenolic compounds, flavonoids, structure activity relationship, biologically active compounds and isolation, and so on.

This volume consists of total ten chapters and each chapter has been written by the various learned experts of their field. Learned experts come from different countries like India, Iran and Egypt. This unique collection of chapters may be highly beneficial for the students of graduate and post graduate level studying in the field of life sciences, biotechnology and biochemistry, plant sciences and for researchers and scientists working research in the field of spices, herbs, compounds with biological activity, natural treatment and natural pharmacology. The book is full of updated knowledge, information and recent researches, and without any doubt, it will be very much fruitful for the readers.

Chapter 1, titled "Spices Biotechnology: Opportunities and challenges", written by Hamid *et al.*, provides an overview of various biotechnological solutions that increase the quality and productivity of spice plants.

Chapter 2, titled "Spices, the guards against the evil microbes: Antimicrobial properties of spices", written by Jacob *et al.*, highlights the effect of various spices on various microorganisms, the various metabolites in spices that lend this ability and also reviews.

Chapter 3, titled "Spices and Herbs in the Treatment of Neurobiological Disorders", written by Trivedi *et al.*, deals with the role of spices and herbs for the cure of neurobiological disorders. Based on the investigations on herbal plants and neurological substrates in disease conditions, herbal medicines can be effectively used in the treatment of various neurological disorders.

Chapter 4, titled "Spices and Herbs in Bacterial and Fungal Resistance", written by Trivedi *et al.*, describes the use of spices and herbs against bacteria and viruses. The use of spices and herbs presents a great potential alternative or supplementary medicine to reduce side effects, progressively increasing the resistance of pathogens induced by the use of allopathic drugs.

Chapter 5, titled "Naturally Isolated Compounds from Spices and Herbs and their Medicinal Uses", is written by Ramteke, A.M. This chapter includes a wide variety of isolated compounds such as phenolic compounds and flavanoids present in spices, which are now experimentally documented to possess antioxidant, anti-inflammatory, antimutagenic and anticarcinogenic activities. It also includes a list of spices compounds that are experimentally evidenced to control cardiovascular diseases, diabetes, cataract, cancer, *etc.*

Chapter 6, titled"Naturally-derived Analgesics and Anti-Inflammatory Agents", written by Fayez *et al.*, covers all the nutraceuticals and phytochemicals – derived from medicinal plants– which have been reported to possess analgesic and/or anti-inflammatory effects over the period between 2018 up to June 2020.

In Chapter 7, titled "Phenolic compounds and their Biological and Pharmaceutical activities", Kumar *et al.* have summarized information on the biological and pharmaceutical activities related to different classes of phenolic compounds.

Chapter 8, titled "Structure Activity Relationship of flavonoids: An update", written by Khare *et al.*, focuses on the majority of polyphenols present in the daily diet, which mainly exist as glycosides with different sugar units and acetylated sugars at different positions of the polyphenol skeletons.

Chapter 9, titled "Biologically active compounds and Structure-Activity Relationship" has been written by Ganatra, S.H. He has discussed all three methods in detail, along with examples. It also provides the practical procedure to use available computational tools. The final aim of this chapter is not only to provide the theoretical background of drug discovery using structure activity relationships, but also to provide practical methods.

Chapter 10, titled "Turmeric Supplementation and Its Valued Clinical Connections", demonstrates the renowned significance of turmeric in the treatment of various health issues and its role as a food supplement concisely.

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Spices Biotechnology: Opportunities and Challenges

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Abstract: Spices have been used since ancient times as a flavoring agent as well as an important medicinal resource. Biotechnology, using strategies such as cell, organ, and tissue culture, genetic engineering, and the application of nucleic acid markers can escalate the productivity and efficiency of spices. Cell, tissue, and plant organ culture have enabled the rapid and mass reproduction of many disease-free spice plants, which are uniform genetically and qualitatively. In recent years, cell and limb suspension (stem and hair roots) have been considered for producing secondary metabolites and for studying the biosynthesis pathway of metabolites. Plant genetic engineering has helped in the genetic identification and manipulation of enzymes of the biosynthetic pathway of secondary metabolites. Gene transformation has improved the production of secondary metabolites that have yield limitations. Molecular markers are powerful tools for accurately identifying important medicinal species, examining genetic diversity, classifying hereditary reserves, and determining their genetic map irrespective of their age, physiological, and environmental conditions. Next-generation sequencing (NGS) methods like restriction-site-associated DNA sequencing (RAD-seq) have revolutionized the study of genetic diversity, and the enzymes and genes implied in the secondary metabolites biosynthetic pathways can be studded by transcriptome profiling (RNA-seq). The ground-breaking genome editing techniques like Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR), sequence-specific nucleases of transcription activator-like effector nucleases (TALENS), and zinc-finger nucleases could help in customizing the plants according to the requirements. This article provides an overview of various biotechnology solutions that increase the quality and productivity of spice plants.

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INTRODUCTION

Spices are mainly the aromatic parts of plants that have been dried. The Food and Drug Administration (FDA) has defined spices as: "aromatic vegetable substances in whole, crushed, or ground form, the notable characteristic of which in food is preparation as opposed to nutrition." [1]. Flavors are regularly derived from the dried part of plant-like buds, barks (cinnamon), fruits/berries (cloves, black pepper, chili), blooms (cloves, saffron), seeds (cumin), or roots (ginger, turmeric) that contain unstable oils or fragrant scents and aromas [1, 2]. The majority of the well-known spices and herbs come from Asia, the Middle East, or Mediterranean countries and have been used since ancient times [3]. Spices and herbs have occupied, and still occupy, significant roles as seasoning specialists, food additives, and meds for quite a long time. Over the last few decades, the investigation into their medical advantages has expanded essentially; the same number of spices and flavors are considered to have properties that reduce the risk of chronic disease development. Specifically, a few of the potential wellbeing benefits of herbs and flavors include conferring security against cancer, chronic inflammation, cardiovascular illness, type 2 diabetes, neurodegenerative conditions and obesity [3 - 6]. Several herbs have been renowned for their antiinflammatory, antioxidant, and anti-microbial properties [7, 8]. Additionally, the use of certain herbs and flavors will help in reducing the use of salt as the sole flavoring agent (*i.e.*, lower sodium admissions), which has cardiovascular benefits [9]. Black pepper, turmeric, clove, vanilla, cardamom, nutmeg, ginger, cinnamon, tamarind, etc., constitute the major flavors, whereas fennel, fenugreek, coriander and cumin are imperative seed flavors. While anise, celery, lavender, oregano, saffron, sage and thyme are critical homegrown flavors. The transcriptomes of Piper player nigrum and Piper player colubrinum were analyzed to understand the host-pathogen activity in black pepper, with a focus on Phytophthora foot rot tolerance. The productivity of spices is poor, owing to the lack of high-yielding, pest and diseases resistant varieties, and also due to postharvest losses. Ordinary breeding programs were found to be time-devouring and lumbering in perpetual flavors, such as cardamom and black pepper. Dearth of sources of biotic and abiotic stress resistance within the evolved germplasm made the process even more arduous. Furthermore, crops like ginger and turmeric have no or very few seeds, rendering traditional breeding systems ineffective. Creating varieties with high yielding and disease resistance, under such circumstances, through biotechnology, is imperative for the improvement of spices. The use of biotechnological methods to achieve the above has increased dramatically in

recent years through marker-assisted breeding, development of novel varieties, and commercial propagation.

COMPARATIVE GENOMICS AND GENE TAGGING

Comparative genomics compares various genomic features like genes, regulatory sequences, DNA sequence, gene order and various genomic structural landmarks of several organisms. A crucial step in breeding is recognizing the loci of beneficial genes (high yield, quality, cost-efficiency, and pest and disease resistance). It may be a capable and swift strategy since it does not necessitate several generations of closely supervised parent strain breeding [9]. The detailed transcriptome of Piper nigrum and Piper colubrinum was conducted w.r.t hostpathogen interaction in black pepper with more focus to the Phytophthora foot rot tolerance [10]. The root transcriptome sequencing of black pepper [11] was done by the SOLiD platform and a detailed dataset of 10,338 UniGenes was found to be crucial for the molecular breeding of black pepper. The 4472 anticipated proteins appeared to have approximately 52% homology with the Arabidopsis proteome. The comparative proteome analysis of two roots revealed 615 differentially expressed proteins [12]. Hu, Hao [13] depicted the black pepper fruit transcriptome in conjunction with the piperine biosynthetic pathway and found 40,537 UniGenes included in piperine biosynthesis. The molecular mechanisms underlying foot rot susceptibility were understood by comparing the transcriptome of resistant (*Piper flaviflorum*) and susceptible (*P. nigrum* cv. Reyin-1) species. It was observed that the genes consolidated within the phenylpropanoid metabolism pathway were highly up-regulated in resistant species [10]. Karthika, Prasath [14], compared the ginger (Zingiber officinale Rosc.) and mango ginger (Curcuma amada Roxb.) transcriptomes in response to bacterial wilt infection and they observed that 105 genes were only expressed in C. amada (safe species) in reaction to contamination by Ralstonia solanacearum. These genes were linked to pathogen defence through hypersensitive, systemic acquired, and cell death responses mediated by salicylic acid (SA). Out of the 54 differentially expressed transcription factors, 32 showed upregulation in C. amada, which includes GATA, WRKY, zinc finger, MYB and leucine zipper protein domain transcription factors. The transcriptome of two samples of the elite ginger variety Suprabha obtained from two separate agro-climatic zones of Odisha was analyzed by Gaur, Das [15]. The novel transcripts coding for terpenoids related to anticancer and antimalarial in the transcriptome of Curcuma longa was reported by Annadurai, Neethiraj [16]. Comparative transcriptome (rhizome-specific) evaluation of C. longa and Curcuma aromatica associated with curcumin content provided information about the genetic basis and regulation of curcumin biogenesis [17]. Differential expression analysis identified two novel polyketide synthase genes (clpks1 and clpks2), which showed similarity to *Musa acuminata*

polyketide synthase type 2 (MaPKS2) and M. acuminata polyketide synthase type 4 (MaPKS4) that were found to be upregulated in C. longa [17]. Babu, Jose [18] analyzed the transcriptome assembly of the turmeric variety Suvarna (CL-Suv). The transcriptome from seeds, leaves, and flowers of Coriander (Coriandrum sativum L.) was sequenced and analyzed by Tulsani, Hamid [19], 8676 unigenes were assigned to 153 KEGG pathways in this study. Among them, 291 unigenes were related to terpenes biosynthesis. Paul, Mathew [20] explored the possibility of using comparative transcriptome analysis to point out the candidate genes responsible for the black pepper foot rot field tolerance. DD-RT PCR on cDNA fragments was used to compare transcriptome profiles, and the bands that were differentially expressed were sequenced. Sequence analysis showed the participation of signal proteins and defence enzymes like Aspartyl protease, betaglucosidase enzyme, Cytochrome P450 signal protein, Nitrous oxide reductase family maturation protein, nucleoredoxin 1-1 enzyme, Phosphatase 2C-like domain-containing protein, Premnaspirodiene oxygenase, putative disease resistance protein RGA3 and Serine/Threonine Protein kinase WAG1 in field tolerance of black pepper to foot rot. Additional insights into the molecular function of tolerance were acquired by pathway analysis. Jiang, Liao [21], analyzed the transcriptome and phytohormone profiles of ginger (Zingiber officinale Rose) in reaction to postharvest dehydration stress. Transcriptome profiling found out a total of 1415, 2726, and 6641 genes were differentially expressed after 2 h, 12 h, and 24 h of water-loss stress treatment, respectively in comparison with that during zero h of ginger rhizomes. Moreover, 518 DEGs shared comparable expression patterns throughout twenty-four h of dehydration stress. These genes are specifically enriched in plant hormone signalling, carotenoid biosynthesis, starch and sugar metabolism, phenylalanine metabolism, fatty acid elongation, and phenylpropanoid biosynthesis.

Cloning and Genes Isolation

Genes involved in biotic and abiotic stresses and agronomically critical characters were distinguished in most spice crops [22]. Pathogenesis related candidate genes may also be distinguished using sequence data from libraries, extracted, and then integrated into promising varieties utilizing transgenic techniques. A family or genus, wild relatives of crops may have a set of genes for various biotic and abiotic resistance, agronomically important characteristics, *etc* [23]. Since hybridization based breeding programs to mobilize genes from wild relatives are challenging, the transgenic approach to join the genes is preferable.

Genetic Transformation

Diseases, a lack of resistant varieties, and post-harvest declines are the main

causes of lower spice yields [24]. Genetic transformation has great potential to overcome restrictions of conventional breeding methods and produce high yielding and disease resistant transgenic plants [25]. Plant transformation is considered as both a basic scientific method in plant biology and a practical tool for transgenic plant advancement [26].

Gene transformation is a powerful tool for increasing productivity. There are various methods for gene transformation; such as *Agrobacterium-tumefaciens* transformation, particle bombardment, and electroporation for gene transfer on herb and spice plants, but there are two fundamental classifications for gene delivery: biological and non-biological system [27].

NON- BIOLOGICAL GENE TRANSFORMATION SYSTEM

There are several non-biological systems, which are used for gene delivery *via* plant or protoplast. Non-biological systems like chemical treatment of isolated protoplasts by PEG, electroporation, lipofection, or fusion of protoplasts with liposomes, microinjection, and biolistic. In a direct gene transformation system; chemical solution including PEG, Polyethylene glycol (generally is used only PEG) is incubated with DNA fragment and protoplast. Protoplast is the most appropriate explant in this technique. Due to accessibility and simplicity, this protocol has been reported in numerous plants [28, 29]. There are some reports of using protoplast fusion mediated (PEG mediated) for production of abiotic or biotech disease tolerance or somatic hybridization in vanilla species [30], ginger [31], and coriander [32].

Lipofection mediated transformation involves liposomes (as artificial circular lipid with an aqueous interior for carrying DNA fragment), which can be stimulated *via* PEG to integrate into protoplasts [33]. A sudden electrical discharge for creating small pores in the plasma membrane is used in the electroporation system for the transformation of DNA to protoplast. Transformed protoplasm has the potential to regenerate transgenic plants. Electroporation is introduced as a reproducible system if a good quality protoplast is produced. In the microinjection method, DNA fragment is transferred mechanically to a specific target, which normally is the protoplast. The process is applied through a glass micro capillary-injection pipette. Using a micromanipulator is not practical for transformation in the plant due to the presence of the cell wall, however, it has been effectively used for the transformation of large animal and human cells [29, 34]. Although used rarely for gene transformation, biolistic gene transformation is an alternative non-biological method and has been referred to as an important and famous method for gene transformation to spices plants.

Biolistic Micro-Projectile Bombardment Gene Transformation

The micro projectile bombardment method (also mentioned as particle bombardment, particle gun method, particle acceleration, and biolistic) has been widely introduced as a routine, reliable, and physical gene delivery system [33]. In this method, DNA or RNA gene is coated on microinjection (which normally is tungsten or gold with the size of 1-4 m) then bombarded into callus explants. Micro-carrier size, explant target distance, and helium bombardment pressure and the constructs (circular or linear plasmid) used are factors affecting the efficiency of biolistic transformation. Among the various explants such as microspore, pollen, and shoot meristem reported as explants, embryogenic callus has a higher potential for uniform regeneration after the bombardment and has been considered as an optimum explant for biolistic gene transformation [35]. There are numerous reports of reproducible transformation protocol in capsicum [35], ginger [36], turmeric [37], cumin [38] *via* biolistic system. However, multi-copy integration, which causes transgenic silencing, has been reported as a major concern [39].

Biological System Agrobacterium-Mediated Gene Transformation

Agrobacterium tumefaciens mediated transformation is a natural mechanism for gene transformation in numerous plants. Even though, there are various approaches for gene transformation, the use of Agrobacterium is superior and more popular than other methods especially in dicotyledonous plants, due to more efficiency with lower cost, reproducibility, high capacity to transfer large inserts of DNA, and low copy number. This technology has been widely used for gene transformation (stable or transient) in many spices [40].

IN PLANTA GENE TRANSFORMATION

Another way to use agrobacterium in gene transformation is *in planta*, *i.e.* DNA transfer directly in the intact plants without using tissue culture methods [41]. This minimizes somaclonal variation and saves time significantly, decrease the costs and labour. The pollen tube pathway method is an in planta method that is effective only after pollination in plants. The DNA transformation process takes place by cutting the styles then using a syringe to transfer the DNA material down the pollen tube. This technique was successfully used in black pepper for improving *Phytophthora capsici* resistance [42]. Several genes have been transferred to spice plants *via* biological and non-biological methods for various purposes, which is discussed below (Fig. 1).

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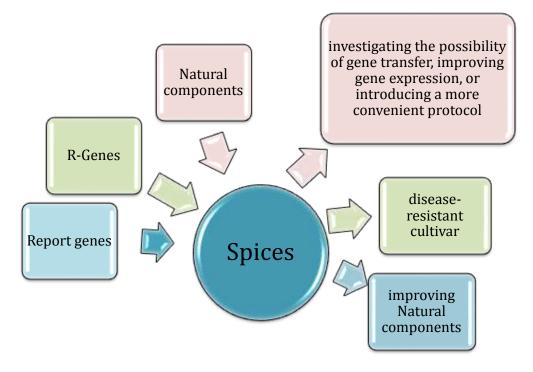


Fig. (1). Important purposes of gene transfer *via* biological and non-biological gene transformation systems and their results in spice plants.

Introducing a More Convenient Protocol

Varghese and Bhat [43], reported an efficient Agrobacterium-mediated gene transformation method in black pepper using somatic embryo explants for and GUS reporter gene. They succeeded in regenerating 9 plants per gram of embryo genic mass for the first time without using growth regulators and any genetic variation [43]. Sinojo *et al* 2014, also optimized somatic embryogenesis methods for Agrobacterium-mediated genetic transformation of a pathogenesis-related gene (PR5) in black pepper [44]. Compared with other solanaceous crops, pepper varieties (*Capsicum annuum*) are highly recalcitrant, so they have shown a very poor response toward transformation by Agrobacterium and regenerative capacity [45, 46]. In capsicum varieties, transformation frequency and shoot regeneration rate are highly genotype-dependent, also Agrobacterium-mediated transformation rate was low for cut-injured cotyledon and hypocotyl [47, 48].

A protocol for generation and gene transformation of two elite Indian cultivars of chili pepper (*Capsicum annuum* L.) was established through *Agrobacterium tumefactions*, strain LBA4404 containing pCAMB1A2301 plasmid for expression of GUS and NPT-II as reporter and marker genes respectively. Results of GUS

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assay, PCR, Southern blotting as well as RT-PCR analyses confirmed transformation [49].

Due to the lack of seed set in ginger, there is a high limitation of diversity in its gene pool. Moreover, all breeding programs and vegetative reproduction via rhizomes lead to the spread of soil-borne diseases [50]. However, there are many successful reports of ginger (Zingiberaceae) transformation via Agrobacteriummediated and biolistic methods. The opening report of gene transformation of ginger was reported *via* biolistic methods on embryogenic callus as an explant for GUS expression [51]. Successful transformation with the biolistic method through protoplast explant of ginger was published with high GUS gene expression [45]. In Agrobacterium-mediated methods, two strains of Agrobacterium LBA 4404 including p35SGUSInt and EHA 105 with binary vector pCAMBIA1301 containing GUS reporter were used. Gene transformation stability was confirmed by PCR [25]. High transformation efficiency in Agrobacterium transformation was reported in a new quick transformation protocol by using LBA4404 strain containing pGFPGUSPlus when the explants incubated with Agrobacterium for 2 days as the co-cultivation stage [26]. In comparison with ginger, there are a few reports of gene transformation in turmeric. He and Gange, (2013) reported two-development transformation systems (leaf-based transient expression and callus-based stable expression) via Agrobacterium transformation. Agrobacterium strain EHA105 consisting of plasmid pBISN1, optimized for both transient and stable transformation. Transgenic plants were confirmed by PCR, Southern blot as well as GUS essay analysis [52]. There is a report of using biolistic as an alternative transformation method for Capsicum species too [53, 54].

Natural Component Gene

There are several diseases, which reduce performance in spice and herb plants and cause annual losses around the world. In this section, some of these diseases, as well as the solution proposed by genetic engineering, are mentioned.

As some of the spice plants (such as Turmeric and Ginger) have underground rhizome, they are vulnerable to accumulate pathogens and are susceptible to soilborne diseases. Pepper varieties (Capsicum) are susceptible to numerous pathogens counting bacteria, fungi, viruses, and nematodes. So some approaches are aiming at the production of red pepper transgenic with high resistance [55].

Appropriate conventional crop improvement methods in the field of disease resistance in plants are problematic and insufficient [37]. Genetic engineering methods by identifying candidate R-Genes (resistance-Genes), cloning, and transformation, are suggested as the novel solution to obtain disease-resistant cultivar [55]. Joshi *et al.*, 2010 isolated five NBS-LRR resistance gene candidates,

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via generated primers based on conserved domains of resistance genes. They suggested this NBS analogs can be a guideline for isolating more R-Gene in wild relatives turmeric for the genetic improvement of Curcuma [56]. In another report, by using molecular genetic methods, tree R-Gene was found out to be the most stable reference genes for developing Phytophthora- resistant black pepper [24, 42].

Furthermore, there are many available reports for validation, cloning, or expression of the genes related to defense mechanisms against various diseases in spices. Primary genes, expressed in black pepper *via* Agrobacterium-mediated transformation, were NPT II gene (neomycin phosphotransferase) and GUS gene in 1994 and 1998 respectively. There are some reports of the transformation of CP genes (as the genes resistance to CMvirus and ToMvirus) through Agrobacterium-mediated transformation to chili pepper [46, 57]. Gene of BC1 (linked with chili leaf curl Joydebpur virus) was induced in hypocotyl explants of six deferent cultivars of red pepper, by a methodical Agrobacterium-mediated transformation protocol. Transgenic lines were validated by PCR and Southern blotting analysis [58].

The primary efficient biolistic gene transfer method in turmeric was reported for the transformation of plasmid pAHC25 that included by the bar (Glufosinate) as an herbicide gene and GUS reporter. The stability of transformation was confirmed by the results of the GUS assay and PCR analysis [59].

Plant Tissue Culture

Plant tissue culture involving *in vitro* direct or indirect regeneration from various explants is a fundamental approach to take advantage of biotechnological applications in plants [60]. Progress in plant tissue culture has led to the development of other biotechnological methods. In the case of spice plants *in vitro* method has generally been used for overcoming the poor germination seed problem and improving mass propagation, producing disease-free plants and germplasm conservation.

Mass Propagation

The most crucial factors for plant mass propagation efficiency are genotypes, growth regulators, the culture medium, and the physical factors. The different parts of a plant such as leaves, terminal buds [61], bulblet [62], rhizomes [63, 64], stem and root fragments [65], have been studied as explants for spice micro-propagation. In addition, depending on the genotype, different compounds of growth regulators affect productivity [66, 67]. Numerous *in vitro* techniques led to established several efficient protocols for large-scale *in vitro* propagation in

various spices. Using different dosages of cytokinins such as BA or 2ip, were reported for improving shoot regeneration in black pepper [68], ginger [65], garlic [62], turmeric [69]. An efficient protocol for micro-propagation of large cardamom was established by culturing rhizome buds as explant in MS medium containing 1 mg each of BA and IBA (tissue-cardamom3). *In vitro*, culture methods have been able to improve reproduction in garlic also in MS medium containing 2ip and NAA for proliferation step [62]. In Myrtle micro-propagation, modified WPM medium supplied with BA and IBA was reported as an optimum culture medium in comparison with MS and applying different concentrations of IBA or NAA were used for rooting step in another report [70]. A high rate of *in vitro* propagation of curcuma (almost 18 shoots) was reported by using thidiazurone as a growth regulator in MS medium [71].

Somatic Embryogenesis

Effective and developmental production of somatic embryos is a prerequisite for commercial crop production. Somatic embryogenesis is the process by which somatic embryos develop from a group of somatic cells or tissues. These embryos are similar to zygote embryos (embryos from sexual fertilization) and can be transformed into seedlings in a suitable culture medium. Plant reproduction using somatic embryogenesis from a single cell has been demonstrated in many spices and herbs. Therefore, in this case, according to the different potential in different cells for the production of natural compounds, plants with superior characteristics can be produced compared to the primary plant. Most of the reports confirmed that decreasing the concentration of growth regulators in culture medium improves somatic embryogenesis. 2,4-D is referred to as an important auxin for callus induction and somatic embryogenesis. A blend of 2,4-D and with a cytokinin, same as BA on MS medium, has a progressive effect on somatic embryogenesis and callus induction in spices. In ginger, a high number of somatic embryos, 87.7% and 93.3%, were formed by indirect and direct culturing in MS liquid medium using a combination of 2,4-D and BA via leaf sheath explants, respectively [72, 73]. Guo and Zhang reported the somatic embryogenesis of four ginger cultivars by cell suspension culture in liquid MSN medium containing 2,4-D and Kinetin [74]. The first report for direct somatic embryogenesis of turmeric with 91.1% efficiency was reported via using solid MS medium containing 2,4-D in dark condition and liquid MS medium with BA [48]. High-frequency black pepper plantlet regeneration via somatic embryogenesis was reported in several protocols [75, 76]. Application of endophytic fungi in somatic embryogenesis culture for promoting growth and hardening of in vitro cultured plants was established in Black pepper [70]. The highest somatic embryogenesis frequency (100%) was reported in *Panax notoginseng* in liquid MS medium contusing 2,4-D via Bioreactor cultures [77].

In-vitro Culture by Bioreactors

Automation of the micropropagation process can play a major role in overcoming the limitations of conventional laborious methods. Bioreactors are widely used for producing microbial, animal, or plant metabolism. Although applying bioreactors has been largely intended for cell suspension or hairy roots of spices and herb plants, the optimization of bioreactors for embryogenesis and tissue or organ culture has been reported in the number of studied spices [78]. The temporary Immersion (TI) system is a famous kind of bioreactor for tissue and organ culture. AKA et al., 2019, reported the optimized protocol for Myrtle micropropagation and rooting by TIB. The efficiency of Myrtle plantlet in all growth factors (Number of roots, plantlet and root length, root fresh and dry weight) in TIS was better when compared to the conventional method [79]. TI system was used for the mass improvement of the propagation of Vanilla also [80]. Three kinds of bioreactor systems were compared for micropropagation of Vanilla planifolia, TI, and RITA systems were introduced as a suitable system for commercial mass propagation and reduction of cost and labour in this spice respectively [77]. The same experiment was carried out for improving shoot and bulblet generation in garlic. However shoot propagated performance was significantly upper in the CI system, the BI system was introduced as an optimal system for bulblet formation in garlic [81].

In vitro Conservation and Cryopreservation

It is important to slow down the growth of spice shoots for the maintenance of their germplasm. In vitro conservation is one of the reliable methods for the maintenance of different vegetatively produced plant germplasm [82]. Increasing the concentration of sucrose in rhizome formation medium, using different concentrations of macroelements including EDTA and iron in MS medium and various kinds and amount of gelling agents are *in vitro* approaches reported for extending conservation period in spices [83]. In a successful report of in vitro turmeric conservation, low-cost medium (up to 73% cost reduction) including commercial sugar and bacteriological agar as a carbon source and gelling agent were used respectively. In vitro, conserved turmeric after 12 months does not have any significant variation in their RAPD profile when compared to the mother plants [84]. Primary *in vitro* conserved cardamom plantlets were achieved using ¹/₂ MS medium without growth regulators and decreasing osmotic potential in the culture medium. In a subsequent study, the efficiency of carbendazim as a fungicide on the conservation of Curcuma and ginger shoot explants was reported. The genetic stability of conserved plants was confirmed by the RAPD profile after 3 years [85]. There are several successful reports for in vitro clonal micropropagation and conservation in ginger and turmeric [86 - 88] also.

One of the important approaches to micropropagation is cryopreservation [89]. Cryopreservation refers to the storage and degradation of germplasm usually in liquid nitrogen at -196°C. During this time, all cell division and metabolism operations are stopped, and germplasm can be maintained safely without any genetic changes. *In vitro* maintenance of some spices germplasm such as wasabi [89 - 91], garlic [92 - 94], piper [95, 96], ginger [96, 97], *via* cryopreservation is increasingly applied. The cryopreservation technology for black pepper, cardamom, turmeric, and their germplasms using methods like vitrification, encapsulation, and encapsulation-vitrification methods is available [98 - 101]. Cryopreservation of Coriander (*Coriandrum sativum* L.) somatic embryos using air desiccation and sucrose preculture was reported by Popova, Kim [102]. González-Benito and Iriondo [103], also used LN 2 for Celery Cryopreservation.

Secondary Metabolites Production

Secondary metabolites are complex chemical organic matter that plants produce during their lifetime; however, they do not have any important role in their growth and vital activities, mainly produce against biotic and abiotic stresses or attracting pollinating insects. Mass production of these natural components on a large scale through chemical methods is mainly "difficult or impossible". Appling tissue culture methods like cell suspension cultivation, organ culture, and polyploidy induction are suitable solutions for the rapid and mass secondary metabolites production in plants.

There are available reports for enhancing natural components in spices by micropropagation. The significance of various MS salt concentrations, as well as sucrose were evaluated on four major volatile constituents of *Chenopodium ambrosioides* L. *in vitro* condition. The results showed that all four natural compounds have changed under the influence of changing culture medium [104]. Another successful protocol for *in vitro* culture of *Spilanthes acmella* MURR *via* shoot tip explants was given recently [105, 106].

In most plant species, the induction of polyploidy by increasing cell size has created the ability to produce stronger vegetative organs. Growth organs are the source of a variety of commercially valuable secondary metabolites. Therefore, it is possible to induce polyploidy which can play an imperative role in improving the quantity and quality of these valuable compounds [107]. A significant rise in the production of secondary metabolism has been observed in comparison with numerous polyploidy plants with their diploid counterparts, such as Astragalus [108], Artemisia [109], Jujube [110], Lemon balm [111]. Colchicine is the most important chemical agent in chromosomal doubling, which is widely used in spice and herb plants. Colchicine inhibits the formation and polymerization of

microtubules through binding to a microtubule protein, called tubulin; hence chromosomes enter the cell together at the metaphase stage, making it an active polyploid inductor [112]. In various experiments, the range of 0.01 up to 0.5% has been reported as an optimal concentration for colchicine [108].

Agrobacterium rhizogenes soil born Gram-negative bacterium is a principal agent for Hairy root disease. The infection by the bacteria culminated in production of hairy roots near the site of bacterial entry. Hairy root induction has been tried on various spices plants, hence resulted in an upturn in the production capacity of metabolites by them. Rapid growth, short duplication duration, and having more efficiency for the production of the various natural component of hairy root make them a permanent source for the secondary metabolites production. Many available reports for the usage of hairy root culture for secondary metabolites production such as; Sotolon from *Trigonella foenum* in airlift bioreactor [113], Sarpagin alkaloids from *Rauvolfia serpentine* [114, 115], α -phellandrene and apiole (as an essential oil from) in dill [116].

Protoplast Culture

Protoplast is a plant cell in which the cellulose wall has been removed. In other words, protoplasm has only a thin plasma membrane that surrounds the cell. Plasma has many applications in direct and indirect DNA transformation through electroporation and PEG mediated transformation as well as in the transient system. Protoplast culture has been reported successful in spices. Effective protocol in protoplast culture from cell suspension and leaf tissue of turmeric, cardamom, black pepper, and ginger, from the root of fennel, from mesophyll of fenugreek, from the shoot of garlic, have been elucidated [117].

Molecular Markers

Germplasm diversity is essential for a successful breeding program. Variation is significant for the increase in the genetic base since it raises the chances of discovering dynamically exceptional genes for which the alleles from the two parents are different (that is, the genetic distance) [118]. DNA markers are a powerful tool for distinguishing spice species effectively, as they are independent of age, physiological and environmental conditions. The profile obtained from the DNA fingerprint of a spice plant is the same. Also, the physical shape of the sample is not important for its evaluation and in addition to fresh tissue; it can be extracted from the dry tissue of the DNA. For species or varieties of medicinal plants that are morphologically and phytochemically similar, DNA markers are very important, as they can be used to accurately differentiate. Several kinds of nucleic acid-based markers, like RFLP, RAPD, AFLP, SNP, and SSR are used to study the genetic structure of organisms (Table 1). In last years, many studies

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have been performed to find out the relationship between DNA markers and quantitative and qualitative variations of active drug compounds among species and near relatives of spice plants (Table 2). In order to study geographical origins and investigation of interrelationships of Indian cardamom, the molecular level profiling of 11 species including 5 major tribes, viz., Amomum, Alpinia, Aframomum, Elettaria, and, Hedychium as well as 96 collections of cardamom germplasm using RFLP, ISSR and RAPD markers was performed by Babu, Divakaran [119]. Tamayo 2007 performed the molecular profiling of chosen cardamom genotypes in Columbia using AFLP molecular markers. The genetic diversity among the various species was appraised using various strategies (Table 2), in addition to conventional molecular markers. Next-generation sequencing (NGS) based genotyping approaches are being used of late in whole-genome sequencing and re-sequencing research programs. An enormous number of singlenucleotide polymorphisms (SNPs) identified from multiple specimens by sequencing can be used to investigate within-species polymorphism, establish haplotype maps, and conduct genome-wide association studies (GWAS). NGS has made the tedious screening of plant germplasm feasible and cost-effective [120]. Using SNP markers has been pretty powerful due to its abundance in plants, costeffectiveness, the flexible technique, little error rate and high speed of detection [121]. GBS (genotyping by sequencing) is a modern method that uses secondgeneration sequencing methods to classify and represent SNPs in a smaller scale genome-wide [122]. Using restriction enzymes in GBS reduces repetitive regions and thus the genome complexity, resulting in the most rapid bioinformatics analysis for large genomes [123, 124]. Therefore, this technique, is a rapid, genome-wide, high-throughput, and cost-effective method for SNP finding [125].

Category	Marker name	Features	
Hybridization based	RFLP	-Co-dominant -Unlimited number of loci generated Conveyable across population and species -A large quantity of DNA needed, frequently results in less levels of polymorphism	
PCR based	AFLP	-Dominant -Small DNA quantities required -High levels of polymorphism generated -Dominant Complicated methodology	
	RAPD	-Dominant -Fast Results -Requires less DNA quantities -High genomic abundance -Low repeatability within and across laboratories	

Table 1.	Most	usable	molecular	markers.

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Category	Marker name	Features
PCR based	SSR	Co-dominant -Highly polymorphic -Technically simple -Species-specific
	ISSR	-Dominant -Highly polymorphic -Species-specific
	STS/EST	-Co-dominant/Dominant -Highly reliable -Small DNA quantities required -Prior sequence information required
	SNP	-Co-dominant Suitable for high throughput technology -Very high development costs, Sequence information required
	SCARS/CAPS	-Co-dominant -Small DNA quantities required, Highly reliable -Sequence information required

Table 2.	Application	of differen	t molecular	markers in	spices.
I abic 2.	reprication	i or uniteren	t morecular	mai kei 5 m	spices.

Molecular markers	Spice: aim and goal of the research	References
RAPD	Black pepper: Evolution of genetic variability and identification of important cultivars, varieties and related species; evolving mapping population for preparation of the genetic map	[135 - 138]
RAPD	Turmeric: Developing molecular genetic fingerprints; assessing genetic diversity	[82, 139, 140, 140]
	Coriander: investigation and assessment of genetic diversity	[141, 142]
	Cardamom: Associated with katte resistance	[143, 144]
	Basil: Assessing genetic variability	[145, 146, 147].
	Fenugreek: Assessing genetic diversity	[148, 149, 150]
	Lavender: Assessing genetic diversity	[151, 152]
	Ginger: Assessing genetic diversity	[153]
AFLP	Black pepper: study of genetic variation and identification of vital cultivars and varieties	[154]
	Fenugreek: Assessing genetic diversity	[155, 156]
SSR/ISSR/EST SSR	black pepper: Developing new EST-SSR markers in black pepper, Evolution of genetic diversity, exploration of important varieties	[157, 51, 158]
	Turmeric: Developing molecular genetic markers; studying genetic diversity	[159, 160]

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Molecular markers	Spice: aim and goal of the research	References	
	Coriander: Developing molecular genetic fingerprints; genetic variation investigation	[19, 161]	
	Cumin: genetic diversity examination, introducing vital varieties	[162, 163]	
	Large Cardamom: Analyzing genetic diversity	[164]	
	Garlic: Assessment of genetic variability	[165, 166, 167]	
SNP/SCAR	Garlic: Characterization of bolting behavior in garlic	[168]	
	Black pepper: <i>Phytophthora</i> resistance responsible gene identification	[169]	
	Saffron: Assessing genetic variability	[170]	

This technique could aid in genotyping genomes without prior knowledge also it is useful for plant genetic diversity investigation in genome-wide spectrum [126]. Recently, GBS has been utilised in exploring the genetic heterogeneity of many crop species, such as capsicum, barley, maize, sorghum, soybean, tomato, and wheat [127 - 134]. Employing molecular markers, transcriptome and genome sequencing, qRT-PCR approaches can aid classical methods of breeding through the clonal selection and improving elite genotypes.

DNA BARCODING TECHNOLOGY

DNA barcoding is a new molecular recognition tool in which short genomic DNA fragments are used as and identifier marker in different species. Paul Hebert in 2003, firstly suggested this technique, wherein a comprehensive barcode is habituated by DNA screening; a DNA barcode database and recognition platform are settled, and the DNA data are analysed and compared by bioinformatics analysis to identify species [171, 172]. The use of DNA barcoding has helped overcome the limitations of conventional morphology-based identification methods that bank on long-term skills. In due course, instinctive identification might be possible. Barcoding DNA is a step forward and an effective alternate to classic biological characterisation methods [173, 174]. Chen et al. studied the variabilities of nuclear gene sequences as well as plastid genomes, of herbal plants and their closely related species and generated a medicinal plant DNA barcode investigation system [175, 176]. Afterwards, Wang et al. showed that the ITS2 sequences can be successfully utilized to identify various types of components of soybean pods, which can serve as a new technique for certifying clinical drug safety [177, 178]. M. Zhang, et al., used the ITS2 and psbA-trnH sequences for developing a DNA barcoding technique for the verification and uncovering of adulterants in powdered spices. The ITS2 and psbA-trnH sequences effectively distinguished sixteen types of spices and their usual adulterants. A significant

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degree of adulteration was observed in the adulteration detection test of 91 commercially available powdered spices. Congeneric plant, vegetal admixture, or cheaper crop-based products were the commonly detected adulterants [179]. The DNA barcoding method is thus a powerful tool for the regulation of the spice market.

CRISPR–Cas9 System for Engineering Resistance to Viruses Infecting Spices

Modern preventive methods are used to manage viral diseases. The control of viral vectors, the development of virus-free plants using different methods, and quarantine regulation are the preventive measures used. Nonetheless, these controls have limitations as the virus vectors develop resistance to pesticides [180]. The recent methods consist of pathogen-derived resistance, RNA interference-mediated resistance, and ribozyme-mediated resistance. Since the last few years, clustered, regularly interspaced, short palindromic repeats (CRISPR)–Cas 9 tools have been used for targeted silencing of viral pathogens in plants. With this technology, it is possible to simultaneously target multiple viruses at various sites and the results are quite favourable. CRISPR is widely distributed in bacterial and archaeal genomes and provides defence against invading viruses and plasmids [181]. The CRISPR locus has short repeats of prokaryotic DNA interspersed with short segments of 'spacer DNA' from the bacterial virus or plasmids they were previously exposed to. CRISPR spacers identify and cleave these exogenous genetic elements like RNA interference in eukaryotic organisms. This interference technique has massive potential and applications like altering the germline of humans, animals, other organisms, and plants [182]. The Cas9 protein and guide RNAs are delivered into the cell so that the genome can be specifically cut at any desired location. Thus CRISPR-Cas system can be efficiently used to develop resistance to DNA and RNA plant viruses through editing or introducing novel traits, precisely at the loci of interest, into plants [183]. It can also be used for manipulating the host genome itself to insert viral immunity. To date, there are only a few reports of using this technology in spices genome editing, Costa et al used CRISPR-Cas9 based strategy to engineer Saccharomyces cerevisiae for producing curcumin from ferulic acid [184]. Thus, this method has great potential to overcome viral and bacterial diseases in spices.

CONCLUSION AND FUTURE PERSPECTIVES

The availability of nutritious food to nourish the ever-growing population is crucial. Conventional breeding is inadequate to improve the growth and yield of these forgotten plants. Biotechnology based breeding methods (BBBMs) are the solution for high throughput improvement of spice plants in a rapid ways.

Biotechnology can be a key device to accomplish maintainable farming and agriculture-based industry, by the progress of food creation in terms of amount, quality, and wellbeing and at the same time protecting the earth. There has been noteworthy advancement in the field of biotechnology for molecular characterization, micropropagation, and protection, and genetic resources management, management of infections, diseases and pests. Distinguishing markers connected to significant agronomic characters will help in MAS to cut short reproducing time. The utilization of recombinant DNA innovation for biotic and abiotic stress tolerance needs a lot of research before they can be adequately used. Although projects have been started in numerous research facilities for in *vitro* optimal metabolite creation, these methods are to be refined and scaled up for conceivable mechanical creation of the items. Due to their business potential, strengthening, and using biotechnology in spices will be significant in the coming decade. Microbial mediation through T. harzianum, T. viride, P. florescens, and AM fungi has been discovered powerful in disease repression and plant growth promotion. Induced systemic resistance (ISR) as mirrored in defence response seems to be one among the modes of action in disease suppression in ginger and black pepper. The farming community is currently using this microbial biocontrol technology to effectively treat disease in spice crops. The powerful methodology of NGS-based polymer barcoding, which solely targets short regions of genomic polymer and does not need full genome-scale information, will facilitate breeders in quicker identification and classification of untamed populations of these plants. Plant tissue culture could be a core part of BBBMs, which helps in the conservation and micropropagation of spice plants. In in vitro condition, induction and bioreactors are the two alternative tissue culture-derived methods that have led to prominent improvement of spices plants. Agrobacterium-mediated transformation (A. tumefaciens and A. rhizogenes) is another technique that has led to the wide-ranging improvement of spices by overexpressing the key genes which respond to an organic phenomenon or abiotic stress and special substance pathway. Genome editing approaches (TALLENs, ZFNs, CRISPR/Cas9) are other alternative ways to improve spices plants. Among all genome editing methods, CRISPR/Cas9 has emerged as a promising approach to induce targeted mutation within spice plant genomes, ultimately purposefully altering their organic chemistry profile. Thus, biotechnology-based techniques are definitely the way forward for the overall development of spice plants.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The authors declared that there is no conflict of interest.

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LIST OF ABBREVIATIONS

NGS	Next-generation sequencing techniques	
RAD-seq	Restriction-site-Associated DNA sequencing	
TALENs	Transcription Activator-Like Effector Nucleases	
FDA	Food and Drug Administration association	
KEGG	Kyoto Encyclopedia of Genes and Genomes	
DEGs	Differentially Expressed Genes	
PEG	Polyethylene glycol	
RAPD	Random amplification of polymorphic DNA	
MS medium	Murashige and Skoog medium	
BBBMs	Biotechnology based breeding methods	
RFLP	Restriction fragment length polymorphism	
SSRs	Simple-sequence repeats	
AFLP	Amplified Fragment Length Polymorphism	
SNP	Single nucleotide polymorphisms	
SCAR	Species-specific sequence-characterized amplified region	
EST-SSRs	Expressed sequence tag-derived simple sequence repeat markers	
GWAS	Genome-wide association studies	
CAPS	Cleaved Amplified Polymorphic Sequences	
CRISPR	Short palindromic repeats	
ISR	Induced systemic resistance	

REFERENCES

- [1] Douglas M, Heyes J, Smallfield B. Herbs, spices and essential oils: post-harvest operations in developing countries. UNIDO and FAO 2005; p. 61.
- [2] Peter K. Handbook of Herbs and Spices. Woodhead publishing 2006; Vol. 3. [http://dx.doi.org/10.1533/9781845691717]
- [3] Radulović NS, Blagojević PD, Stojanović-Radić ZZ, Stojanović NM. Antimicrobial plant metabolites: structural diversity and mechanism of action. Curr Med Chem 2013; 20(7): 932-52.
 [PMID: 23210781]

- [4] Opara EI, Chohan M. Culinary herbs and spices: their bioactive properties, the contribution of polyphenols and the challenges in deducing their true health benefits. Int J Mol Sci 2014; 15(10): 19183-202.
 [http://dx.doi.org/10.3390/ijms151019183] [PMID: 25340982]
- [5] Kaefer CM, Milner JA. The role of herbs and spices in cancer prevention. J Nutr Biochem 2008; 19(6): 347-61.

[http://dx.doi.org/10.1016/j.jnutbio.2007.11.003] [PMID: 18499033]

- [6] Vázquez-Fresno R, Rosana ARR, Sajed T, Onookome-Okome T, Wishart NA, Wishart DS. Herbs and spices-biomarkers of intake based on human intervention studies–a systematic review. Genes Nutr 2019; 14(1): 18. [http://dx.doi.org/10.1186/s12263-019-0636-8] [PMID: 31143299]
- [7] El-Sayed SM, Youssef AM. Potential application of herbs and spices and their effects in functional dairy products. Heliyon 2019; 5(6): e01989.
 [http://dx.doi.org/10.1016/j.heliyon.2019.e01989] [PMID: 31338458]
- [8] Bhatt N. Herbs and herbal supplements, a novel nutritional approach in animal nutrition 2015.
- [9] Piquerez SJ, Harvey SE, Beynon JL, Ntoukakis V. Improving crop disease resistance: lessons from research on Arabidopsis and tomato. Front Plant Sci 2014; 5: 671.
 [http://dx.doi.org/10.3389/fpls.2014.00671] [PMID: 25520730]
- Hao C, Xia Z, Fan R, *et al.* De novo transcriptome sequencing of black pepper (*Piper nigrum* L.) and an analysis of genes involved in phenylpropanoid metabolism in response to *Phytophthora capsici*. BMC Genomics 2016; 17(1): 822.
 [http://dx.doi.org/10.1186/s12864-016-3155-7] [PMID: 27769171]
- [11] Gordo SM, Pinheiro DG, Moreira EC, et al. High-throughput sequencing of black pepper root transcriptome. BMC Plant Biol 2012; 12(1): 168.
 [http://dx.doi.org/10.1186/1471-2229-12-168] [PMID: 22984782]
- [12] Albenne C, Canut H, Jamet E. Plant cell wall proteomics: the leadership of Arabidopsis thaliana. Front Plant Sci 2013; 4: 111. [http://dx.doi.org/10.3389/fpls.2013.00111] [PMID: 23641247]
- [13] Hu L, Hao C, Fan R, Wu B, Tan L, Wu H. De novo assembly and characterization of fruit transcriptome in black pepper (*Piper nigrum*). PLoS One 2015; 10(6): e0129822. [http://dx.doi.org/10.1371/journal.pone.0129822] [PMID: 26121657]
- [14] Karthika R, Prasath D, Anandaraj M. Transcriptome-wide identification and characterization of resistant gene analogs (RGAs) of ginger (*Zingiber officinale* Rosc.) and mango ginger (*Curcuma amada* Roxb.) under stress induced by pathogen. Sci Hortic (Amsterdam) 2019; 248: 81-8. [http://dx.doi.org/10.1016/j.scienta.2019.01.003]
- [15] Gaur M, Das A, Sahoo RK, Mohanty S, Joshi RK, Subudhi E. Comparative transcriptome analysis of ginger variety Suprabha from two different agro-climatic zones of Odisha. Genom Data 2016; 9: 42-3. [http://dx.doi.org/10.1016/j.gdata.2016.06.014] [PMID: 27408809]
- [16] Annadurai RS, Neethiraj R, Jayakumar V, *et al.* De Novo transcriptome assembly (NGS) of *Curcuma longa* L. rhizome reveals novel transcripts related to anticancer and antimalarial terpenoids. PLoS One 2013; 8(2): e56217.
 [http://dx.doi.org/10.1371/journal.pone.0056217] [PMID: 23468859]
- [17] Sheeja T, Deepa K, Santhi R, Sasikumar B. Comparative transcriptome analysis of two species of Curcuma contrasting in a high-value compound curcumin: insights into genetic basis and regulation of biosynthesis. Plant Mol Biol Report 2015; 33(6): 1825-36. [http://dx.doi.org/10.1007/s11105-015-0878-6]
- [18] Babu KN, Jose C, Suraby EJ, Peter KV. Chapter 16-Transgenic Research in Spices. In: Rout GR, Peter KV (Eds) Genetic Engineering of Horticultural Crops. Elsevier 2018; 387-412.

[http://dx.doi.org/10.1016/B978-0-12-810439-2.00016-7]

- [19] Tulsani NJ, Hamid R, Jacob F, et al. Transcriptome landscaping for gene mining and SSR marker development in Coriander (*Coriandrum sativum* L.). Genomics 2020; 112(2): 1545-53. [http://dx.doi.org/10.1016/j.ygeno.2019.09.004] [PMID: 31505244]
- [20] Paul BB, Mathew D, Beena S, Shylaja MR. Comparative transcriptome analysis reveals the signal proteins and defence genes conferring foot rot (*Phytophthora capsici* sp. nov.) resistance in black pepper (*Piper nigrum* L.). Physiol Mol Plant Pathol 2019; 108: 101436. [http://dx.doi.org/10.1016/j.pmpp.2019.101436]
- [21] Jiang Y, Liao Q, Zou Y, Liu Y, Lan J. Transcriptome analysis reveals the genetic basis underlying the biosynthesis of volatile oil, gingerols, and diarylheptanoids in ginger (*Zingiber officinale* Rosc.). Bot Stud (Taipei, Taiwan) 2017; 58(1): 41. [http://dx.doi.org/10.1186/s40529-017-0195-5] [PMID: 29058093]
- [22] Roorkiwal M, Nayak SN, Thudi M, *et al.* Allele diversity for abiotic stress responsive candidate genes in chickpea reference set using gene based SNP markers. Front Plant Sci 2014; 5: 248. [http://dx.doi.org/10.3389/fpls.2014.00248] [PMID: 24926299]
- [23] Parmar N, Singh KH, Sharma D, *et al.* Genetic engineering strategies for biotic and abiotic stress tolerance and quality enhancement in horticultural crops: a comprehensive review. 3 Biotech 2017; 7(4): 239.
- [24] Babu N, et al. Status of transgenics in Indian spices. J Trop Agric 2013; 51(1): 1-14.
- [25] Min J, Shin SH, Jeon EM, Park JM, Hyun JY, Harn CH. Pepper, chili (*Capsicum annuum*). Agrobacterium protocols. Springer 2015; pp. 311-20. [http://dx.doi.org/10.1007/978-1-4939-1695-5 25]
- [26] Hood EE, Gelvin SB, Melchers LS, Hoekema A. NewAgrobacterium helper plasmids for gene transfer to plants. Transgenic Res 1993; 2(4): 208-18. [http://dx.doi.org/10.1007/BF01977351]
- [27] Keshavareddy G, Kumar A, Ramu VS. Methods of Plant Transformation-A Review. Int J Curr Microbiol Appl Sci 2018; 7(7): 2656-68. [http://dx.doi.org/10.20546/ijcmas.2018.707.312]
- [28] Jogdand S. Gene biotechnology 2009.
- [29] Liu H, Kawabe A, Matsunaga S, *et al.* Obtaining transgenic plants using the bio-active beads method. J Plant Res 2004; 117(2): 95-9.
 [http://dx.doi.org/10.1007/s10265-003-0141-3] [PMID: 15108034]
- [30] Divakaran M, Pillal GS, Babu KN, Peter KV. Isolation and fusion of protoplasts in Vanilla species. Curr Sci 2008; 115-20.
- [31] Guan Q, et al. Regeneration of somatic hybrids of ginger via chemical protoplast fusion. Plant Cell Tissue Organ Cult 2010; 102(3): 279-84. [PCTOC]. [http://dx.doi.org/10.1007/s11240-010-9730-8]
- [32] Stephen R, Jayabalan N. Artificial seed production in coriander (*Coriandrum sativum* L.). Plant Tissue Cult 2000; 10(1): 45-9.
- [33] Khan K. Gene transfer technologies in plants: Roles in improving crops. Recent Research in Science and Technology 2009.
- [34] Morikawa H, Yamada Y. Capillary microinjection into protoplasts and intranuclear localization of injected materials. Plant Cell Physiol 1985; 26(2): 229-36.
- [35] Keshavareddy G, Kumar ARV, Ramu VS. Methods of Plant Transformation- A Review. Int J Curr Microbiol Appl Sci 2018; 7(07): 2656-68. [http://dx.doi.org/10.20546/ijcmas.2018.707.312]

- [36] Nair KP. The Biotechnology of Ginger. Turmeric (*Curcuma longa* L.) and Ginger (*Zingiber officinale* Rosc.)-World's Invaluable Medicinal Spices. Springer 2019; pp. 405-32. [http://dx.doi.org/10.1007/978-3-030-29189-1_19]
- [37] Shirgurkar MV, Naik VB, von Arnold S, Nadgauda RS, Clapham D. An efficient protocol for genetic transformation and shoot regeneration of turmeric (*Curcuma longa L.*) via particle bombardment. Plant Cell Rep 2006; 25(2): 112-6. [http://dx.doi.org/10.1007/s00299-005-0033-1] [PMID: 16397786]
- [38] Singh N, et al. Microprojectile bombardment mediated genetic transformation of embryo axes and plant regeneration in cumin (*Cuminum cyminum* L.). Plant Cell Tissue Organ Cult 2010; 103(1): 1-6. [PCTOC]. [http://dx.doi.org/10.1007/s11240-010-9746-0]
- [39] Ismagul A, Yang N, Maltseva E, *et al.* A biolistic method for high-throughput production of transgenic wheat plants with single gene insertions. BMC Plant Biol 2018; 18(1): 135. [http://dx.doi.org/10.1186/s12870-018-1326-1] [PMID: 29940859]
- [40] Guo M, Ye J, Gao D, Xu N, Yang J. Agrobacterium-mediated horizontal gene transfer: Mechanism, biotechnological application, potential risk and forestalling strategy. Biotechnol Adv 2019; 37(1): 259-70.

[http://dx.doi.org/10.1016/j.biotechadv.2018.12.008] [PMID: 30579929]

- [41] Bahramnejad B, Naji M, Bose R, Jha S. A critical review on use of Agrobacterium rhizogenes and their associated binary vectors for plant transformation. Biotechnol Adv 2019; 37(7): 107405. [http://dx.doi.org/10.1016/j.biotechadv.2019.06.004] [PMID: 31185263]
- [42] Suraby E, *et al.* Expression analysis of resistance gene analogs in *Piper colubrinum-Phytophthora capsici* pathosystem. International Journal of Innovative Horticulture 2015; 4(2): 107-12.
- [43] Varghese JM, Bhat AI. An efficient Agrobacterium-mediated transformation protocol for black pepper (*Piper nigrum* L.) using embryogenic mass as explant. J Crop Sci Biotechnol 2011; 14(4): 247-54. [http://dx.doi.org/10.1007/s12892-011-0031-5]
- [44] Sinoj J, *et al.* Somatic embryogenesis and transgenic development in black pepper for delayed infection and decreased spread of foot rot caused by *Phytophthora capsici.* J Plant Crops 2014; 20-8.
- [45] Hu L, Xu Z, Wang M, et al. The chromosome-scale reference genome of black pepper provides insight into piperine biosynthesis. Nat Commun 2019; 10(1): 4702. [http://dx.doi.org/10.1038/s41467-019-12607-6] [PMID: 31619678]
- [46] Lee YH, Jung M, Shin SH, *et al.* Transgenic peppers that are highly tolerant to a new CMV pathotype. Plant Cell Rep 2009; 28(2): 223-32.
 [http://dx.doi.org/10.1007/s00299-008-0637-3] [PMID: 19018536]
- [47] Choi B, Kwon SJ, Kim MH, et al. A plant virus-based vector system for gene function studies in pepper. Plant Physiol 2019; 181(3): 867-80.
 [http://dx.doi.org/10.1104/pp.19.00836] [PMID: 31481630]
- [48] Lee YH, Kim HS, Kim JY, et al. A new selection method for pepper transformation: callus-mediated shoot formation. Plant Cell Rep 2004; 23(1-2): 50-8. [http://dx.doi.org/10.1007/s00299-004-0791-1] [PMID: 15221276]
- [49] Mahto BK, et al. An efficient method for Agrobacterium-mediated genetic transformation of chilli pepper (Capsicum annuum L.). Indian J Plant Physiol 2018; 23(3): 573-81. [http://dx.doi.org/10.1007/s40502-018-0389-1]
- [50] Mehaboob VM, et al. Direct organogenesis and microrhizome production in ginger (Zingiber officinale Rosc.). J Pharmacogn Phytochem 2019; 8(3): 2880-3.
- [51] Babu KN. Biotechnology in conservation and development of spices. Souvenir Abstr SYMSAC 2011; VI: 79-93.

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- [52] He R, Gang DR. Somatic embryogenesis and Agrobacterium-mediated transformation of turmeric (*Curcuma longa*). Plant Cell Tissue Organ Cult 2014; 116(3): 333-42. [PCTOC]. [http://dx.doi.org/10.1007/s11240-013-0407-y]
- [53] Brito-Sánchez SC, et al. Biolistics transformation of callus and cell suspension cultures of Capsicum annuum L. 'Serrano'is useful for in vitro studies of the relative contents of secondary metabolites. Acta Agrobot 2019; 72(4) [http://dx.doi.org/10.5586/aa.1792]
- [54] Ochoa-Alejo N, Ramirez-Malagon R. *In vitro* chili pepper biotechnology. *In Vitro* Cell Dev Biol Plant 2001; 37(6): 701-29.
 [http://dx.doi.org/10.1007/s11627-001-0121-z]
- [55] Kumar RV, Sharma VK, Chattopadhyay B, Chakraborty S. An improved plant regeneration and Agrobacterium - mediated transformation of red pepper (*Capsicum annuum* L.). Physiol Mol Biol Plants 2012; 18(4): 357-64. [http://dx.doi.org/10.1007/s12298-012-0132-8] [PMID: 24082498]
- [56] Joshi RK, Mohanty S, Subudhi E, Nayak S. Isolation and characterization of NBS-LRR- resistance gene candidates in turmeric (*Curcuma longa* cv. surama). Genet Mol Res 2010; 9(3): 1796-806. [http://dx.doi.org/10.4238/vol9-3gmr910] [PMID: 20830672]
- [57] Cai W-Q, et al. Development of CMV-and TMV-resistant chili pepper: field perfermance and biosafety assessment. Mol Breed 2003; 11(1): 25-35. [http://dx.doi.org/10.1023/A:1022655204552]
- [58] Tuhaise S, Nakavuma JL, Adriko J, Ssekatawa K, Kiggundu A. *In vitro* regeneration of Ugandan passion fruit cultivars from leaf discs. BMC Res Notes 2019; 12(1): 425. [http://dx.doi.org/10.1186/s13104-019-4469-8] [PMID: 31311592]
- [59] Musfir Mehaboob V, et al. Indirect somatic embryogenesis and Agrobacterium-mediated transient transformation of ginger (*Zingiber officinale* Rosc.) using leaf sheath explants. J Hortic Sci Biotechnol 2019; 94(6): 753-60. [http://dx.doi.org/10.1080/14620316.2019.1624201]
- [60] Phillips GC, Garda M. Plant tissue culture media and practices: an overview. *In vitro* Cell Dev Biol Plant 2019; 55(3): 242-57. [http://dx.doi.org/10.1007/s11627-019-09983-5]
- [61] Prathanturarug S, Soonthornchareonnon N, Chuakul W, Phaidee Y, Saralamp P. High-frequency shoot multiplication in *Curcuma longa* L using thidiazuron. Plant Cell Rep 2003; 21(11): 1054-9. [http://dx.doi.org/10.1007/s00299-003-0629-2] [PMID: 12835998]
- [62] Keller EJ, Senula A. Micropropagation and cryopreservation of garlic (*Allium sativum* L.). Protocols for micropropagation of selected economically-important horticultural plants. Springer 2012; pp. 353-68.
 [http://dx.doi.org/10.1007/978-1-62703-074-8 28]

- [63] Valdiani A, Hansen OK, Nielsen UB, et al. Bioreactor-based advances in plant tissue and cell culture: challenges and prospects. Crit Rev Biotechnol 2018; 39(1): 1-15. [PMID: 30431379]
- Yasuda K, Tsuda T, Shimizu H, Sugaya A. Multiplication of curcuma species by tissue culture. Planta Med 1988; 54(1): 75-9.
 [http://dx.doi.org/10.1055/s-2006-962344] [PMID: 17265210]
- [65] Sultana A, et al. In vitro regeneration of ginger using leaf, shoot tip and root explants. Pak J Bot 2009; 41(4): 1667-76.
- [66] Hashemabadi D, Kaviani B. Rapid micro-propagation of Aloe vera L. via shoot multiplication. Afr J Biotechnol 2008; 7(12)

- [67] Garro-Monge G, Gatica-Arias AM, Valdez-Melara M. Somatic embryogenesis, plant regeneration and acemannan detection in aloe (Aloe barbadensis Mill.). Agron Costarric 2008.
- [68] Nazeem P, et al. A viable protocol for large scale in vitro multiplication of black pepper (P. nigrum L.). J Plant Crops 2004; 32: 163-8.
- [69] Kaliyadasa E, Samarasinghe BA. A review on golden species of Zingiberaceae family around the world: Genus Curcuma. Afr J Agric Res 2019; 14(9): 519-31. [http://dx.doi.org/10.5897/AJAR2018.13755]
- [70] Sreeja K, Anandaraj M, Bhai R. Colonization and plant growth promotion in somatic embryo derived black pepper plants by fungal endophytes. Journal of Global Biosciences 2019; 8(11): 6525-39.
- [71] Jie EY, et al. Establishment of a high-frequency plant regeneration system from rhizome-derived embryogenic cell-suspension cultures of *Curcuma longa* L. Plant Biotechnol Rep 2019; 13(2): 123-9. [http://dx.doi.org/10.1007/s11816-019-00519-2]
- [72] Raju CS, et al. An efficient regeneration system via somatic embryogenesis in mango ginger (Curcuma amada Roxb.). Plant Cell Tissue Organ Cult 2013; 112(3): 387-93. [PCTOC]. [http://dx.doi.org/10.1007/s11240-012-0244-4]
- [73] Raju CS, Aslam A, Shajahan A. Germination and storability of calcium-alginate coated somatic embryos of mango ginger (*Curcuma amada* Roxb.). Hortic Environ Biotechnol 2016; 57(1): 88-96. [http://dx.doi.org/10.1007/s13580-016-0096-7]
- [74] Guo Y, Zhang Z. Establishment and plant regeneration of somatic embryogenic cell suspension cultures of the *Zingiber officinale* Rosc. Sci Hortic (Amsterdam) 2005; 107(1): 90-6. [http://dx.doi.org/10.1016/j.scienta.2005.07.003]
- [75] Nair RR, Dutta Gupta S. High-frequency plant regeneration through cyclic secondary somatic embryogenesis in black pepper (*Piper nigrum* L.). Plant Cell Rep 2006; 24(12): 699-707. [http://dx.doi.org/10.1007/s00299-005-0016-2] [PMID: 16249871]
- [76] Koleva-Gudeva LR, Spasenoski M, Trajkova F. Somatic embryogenesis in pepper anther culture: The effect of incubation treatments and different media. Sci Hortic (Amsterdam) 2007; 111(2): 114-9. [http://dx.doi.org/10.1016/j.scienta.2006.10.013]
- [77] You XL, et al. Large-scale somatic embryogenesis and regeneration of Panax notoginseng. Plant Cell Tissue Organ Cult 2012; 108(2): 333-8. [PCTOC]. [http://dx.doi.org/10.1007/s11240-011-0030-8]
- [78] Arab MM, et al. Modeling and optimizing a new culture medium for in vitro rooting of G× N15 Prunus rootstock using artificial neural network-genetic algorithm. Sci Rep 2018; 8(1): 1-18. [http://dx.doi.org/10.1038/s41598-018-27858-4] [PMID: 29311619]
- [79] Aka Kaçar Y, et al. Evaluation and Comparison of A New Type of Temporary Immersion System (TIS) Bioreactors for Myrtle (*Myrtus communis* L.). Appl Ecol Environ Res 2020; 18(1): 1611-20. [http://dx.doi.org/10.15666/aeer/1801 16111620]
- [80] Ramos-Castellá A, et al. Improved propagation of vanilla (Vanilla planifolia Jacks. ex Andrews) using a temporary immersion system. In vitro Cell Dev Biol Plant 2014; 50(5): 576-81. [http://dx.doi.org/10.1007/s11627-014-9602-8]
- [81] Zapata EV, et al. In vitro regeneration and acclimatization of plants of turmeric (Curcuma longa L.) in a hydroponic system. Biotecnol Apl 2003; 20: 25-31.
- [82] Tyagi RK, et al. Low-cost media for in vitro conservation of turmeric (Curcuma longa L.) and genetic stability assessment using RAPD markers. In Vitro Cell Dev Biol Plant 2007; 43(1): 51-8. [http://dx.doi.org/10.1007/s11627-006-9000-y]
- [83] Tyagi RK, et al. Micropropagation and slow growth conservation of cardamom (Elettaria cardamomum Maton). In Vitro Cell Dev Biol Plant 2009; 45(6): 721-9. [http://dx.doi.org/10.1007/s11627-009-9234-6]

Spices Biotechnology

- [84] Jain A, et al. Effect of carbendazim on in vitro conservation and genetic stability assessment in Curcuma longa. and Zingiber officinale. J Herbs Spices Med Plants 2018; 24(2): 160-72. [http://dx.doi.org/10.1080/10496475.2017.1423528]
- [85] Yang L, Wu Y, Zhang M, et al. Transcriptome, cytological and biochemical analysis of cytoplasmic male sterility and maintainer line in CMS-D8 cotton. Plant Mol Biol 2018; 97(6): 537-51. [http://dx.doi.org/10.1007/s11103-018-0757-2] [PMID: 30066309]
- [86] Balachandran SM, Bhat SR, Chandel KP. *In vitro* clonal multiplication of turmeric (Curcuma spp.) and ginger (*Zingiber officinale* Rosc.). Plant Cell Rep 1990; 8(9): 521-4. [http://dx.doi.org/10.1007/BF00820200] [PMID: 24226277]
- [87] Dekkers A, Rao A, Goh C. *In vitro* storage of multiple shoot cultures of gingers at ambient temperatures of 24-29 degrees C. Netherlands: Scientia Horticulturae 1991.
- [88] Nair KP. Turmeric (*Curcuma longa* L.) and Ginger (*Zingiber officinale* Rosc.)-World's Invaluable Medicinal Spices: The Agronomy and Economy of Turmeric and Ginger.. Springer Nature 2019. [http://dx.doi.org/10.1007/978-3-030-29189-1]
- [89] Wang M-R, *et al.* Advances in cryopreservation of *in vitro*-derived propagules: technologies and explant sources. Plant Cell Tissue Organ Cult 2020; 1-14. [PCTOC].
- [90] Ruta C, et al. Long-term preservation of Cicer arietinum L. germplasm by in vitro propagation and cryopreservation. Genet Resour Crop Evol 2020; 67(2): 263-71. [http://dx.doi.org/10.1007/s10722-019-00867-6]
- [91] Takagi H. Recent developments in cryopreservation of shoot apices of tropical species. 2000.
- [92] Taesan K, et al. Implementation of cryopreservation for garlic genetic resources by the droplet vitrification procedure. in XXVII International Horticultural Congress-IHC2006: II International Symposium on Plant Genetic Resources of Horticultural 760 2006.
- [93] Kim H-H, Cho EG, Baek HJ, Kim CY, Joachim Keller ER, Engelmann F. Cryopreservation of garlic shoot tips by vitrification: effects of dehydration, rewarming, unloading and regrowth conditions. Cryo Lett 2004; 25(1): 59-70.
 [PMID: 15031746]
- [94] Keller ER. Improvement of cryopreservation results in garlic using low temperature preculture and high-quality *in vitro* plantlets. Cryo Lett 2005; 26(6): 357-66.[PMID: 16547550]
- [95] Peter K, et al. Establishing in vitro conservatory of spices germplasm. ICAR Project Report.. Kerala: Indian Institute of Spices Research 2002.
- [96] Chaudhury R, Malik S. Conservation of Spices and Tree Borne Oil Seed Crops. Conservation of Tropical Plant Species. Springer 2013; pp. 419-36. [http://dx.doi.org/10.1007/978-1-4614-3776-5 17]
- [97] Yamuna G, Sumathi V, Geetha SP, Praveen K, Swapna N, Babu KN. Cryopreservation of *in vitro* grown shoots of ginger (*Zingiber officinale* Rosc.). Cryo Lett 2007; 28(4): 241-52. [PMID: 17962828]
- [98] Babu KN, et al. Cryopreservation of spices genetic resources. Current frontiers in cryobiology 2012; 457-84.
- [99] Chaudhury R, Chandel K. Studies on germination and cryopreservation of cardamom (*Elettaria cardamomum* Maton) seeds.. Switzerland: Seed Science and Technology 1995.
- [100] Agrawal A, et al. In vitro conservation and cryopreservation of clonally propagated horticultural species. Conservation and Utilization of Horticultural Genetic Resources. Springer 2019; pp. 529-78. [http://dx.doi.org/10.1007/978-981-13-3669-0 18]
- [101] Dolce NR, Hernández-Ramírez F, González-Arnao MT. Cryopreservation of vanilla (Vanilla

planifolia) root-tips: a new alternative forin vitro long-term storage of its germplasm. III International Symposium on Plant Cryopreservation 1234.

- [102] Popova E, Kim H-H, Paek K-Y. Cryopreservation of coriander (*Coriandrum sativum* L.) somatic embryos using sucrose preculture and air desiccation. Sci Hortic (Amsterdam) 2010; 124(4): 522-8. [http://dx.doi.org/10.1016/j.scienta.2010.02.012]
- [103] González-Benito ME, Iriondo JM. Cryopreservation of *Apium graveolens* L.(celery) seeds. Cryopreservation of Plant Germplasm II. Springer 2002; pp. 48-56. [http://dx.doi.org/10.1007/978-3-662-04674-6 4]
- [104] Carvalho AAd, et al. Growth and volatiles in the micropropagation of Santa Maria herb. Rev Cienc Agron 2018; 49(4): 624-35. [http://dx.doi.org/10.5935/1806-6690.20180071]
- [105] Algabri AA. INTERNATIONAL JOURNAL OF SCIENTIFIC RESEARCH.
- [106] Nandini B, Giridhar P. Insight View of Topical Trends on Synthetic Seeds of Rare and Endangered Plant Species and Its Future Prospects 2019. [http://dx.doi.org/10.1007/978-3-030-24631-0 5]
- [107] Bielak-Zmijewska A, Grabowska W, Ciolko A, et al. The role of curcumin in the modulation of ageing. Int J Mol Sci 2019; 20(5): 1239. [http://dx.doi.org/10.3390/ijms20051239] [PMID: 30871021]
- [108] Chen L-L, Gao S-L. In vitro tetraploid induction and generation of tetraploids from mixoploids in Astragalus membranaceus. Sci Hortic (Amsterdam) 2007; 112(3): 339-44. [http://dx.doi.org/10.1016/j.scienta.2006.12.045]
- [109] De Jesus L. Effects of artificial polyploidy in transformed roots of Artemisia annua L. 2003.
- [110] Gu XF, Yang AF, Meng H, Zhang JR. In vitro induction of tetraploid plants from diploid Zizyphus jujuba Mill. cv. Zhanhua. Plant Cell Rep 2005; 24(11): 671-6. [http://dx.doi.org/10.1007/s00299-005-0017-1] [PMID: 16094528]
- [111] Mahmodi R, et al. Antioxidant and antibacterial properties of the Melissa officinalis essential oil. the journal of qazvin university of medical sciences 2016; 20(2): 49-57.
- [112] Talei D, et al. Improving productivity of steviol glycosides in Stevia rebaudiana via induced polyploidy. J Crop Sci Biotechnol 2020; 1-9. [http://dx.doi.org/10.1007/s12892-020-00038-5]
- [113] Peraza-Luna F, Rodríguez-Mendiola M, Arias-Castro C, Bessiere JM, Calva-Calva G. Sotolone production by hairy root cultures of *Trigonella foenum*-graecum in airlift with mesh bioreactors. J Agric Food Chem 2001; 49(12): 6012-9. [http://dx.doi.org/10.1021/jf010818j] [PMID: 11743801]
- [114] Sheludko Y, Gerasimenko I, Kolshorn H, Stöckigt J. New alkaloids of the sarpagine group from Rauvolfia serpentina hairy root culture. J Nat Prod 2002; 65(7): 1006-10. [http://dx.doi.org/10.1021/np0200919] [PMID: 12141861]
- [115] Madhusudanan KP, Banerjee S, Khanuja SP, Chattopadhyay SK. Analysis of hairy root culture of Rauvolfia serpentina using direct analysis in real time mass spectrometric technique. Biomed Chromatogr 2008; 22(6): 596-600. [http://dx.doi.org/10.1002/bmc.974] [PMID: 18205139]
- [116] Santos PA, et al. Hairy root cultures of Anethum graveolens (dill): establishment, growth, time-course study of their essential oil and its comparison with parent plant oils. Biotechnol Lett 2002; 24(12): 1031-6.
 [http://dx.doi.org/10.1023/A:1015653701265]
- [117] Babu KN, et al. Biotechnological approaches in improvement of spices: a review. Plant Biology and Biotechnology. Springer 2015; pp. 487-516.

[http://dx.doi.org/10.1007/978-81-322-2283-5_25]

- [118] Hamid R, *et al.* Evaluation the genetic diversity of 10 Milk thistle (*Silybum marianum* L.) ecotypes using morphological, phenological and phytochemical traits. 2014.
- [119] Babu KN, et al. Protocols for improvement of black pepper (*Piper nigrum* L.) utilizing biotechnological tools. Protocols forin vitro Cultures and Secondary Metabolite Analysis of Aromatic and Medicinal Plants. 2nd ed. Springer 2016; pp. 367-85.
- [120] Hamid R, Jacob F, Marashi H, Rathod V, Tomar RS. Uncloaking lncRNA-meditated gene expression as a potential regulator of CMS in cotton (*Gossypium hirsutum* L.). Genomics 2020; 112(5): 3354-64. [http://dx.doi.org/10.1016/j.ygeno.2020.06.027] [PMID: 32574832]
- [121] Morgil H, Gercek YC, Tulum I. Single Nucleotide Polymorphisms (SNPs) in Plant Genetics and Breeding, in The Recent Topics in Genetic Polymorphisms. IntechOpen 2020.
- [122] Torkamaneh D, Laroche J, Belzile F. Genome-wide SNP calling from genotyping by sequencing (GBS) data: a comparison of seven pipelines and two sequencing technologies. PLoS One 2016; 11(8): e0161333.
 [http://dx.doi.org/10.1371/journal.pone.0161333] [PMID: 27547936]
- [123] Wickland DP, Battu G, Hudson KA, Diers BW, Hudson ME. A comparison of genotyping-b--sequencing analysis methods on low-coverage crop datasets shows advantages of a new workflow, GB-eaSy. BMC Bioinformatics 2017; 18(1): 586. [http://dx.doi.org/10.1186/s12859-017-2000-6] [PMID: 29281959]
- [124] Siadjeu C, Mayland-Quellhorst E, Albach DC. Genetic diversity and population structure of trifoliate yam (Dioscorea dumetorum Kunth) in Cameroon revealed by genotyping-by-sequencing (GBS). BMC Plant Biol 2018; 18(1): 359.
 [http://dx.doi.org/10.1186/s12870-018-1593-x] [PMID: 30563456]
- [125] Kumar S, Banks TW, Cloutier S. SNP discovery through next-generation sequencing and its applications. International journal of plant genomics 2012. [http://dx.doi.org/10.1155/2012/831460]
- [126] Baral K, Coulman B, Biligetu B, Fu YB. Genotyping-by-sequencing enhances genetic diversity analysis of crested wheatgrass. Int J Mol Sci 2018; 19(9): 2587. [Agropyron cristatum (L.) Gaertn.]. [http://dx.doi.org/10.3390/ijms19092587] [PMID: 30200310]
- [127] Schreiber M, Stein N, Mascher M. Genomic approaches for studying crop evolution. Genome Biol 2018; 19(1): 140.
 [http://dx.doi.org/10.1186/s13059-018-1528-8] [PMID: 30241487]
- [128] Zhang X, Pérez-Rodríguez P, Semagn K, et al. Genomic prediction in biparental tropical maize populations in water-stressed and well-watered environments using low-density and GBS SNPs. Heredity 2015; 114(3): 291-9. [http://dx.doi.org/10.1038/hdy.2014.99] [PMID: 25407079]
- [129] Liu H, Bayer M, Druka A, et al. An evaluation of genotyping by sequencing (GBS) to map the Breviaristatum-e (ari-e) locus in cultivated barley. BMC Genomics 2014; 15(1): 104. [http://dx.doi.org/10.1186/1471-2164-15-104] [PMID: 24498911]
- [130] Alipour H, Bai G, Zhang G, Bihamta MR, Mohammadi V, Peyghambari SA. Imputation accuracy of wheat genotyping-by-sequencing (GBS) data using barley and wheat genome references. PLoS One 2019; 14(1): e0208614. [http://dx.doi.org/10.1371/journal.pone.0208614] [PMID: 30615624]
- [131] Carbonell P, et al. Twenty years of tomato breeding at EPSO-UMH: Transfer resistance from wild types to local landraces—from the first molecular markers to genotyping by sequencing (GBS). Diversity (Basel) 2018; 10(1): 12. [http://dx.doi.org/10.3390/d10010012]
- [132] Hu Z, Olatoye MO, Marla S, Morris GP. An Integrated Genotyping-by-Sequencing Polymorphism

Map for Over 10,000 Sorghum Genotypes. Plant Genome 2019; 12(1): 1-15. [http://dx.doi.org/10.3835/plantgenome2018.06.0044] [PMID: 30951089]

- [133] Lemay M-A, Torkamaneh D, Rigaill G, *et al.* Screening populations for copy number variation using genotyping-by-sequencing: a proof of concept using soybean fast neutron mutants. BMC Genomics 2019; 20(1): 634.
 [http://dx.doi.org/10.1186/s12864-019-5998-1] [PMID: 31387530]
- [134] Pereira-Dias L, Vilanova S, Fita A, Prohens J, Rodríguez-Burruezo A. Genetic diversity, population structure, and relationships in a collection of pepper (*Capsicum* spp.) landraces from the Spanish centre of diversity revealed by genotyping-by-sequencing (GBS). Hortic Res 2019; 6(1): 54. [http://dx.doi.org/10.1038/s41438-019-0132-8] [PMID: 31044080]
- [135] George KJ, et al. Identification of hybrids in black pepper (*Piper nigrum* L.) using male parent-specific RAPD markers. Curr Sci 2005; 88(2): 216-8.
- [136] Nazeem P, et al. Assessment of genetic variability in Black Pepper (*Piper nigrum* L.) varieties through RAPD and AFLP analyses. Recent trends in horticultural biotechnology 2007; 485-90.
- [137] Remmia R, et al. Molecular characterization of black pepper (*Piper nigrum*) using RAPD and SSR markers. Biosci Biotechnol Res Asia 2010; 7(2): 1011-5.
- [138] Khan S, et al. Development of RAPD markers for authentication of Piper nigrum (L.). Environ We Int J Sci Tech 2010; 5: 47-56.
- [139] Singh S, Panda MK, Nayak S. Evaluation of genetic diversity in turmeric (*Curcuma longa* L.) using RAPD and ISSR markers. Ind Crops Prod 2012; 37(1): 284-91. [http://dx.doi.org/10.1016/j.indcrop.2011.12.022]
- [140] Jan HU, Rabbani MA, Shinwari ZK. Assessment of genetic diversity of indigenous turmeric (*Curcuma longa L.*) germplasm from Pakistan using RAPD markers. J Med Plants Res 2011; 5(5): 823-30.
- [141] Pareek N, Jakhar M, Malik C. Analysis of genetic diversity in coriander (*Coriandrum sativum* L.) varieties using random amplified polymorphic DNA (RAPD) markers. J Microbiol Biotechnol Res 2011; 1(4): 206-15.
- [142] Singh R, et al. Characterization of coriander (Coriandrum sativum L.) varieties using SDS-PAGE and RAPD markers. Afr J Biotechnol 2013; 12(11): 1189-95.
- [143] Babu KN, et al. Genetic diversity and phylogenetic relationship among small cardamom (*Elettaria cardamomum* Maton.) cultivars and related genera using DNA markers. International Journal of Innovative Horticulture 2012; 1(1): 47-56.
- [144] PHADNIS S, PETER A. Morphological Studies and Genetic Diversity Analysis of Cardamom (*Elettaria cardamomum* Maton.) Genotypes and Hedychium coronarium Using RAPD Markers.
- [145] Vieira RF, Goldsbrough P, Simon JE. Genetic diversity of basil (Ocimum spp.) based on RAPD markers. J Am Soc Hortic Sci 2003; 128(1): 94-9. [http://dx.doi.org/10.21273/JASHS.128.1.0094]
- [146] Rady MR, Nazif NM. Rosmarinic acid content and RAPD analysis of *in vitro* regenerated basil (Ocimum americanum) plants. Fitoterapia 2005; 76(6): 525-33. [http://dx.doi.org/10.1016/j.fitote.2005.04.001] [PMID: 16112496]
- [147] Giachino RRA, et al. RAPD and essential oil characterization of Turkish basil (Ocimum basilicum L.). Plant Syst Evol 2014; 300(8): 1779-91.
 [http://dx.doi.org/10.1007/s00606-014-1005-0]
- [148] Sundaram S, Purwar S. Assessment of genetic diversity among fenugreek (*Trigonella foenum*-graecum L.), using RAPD molecular markers. J Med Plants Res 2011; 5(9): 1543-8.
- [149] Mamatha N, et al. Molecular characterization of Fenugreek (*Trigonella foenum*-graecum L.) genotypes using RAPD markers. Int J Curr Microbiol Appl Sci 2017; 6: 2573-81. [http://dx.doi.org/10.20546/ijcmas.2017.606.306]

Spices Biotechnology

- [150] Choudhary S, et al. Genetic diversity in fenugreek assessed through RAPD-PCR. International J Seed Spices 2011; 1(1): 47-52.
- [151] Zhang Y, Hao J, Yao L. Analysis of homology of lavender with RAPD. Shanghai Jiaotong Daxue Xuebao Nongye Kexueban 2007; 25(6): 578-82.
- [152] Leila G, Abdelkader S, Azdinia Z. Molecular polymorphism in dentate lavender from littoral Algerian. Genetics and Biodiversity Journal 2019; 3(2): 40-8.
- [153] Varghese S, Thomas GE, Thomas G. AFLP analysis reveals exceptionally narrow genetic background in ginger (Zingiber officinale Rosc.). International Journal of Engineering. Science and Mathematics 2018; 7(2): 19-24.
- [154] Paul T, Debnath S. Recent researches on molecular breeding for spice crop improvement. Indian spices. Springer 2018; pp. 317-39.
 [http://dx.doi.org/10.1007/978-3-319-75016-3 11]
- [155] Kumar V, et al. Genetic diversity and identification of variety-specific AFLP markers in fenugreek (*Trigonella foenum*-graecum). Afr J Biotechnol 2012; 11(19): 4323-9.
- [156] Talib I, et al. Assessment of genetic diversity in fenugreek (Trigonella foenum-graecum) in Oman. Int J Agric Biol 2014; 16(4)
- [157] Jose S, Sujatha R, Deeshma K. Novel EST-SSR marker development and validation in black pepper cultivars and varieties. J Trop Agric 2018; 55(2): 175-9.
- [158] Peter K, et al. Breeding of spice crops (black pepper, cardamom, ginger and turmeric). 2008.
- [159] Singh TJ, et al. Molecular Diversity Analysis in Turmeric (Curcuma longa L.) Using SSR Markers. Int J Curr Microbiol Appl Sci 2018; 7(11): 552-60. [http://dx.doi.org/10.20546/ijcmas.2018.711.066]
- [160] Jain A, Jain P. DNA fingerprinting of cultivated and wild genotypes of Curcuma species from agroclimatic regions of Chhattisgarh. 2020. [http://dx.doi.org/10.21203/rs.3.rs-20321/v1]
- [161] Choudhary S. Microsatellites for coriander crop: A cross species amplification. Quarterly Research Journal of Plant & Animal Sciences/Bhartiya Krishi Anusandhan Patrika 2018; 33: 3. [http://dx.doi.org/10.18805/BKAP114]
- [162] Bharti R, Kumar S, Parekh MJ. Development of genomic simple sequence repeat (gSSR) markers in cumin and their application in diversity analyses and cross-transferability. Ind Crops Prod 2018; 111: 158-64.
 [http://dx.doi.org/10.1016/j.indcrop.2017.10.018]
- [163] Radhika RM, Sukanya D, Kaipa H. Study on cross-species transferability and DNA fingerprinting of Ashwagandha genotypes using SSR markers. Medicinal Plants-International Journal of Phytomedicines and Related Industries 2019; 11(4): 444-54. [http://dx.doi.org/10.5958/0975-6892.2019.00059.5]
- [164] Mathew KM, *et al.* Optimization of genomic DNA extraction from fresh and dry leaves of large cardamom (Amomum subulatum Roxb.) for diversity analysis. 2014.
- Barboza K, Beretta V, Kozub PC, *et al.* Microsatellite analysis and marker development in garlic: distribution in EST sequence, genetic diversity analysis, and marker transferability across Alliaceae. Mol Genet Genomics 2018; 293(5): 1091-106.
 [http://dx.doi.org/10.1007/s00438-018-1442-5] [PMID: 29705936]
- [166] Kumar M, Rakesh Sharma V, Kumar V, *et al.* Genetic diversity and population structure analysis of Indian garlic (*Allium sativum* L.) collection using SSR markers. Physiol Mol Biol Plants 2019; 25(2): 377-86.
 [http://dx.doi.org/10.1007/s12298-018-0628-y] [PMID: 30956421]

- [167] Barboza K, et al. Assessment of genetic diversity and population structure in a garlic (Allium sativum L.) germplasm collection varying in bulb content of pyruvate, phenolics, and solids. Sci Hortic (Amsterdam) 2020; 261: 108900.
 [http://dx.doi.org/10.1016/j.scienta.2019.108900]
- [168] Kaur Y, Dhall R, Sharma P. Characterization of bolting behaviour in garlic (*Allium sativum*) using SNP. Indian J Agric Sci 2020; 90(1): 112-7.
- [169] Tyagi R, Kak A. Registration of plant genetic resources in India–A review. Indian J Agric Sci 2012; 82(8): 651-9.
- [170] Yousefi Javan I, Gharari F. Genetic diversity in saffron (*Crocus sativus* L.) cultivars grown in Iran using SSR and SNP markers. J Agric Sci Technol 2018; 20(6): 1213-26.
- [171] Hebert PD, Cywinska A, Ball SL, deWaard JR. Biological identifications through DNA barcodes. Proc Biol Sci 2003; 270(1512): 313-21.
 [http://dx.doi.org/10.1098/rspb.2002.2218] [PMID: 12614582]
- [172] Costa FO, et al. Biological identifications through DNA barcodes: the case of the Crustacea. Can J Fish Aquat Sci 2007; 64(2): 272-95. [http://dx.doi.org/10.1139/f07-008]
- [173] Vohra P, Khera K. DNA barcoding: Current advances and future prospects-a review. Asian Journal of Biological and Life Sciences 2013; 2: 3.
- [174] Yang F, et al. DNA barcoding for the identification and authentication of animal species in traditional medicine. Evidence-Based Complementary and Alternative Medicine 2018.
- [175] Hollingsworth PM, Graham SW, Little DP. Choosing and using a plant DNA barcode. PLoS One 2011;
 6(5): e19254.
 [http://dx.doi.org/10.1371/journal.pone.0019254] [PMID: 21637336]
- [176] Liu Z, Zeng X, Yang D, Chu G, Yuan Z, Chen S. Applying DNA barcodes for identification of plant species in the family Araliaceae. Gene 2012; 499(1): 76-80. [http://dx.doi.org/10.1016/j.gene.2012.02.016] [PMID: 22406497]
- [177] P.P.-H. and S. Pang-Chui, Identification of herbal medicinal materials using DNA barcodes. J Syst Evol 2011; (3): 15.
- [178] Santana AC, et al. Evaluation of the shelf-life of vegetable-type soybean pods. Braz Arch Biol Technol 2012; 55(4): 591-5. [http://dx.doi.org/10.1590/S1516-89132012000400015]
- [179] Zhang M, et al. An efficient DNA barcoding based method for the authentication and adulteration detection of the powdered natural spices. Food Control 2019; 106: 106745. [http://dx.doi.org/10.1016/j.foodcont.2019.106745]
- [180] Hull R. Plant virology. Academic press 2013.
- [181] Barakate A, Stephens J. An overview of CRISPR-based tools and their improvements: new opportunities in understanding plant-pathogen interactions for better crop protection. Front Plant Sci 2016; 7: 765. [http://dx.doi.org/10.3389/fpls.2016.00765] [PMID: 27313592]
- [182] Bhat AI, Rao GP. Production of virus-resistant plants through CRISPR-Cas technology. Characterization of Plant Viruses. Springer 2020; pp. 511-20. [http://dx.doi.org/10.1007/978-1-0716-0334-5_50]
- [183] Chen K, Wang Y, Zhang R, Zhang H, Gao C. CRISPR/Cas genome editing and precision plant breeding in agriculture. Annu Rev Plant Biol 2019; 70: 667-97. [http://dx.doi.org/10.1146/annurev-arplant-050718-100049] [PMID: 30835493]

Spices Biotechnology

[184] Costa JMR, *et al.* CRIPR-Cas9 based strategy to engineer Saccharomyces cerevisiae towards the production of curcumin from ferulic acid. 2019.

Spices, the Guards Against the Evil Microbes: Antimicrobial Properties of Spices

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Abstract: Since time immemorable, spices have been known to combat the onslaught of various microbes like bacteria, fungi and viruses, responsible for various diseases. These microbes also led to food spoilage, which in turn reduced its shelf life. Spices can be used as food preservatives instead of chemical preservatives that are harmful to our health. Studies have proven that the spices commonly used in the kitchen like pepper, clove, ginger, coriander, garlic, cinnamon, etc., are highly potent anti-microbial agents. Moreover, they are also eminent anti-inflammatory and carminative agents. The essential oils in spices are also used for protection against various pathogens in plants. These properties are due to the various chemical compounds like eugenol, gingerol, flavonoids, terpenes, anthocyanins, phenylpropanoids and various organosulphur compounds among others present in spices. Hence, spices can be exploited for food preservation and in the pharmaceutical industries. They can also be used as biopesticides, insecticidal agents, antioxidants and natural colorants. This chapter highlights the effect of various spices on various micro-organisms, the various metabolites in spices that lend this ability, and also reviews the various works undertaken to understand the antimicrobial activity of spices.

Keywords: Spices, Anti-microbial, Metabolites, Food preservatives.

INTRODUCTION

The use of spices and herbs dates back to the prehistoric period when the huntergatherers wrapped the meat they hunted in leaves and found that this added the flavor and fragrance to meat. They also added some plant products and saw that

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spices improved its texture, aroma and also improved its shelf-life [1]. These plant products have been used ever since and have found a prominent position in history, religious and cultural practices. Many explorers carried out sea expeditions for this precious commodity, many lands were taken over and many treaties were signed for this precious commodity. Spices were among the most precious trade items during the medieval and ancient eras.

India is known as the "Land of spices". From the saffron of Kashmir to the pepper from Kerala, India is blessed in terms of the abundance of all types of spices due to favorable edaphic and climatic conditions. The health benefits of using spices in our everyday life have been elucidated in Ayurveda, which developed in India and is among the earliest systems of medicine in the universe. According to Ayurveda, using spices and herbs in a small amount in our diet keeps our body healthy. Over the years, humans have realized the essence of spices and understood their multiple uses as medicine, natural colorants, preservative, antioxidants, nutraceuticals, flavoring agents, immunity boosters and antimicrobial agents. Spices are also widely used in cosmetics, like essential oils and perfumes. In this chapter, we will focus on the anti-microbial activity of spices, their use in food preservation and as a medicine.

ANTIMICROBIAL ACTIVITY OF SPICES

Diseases caused by harmful pathogenic micro-organisms and food poisoning brought about by consuming food spoiled due to the activity of microbes are a great threat to human health all over the world. Essential oils extracted from spices by steam or water distillation of plant parts contain many active antimicrobial compounds. These compounds are found to be quite active against many bacteria, fungi, viruses and even many antibiotic-resistant microbes [2]. Regulatory agencies such as the European Union standards, the US Food and Drug Act, Codex Alimentarius, and Food Safety and Standards Authority of India have recognised spices, herbs, and their constituents as generally recognized as safe (GRAS) [3]. Hence spices have immense prospective to be developed as new and safe antimicrobial agents. Let us have a look at the main antimicrobial activities of the spices we generally use, their biochemical properties and their mode of action.

Anti-bacterial Activity of Spices

Bacteria are small microscopic organisms found everywhere, even inside the human body. Most bacteria are harmless, but the few that are pathogenic to humans cause fatal diseases. Many spices or their active metabolites can either kill or inhibit (reduce the rate of their growth) these pathogenic bacteria. The active ingredients present in the plant-derived spices and extracts have received

growing attention, not only for their active antibacterial activity but also due to the fact that developing resistance to them is relatively challenging. The spices possess antibacterial activity due to their innate ability to degrade the bacterial cell wall and cause cell lysis [4]. They can also cause loss of electrolytes, ATP, proteins, and DNA materials, through leakage caused by the damaged cell membranes [5]. The antibacterial activity of major spices is listed in Table 1.

S. No.	Spices/Herbs	Active Component	Antibacterial Effect On	References
1	Garlic	Allicin	Escherichia. coli, Salmonella species, Citrobacter, Enterobacter, Pseudomon, Klebsiella, Streptococcus, Bacillus anthrax	[6]
2	Ginger	Gingerol, Zerumbone, Zingerone	Porphyromonas gingivalis, P. endodontalis, Prevotella intermedia	[7]
			Pseudmonas aeruginosa, Salmonella choleraesuis, Bacillus subtilis	[8]
			Bacillus cereus, Staphlococcus aureus, E. coli, Yersinia enterocolitica	[9]
3	Tumeric	Curcuminoid,	E. coli, S. aureus, Salmonella typhi	[10, 11]
		Turmerone, Curlone	B. cereus, Bacillus coagulans, B. subtilis, S. aureus. E. coli and P. aeruginosa	[12]
4	Thyme	Thymol	B. subtilis, Salmonella enteritidis, P. aeruginosa	[13]
5	Clove	Eugenol	P.aeruginosa, S. aureus, S. choleraesuis, Klebsiella pneumoniae	[13]
6	Cinnamon	Cinnamaldehyde, eugenol, cinnamic acid, cinnamate	P.aeruginosa, S. aureus, E. coli, Bacillus megaterium, K. pneumonia, Enterobacter cloaca, Corynebacterium xerosis, Streptococcus faecalis, S. typhi, Pseudomonas fluorescens, Bacillus licheniformis, Y. enterocolitica, Proteus spp.	[14]
7	Cardamom	1, 8-cineole, α□terpinyl acetate, linalool	E. coli, P. aeruginosa, S. aureus, Bacillus pumilus	[15]
8	Pepper	Piperine, terpenes, phenols	S. typhimurium, Bacillus, E. coli, S. aureus	[16]
9	Cumin	Cuminaldehyde, cymene, terpenoids	E. coli, S. aureus, S. faecalis, P. aeruginosa, K. pneumoniae, B. megaterium, Brevibacillus brevis, Enterococcus faecalis, Pseudomonas pyocyaneus	[17]
10	Basil	Linalool, estragole, eugenol, 1,8-cineole	S. aureus, E. coli, B. subtilis, Pasteurella multocida	[18]

Table 1. List of spices with	their anti-bacterial effects.
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(Table 1	le 1) cont					
11	Oregano	Carvacrol and thymol	Staphylococcus gallinarum, Salmonella enteritidis, Salmonella typhimurium, S. aureus, B. subtilis, E. coli, P. aeruginosa	[18]		
12	Fenugreek	Galactomannan 4-OH isoleucine, steroidal saponin	E. coli, P. putida, S. typhimurium, S. aureus	[19]		
13	13 Rosemary p-cymene, linalool,		Leuconostoc mesenteroides, Lactobacillus delbruekii, S. cerevisiae, E. coli, S. typhimurium, S. enteritidis, Shigella sonei, Listeria monocytogenes	[20]		

Antifungal Activity of Spices

Fungi are ubiquitous microorganisms. Some of them, beneficial as food or as the basis for effective medicines, while other few are less desirable, such as food-mold or spores that bring about diseases in humans like skin infections, allergies and other diseases. The active component of several spices is known to completely or partially inhibit mycelial growth, penetrate the cell wall of various fungi and damage the organelle membranes [21], hinder the normal synthesis of DNA and proteins, and prevent cell wall formation and aflatoxin production, *etc.* A few examples of the antifungal activities of spices are listed in Table **2**.

S. No.	Spices/Herbs	Active Component	Antifungal Effect On	References
1	Garlic	Allicin	Candida albicans	[18, 22]
2	Ginger	Gingerol and shagelol	Aspergilus niger, Fusarium oxysporum	[23, 24]
3	Turmeric	Curcuminoid, turmerone, curdione, curcumol	C. albicans, Fusarium graminearum, Paracoccidioides brasiliensis, Aspergillus fumigatus, Aspergillus flavus, Aspergillus clavatus	[25, 26]
4	Thyme	Carvacrol and γ -terpinene	C. albicans, F. graminearum, Saccharomyces cerevisae	[27, 28]
5	Clove	Eugenol	C. albicans	[29]
6	Cinnamon	Cinnamaldehyde and eugenol	C. albicans, Aspergillus niger	[30, 31]
7	Cardamom	1,8-cineole, α-terpineol, terpinen-4-ol, spathulenol	A. flavus, C. albicans, S. cerevisiae	[32, 33]
8	Pepper	Piperine	F. graminearum	[34]
9	Cumin	Cuminaldehyde	C. albicans, A. niger, S. cerevisiae	[35]

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(Table 2)	(Table 2) cont					
10	Basil	Estragol	A. niger, A. flavus, Fusarium proliferatum, Fusarium subglutinans, F. oxysporum	[18, 22]		
11	Oregano	Carvacrol and thymol	C. albicans, A. niger, F. oxysporum, A. niger	[36, 37]		
12	Fenugreek	Galactomannan 4-OH isoleucine, steroidal saponin, cumarin	Microsporum gypseum, F. oxysporum, Trichoderma viridae	[38, 39]		
13	Rosemary	Rosmarinic acid	C. albicans	[40]		

Antiviral Activity of Spices

The outbreak of different human pathogenic viruses has brought research on viral diseases and its cure to the forefront. Many researchers are also looking at spices as probable anti-viral agents. These spices can effectually avert viral infections by acting as entry inhibitors, replication inhibitors, protease inhibitors, an integrase inhibitor, Tat protein inhibitor and can also inhibit gene expression. Thus the active chemicals in various spices prevent the virus from establishing itself in the host and thus prevents infection. Molecular docking experiments have shown that many Indian spices have potent activity against the SARS-CoV-2 virus. The antiviral activity of a few spices is mentioned in Table **3**.

S. No.	Spices/Herbs	Active Component	Antiviral Effect On	Family	References
1	Garlic	Allicin	HPV	Papilomaviridae	[41]
			Influenza A and B	Orthomyxoviridae	[42]
			HIV	Retroviridae	[43]
			HSV-1	Herpesviridae	[44]
			Rhinovirus	Picornaviridae	[45]
2	Ginger	Gingerol and shagelol	Influenza	Orthomyxoviridae	[46]
			RSV	Pneumoviridae	[47]
			FCV	Caliciviridae	[48]

Table 3. List of spices with their anti-viral activity.

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Table 3)	cont			·	
3	Turmeric	Curcuminoid, turmerone, curdione, curcumol	CHIKV	Togaviridae	[49]
			DENV	Flaviviridae	[50]
			HBV	Hepadnaviridae	[51]
			HCV	Flaviviridae	[52]
			HIV	Retroviridae	[53]
			HPV	Papilomaviridae	[54]
			HSV	Herpesviridae	[55]
			IAV	Orthomyxoviridae	[56]
			JEV	Flaviviridae	[57]
			MNV	Caliciviridae	[58]
			RSV	Pneumoviridae	[56]
			RVFV	Phenuiviridae	[59]
			ZIKV	Flaviviridae	[60]
			H5N1	Orthomyxoviridae	[61, 62]
5	Clove	Eugenol	FCV	Caliciviridae	[48]
			HSV1	Herpesviridae	[63]
6	Cinnamon	Cinnamaldehyde and	HSV1	Herpesviridae	[64]
		eugenol	H1N1	Orthomyxoviridae	[64]
7	Cardamom	1,8-cineole, α-terpineol,	HSV1, HSV2, HSV6,	Herpesviridae	[65]
		terpinen-4-ol, spathulenol	HBV	Hepadnaviridae	[65]
			HCV	Flaviviridae	[65]
8	Pepper	Piperine	CVB3	Picornaviridae	[66]
9	Cumin	Cuminaldehyde	HSV1	Herpesviridae	[67]
11	Oregano	Carvacrol and thymol	MNV	Caliciviridae	[68]
			HSV-1	Herpesviridae	[68]
			RSV	Pneumoviridae	[69, 70]
12	Sage	safficinolide and sageone	HIV-1	Retroviridae	[71]
			HSV-1	Herpesviridae	[72]
13	Rosemary	Oleanolic acid	HSV1	Herpesviridae	[73]
			H1N1	Orthomyxoviridae	
			HIV	Retroviridae	[74]
14	Basil	Estragol, apigenin and	Herpes viruses	Herpesviridae	[75]
		ursolic acid	HBV	Hepadnaviridae	[76]
			Enterovirus	Picornaviridae	[76]

Apart from the wide range of action against Gram-positive and Gram-negative bacteria, yeasts and fungal pathogens, spices also have a broad spectrum of activity against molds. As mold spoilage is a big concern for both cheese manufacturers and marketers, causing the development of off-flavor and production of toxins the spices like black pepper along with mesquite smoke and hickory smoke oil used as flavouring can also aid in controlling the molds. According to Bachmann [77], when large quantities of spices like cinnamon, cloves, and allspice are used, the growth of molds is retarded, thus acting as a preservative. The mold growth is inversely dependent on the amount of spice used and the inhibition of mold is due to the diffusion and volatilization of the spice. The essential oil derived from bush-basil possess anti-Fungal activity against molds like Penicillium islandicum [78], Alternaria sp., Mucor [79] Aspergillus flavus, Botrytis cinereal cinereal and Aspergillus niger [80]. Coriander EO showed antifungal activity against *M. racemosus*, *A. alternata* and *P chrysogenum* [82]. Alves-Silva, dos Santos [81] reported an antagonistic effect for mold inhibition when cinnamaldehyde was combined with essential oil of clove, also additive effect on mold inhibition was obtained when different eugenol and peppermint combination was used in tomato fruit.

ESSENTIAL OILS EXTRACTION FROM SPICES

Essential oil is the essence of the plant. It consists of concentrated hydrophobic liquid which consists of volatile chemical compounds from plants [83]. An essential oil may comprise of several hundreds of chemical compounds and this is what gives it, its distinctive fragrance and flavour. Essential oils from spices are being widely utilized and are a rich source of several bioactive compounds with quite a lot of antioxidative and antimicrobial properties [84] as mentioned earlier. Instead of being synthetically manufactured in laboratories, essential oils are extracted from plant materials through removal methods that are specific for plant parts containing the oils. Commonly, essential oils are extracted by the process of hydrodistillation, steam distillation and solvent extraction. Other processes include supercritical fluid extraction, absolute oil extraction, solvent extraction, resin tapping, *sfumatura*, wax embedding, and cold pressing, pulsed electric fields, microwave, ultrasound, ohmic-heating and microwave (MW) extraction techniques [85]. Essential oils have been widely used in cosmetics, perfumes, soaps and other products, to add-on fragrances to incense and household cleaning products and also as a food and drink flavouring agent. They may also be fractioned and sold as individual natural components.

USE OF SPICES IN PHARMACEUTICAL INDUSTRY

Spices are used widely for drug development. Ancient medical systems like Ayurveda and Unani have exploited the innumerable health benefits and healing properties of spices effectively. Turmeric is a well-known example of a spice used for wound healing, skin allergies and other infections [86]. Garlic and ginger have been found effective against antibiotic-resistant bacteria such as *B. subtilis, E. coli, K. pneumoniae, P. aeruginosa, Shigella, S. aureus,* and *S.typhi* [87]. Many spices such as mint, clove, camphor, and ginger have been used against microbes that harm oral health. The gingerol in ginger is quite effective against *Porphyromonas gingivalis, P. endodontalis, Prevotella intermedia* [7]. Even deadly diseases like HIV can be controlled by methanolic extracts of rosemary, cinnamon, sage [88]. Thus understanding the specific antimicrobial activity of the spices can help developing new drugs. Molecular docking approaches are being used with active compounds of spices as lead molecules for drug discovery. Extensive research is being carried out for discovering a drug against Covid-19 from Indian spices [89].

SPICES AS FOOD PRESERVATIVES

Have you ever wondered why the food in the tropical regions has more spices than those in the comparatively colder regions? It is because spices delay the process of spoiling due to microbes. Due to higher temperature and humidity in the tropical regions, the contamination of food by microbes was faster and hence they started adding more spices for food preservation. Moreover, it enhanced the taste.

Spoilage of food is an irreversible modification of food, making it unfit for human consumption or a compromise in its quality. It can be due to physical (oxygen, temperature, light) and/or biological (enzymatic activity and microbial growth) factors. Inspite of the advanced technologies available within the assembly chain (for instance, freezing, pasteurization, drying, preservatives), it appears impossible to do away with the risk of food spoiling [90].

One of the primary causes of food spoiling is lipid oxidation. Hence, food industries consider the application of antioxidants such as butylated hydroxytoluene (BHT) and butylated hydroxyanisole (BHA) of the utmost importance so as to prevent spoilage [91]. But the use of these chemicals may not be safe and so the consumers are increasingly demanding the use of natural compounds. The antioxidant capacity of the spices can be attributed to the presence of phenolic compounds, and they do so by chelating transition metals, scavenging free radicals, quenching singlet oxygen, and enhancing the activities

of antioxidant enzymes [92]. Many surveys have discovered the utility of spices against food spoilage [92 - 95].

The antimicrobial activity of spices is exerted in two different ways: by preventing the growth of microorganisms that cause food spoilage (food preservation), and/or by inhibiting/regulating the development of these pathogens (food safety) [96]. Some of the natural antimicrobials reported for meat products include lactoferrin, bacteriocins, essential oils, lysozyme species, and a variety of plant extracts. Spices, for instance, cinnamon, clove, cumin and oregano, were found to be effective against the gram-positive and gram-negative bacteria inoculated on meat [31, 97, 98].

Pros & Cons of Using Spices as Preservatives

The antioxidant and antimicrobial activity of spices has a crucial role in preserving the food. But, for ensuring the effectiveness of spices as preservatives, we need to study several aspects. As reviewed, various spices have different flavours and varying levels of aroma but they are commonly categorised as being strong and so they may interfere with the original flavour of the food, if used in more measures to achieve good antioxidant or antimicrobial activities. Essential oils extracted from spices maybe good substitutes for meat preservation. But when used in meat, the high levels of fat and protein can protect the bacteria from the action of essential oils, as essential oils easily dissolve in the fatty phase of the food, thus reducing their availability to act against the microorganisms [99].

Encapsulated rosemary essential oil has a higher antimicrobial effect against L. *monocytogenes* in pork liver sausage than standard rosemary essential oil and this is associated with the interaction of essential oils with the fatty phase of meat [100]. Thus, using higher concentration of spices may ensure better antioxidant and antimicrobial activity but the strong flavour of spices may affect its commercial value due to the alteration in the flavour of meat. Essential oils also need the additional extraction process from spices which can make it more expensive eventhough it may not have a better antimicrobial activity. Therefore, using whole spices might be a better alternative for food preservation as they are cheaper, less complex, and have equivalent antimicrobial activity. Another fact is that the efficacy of the spice formulation against microorganisms fluctuates depending on the food or media, the same formulation can be effective for one type of meat but not for another.

SPICES IN PACKAGING MATERIAL

Active Packaging (AP) is a high-tech concept for food packaging that combines the advances in packaging, material sciences and food technology, for food preservation. In antimicrobial packaging, antimicrobial agents can be infused into the packaging material, smeared over the surface of packaging film or an antimicrobial compound sachet can be added into the package [101]. Sachets and pads being the most successful applications of active food packaging. Active antimicrobial packaging gives a headspace to reduce, retard, or even inhibit the growth of spoilage and pathogenic microorganisms by interacting with packaged food. The transfer of active compounds to food may be achieved through the direct contact between food and the packaging material or through gas-phase diffusion from the inner packaging layer to the food surface [102].

The addition of EOs to a wax coating for antimicrobial active packaging of strawberry preservation was studied. The release of antimicrobial agents from the coating to prevent contamination by microorganisms was evaluated [103, 104]. In another study, carvacrol was added to chitosan-based films for active packaging, and its antimicrobial efficiency against microorganisms that cause food spoilage was demonstrated by using a headspace chromatographic technique. The shelf-life of a complex bakery product was increased by more than three times using cinnamon-based active material with minimal changes in the packaging and no extra manipulation steps [105].

Prevention of Toxin Production

Certain filamentous fungi produce mycotoxins as secondary metabolites, which may contaminate agricultural commodities. They are toxic to humans and animals, cause a substantial drop in crop yield and thus economic losses. Food contaminated with mycotoxins, particularly with aflatoxins, can lead to fatal acute illness and are even related to increased cancer risk [106]. Many countries have imposed strict limits in the aflatoxin concentration due to its high risk to human health. The US Food and Drug Administration (USFDA) has established 20 ppb as the minimum level of aflatoxin acceptable, in all foods other than milk. The European Union has put a ban on the import of peanuts having >2 ppb of AFB1 content and >4 ppb of total aflatoxins in nuts prepared for human consumption. For the export of tree nuts to the European market, the level of aflatoxin should be <3 ppb [107].

Plant extracts or essential oils can be utilized as promising alternatives to the toxic fungicides, currently used for controlling post-harvest fungal deterioration. The extracts of certain plants, toxic to fungi, can impede the biosynthesis of aflatoxin B1 and thus control fungal growth and mycotoxin production. Extracts of garlic and onion can efficiently check growth and aflatoxin production. The effect of turmeric leaf oil on fungal growth and aflatoxin production was determined [107]. Essential oils from aromatic plants like coriander, ginger, pepper, rosemary,

cinnamon, and thyme were found effective against *A. flavus* IMI 242684 on PDA [108]. P. *betle* chloroform fraction substantially reduced the aflatoxin B1 production. This fraction caused a decrease of 91% in mycelial growth and completely inhibited toxin biosynthesis by A. *flavus* at 500 µg/ml [106].

Edible Films

Edible films are thin films prepared from edible material that acts as a barrier to external elements (factors such as oils, gases, moisture, and vapors) thus protecting the product, improving its quality and extending its shelf life. Edible films can control the transfer of moisture, oxygen, carbon dioxide, flavor, and aroma between food components or the atmosphere and the food [109]. Recently, more research has been conducted on biodegradable films, which are made from plant and animal edible protein sources like cottonseed, soy, wheat gluten, corn zein and peanut protein, albumin, casein, collagen, gelatin, and whey proteins. The addition of essential oils to coatings and edible films inhibits the growth of pathogenic and spoilage bacteria. It boosts the sensory properties of foods like meat and meat products. The essential oils of garlic, cinnamon, lemongrass and oregano are widely used for applications on food wrappings [110]. Using apple films comprising 0.5% carvacrol or cinnamaldehyde- the active ingredients of oregano and cinnamon oils, respectively for wrapping pieces of chicken, inhibited the growth rate of E. coli O157:H7, Salmonella enterica, and L. monocytogenes [36].

CONCLUSION

Spices are valuable resources. Many spices used daily have been shown to have antimicrobial activity and medicinal value. They have been used extensively in pharmaceutical industries and also for food preservation. The active chemical components obtained by different extraction methods are quite effective against various bacteria, fungi, viruses and many other pathogenic microbes. They are safe to use and can also eliminate antibiotic-resistant bacteria. Medicinal systems like Ayurveda and Unani have been using spices, and they are found to be successful in treating many diseases. They are also widely used in food preservation. However, there is a need for much more extensive research to escalate the stability of the active compounds, proper target delivery, drug development, *etc.*, to reap the complete benefits of spices. Moreover, the multiple possibilities of using various spices individually and also in different combinations against the different pathogens have to be explored. **Antimicrobial Properties**

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The authors declared that there is no conflict of interest.

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REFERENCES

- Jessica Elizabeth T, Gassara F, Kouassi AP, Brar SK, Belkacemi K. Spice use in food: Properties and benefits. Crit Rev Food Sci Nutr 2017; 57(6): 1078-88.
 [http://dx.doi.org/10.1080/10408398.2013.858235] [PMID: 26560460]
- [2] Liu Q, Meng X, Li Y, Zhao CN, Tang GY, Li HB. Antibacterial and antifungal activities of spices. Int J Mol Sci 2017; 18(6): 1283. [http://dx.doi.org/10.3390/ijms18061283] [PMID: 28621716]
- [3] Dhiman R, *et al.* Comparative evaluation of antimicrobial activities of commonly used indian spices against microbes associated with juices. Research Journal of Microbiology 2015; 10(4): 170. [http://dx.doi.org/10.3923/jm.2015.170.180]
- [4] Burt S. Essential oils: their antibacterial properties and potential applications in foods--a review. Int J Food Microbiol 2004; 94(3): 223-53. [http://dx.doi.org/10.1016/j.ijfoodmicro.2004.03.022] [PMID: 15246235]
- [5] Zhang J, Ye KP, Zhang X, Pan DD, Sun YY, Cao JX. Antibacterial activity and mechanism of action of black pepper essential oil on meat-borne Escherichia coli. Front Microbiol 2017; 7: 2094. [http://dx.doi.org/10.3389/fmicb.2016.02094] [PMID: 28101081]
- [6] Daka D. Antibacterial effect of garlic (*Allium sativum*) on Staphyloccus aureus: An *in vitro* study. Afr J Biotechnol 2011; 10(4): 666-9.
- [7] Park M, Bae J, Lee DS. Antibacterial activity of [10]-gingerol and [12]-gingerol isolated from ginger rhizome against periodontal bacteria. Phytother Res 2008; 22(11): 1446-9. [http://dx.doi.org/10.1002/ptr.2473] [PMID: 18814211]
- [8] Abdul AB, Abdelwahab SI, Al-Zubairi AS, Elhassan MM, Murali SM. Anticancer and Antimicrobial Activities of Zerumbone from the Rhizomes of Zingiber zerumbut. Int JPharmacol 2008; 4(4): 301-4. [http://dx.doi.org/10.3923/ijp.2008.301.304]
- [9] Singh G, et al. Studies on essential oils, Part 42: chemical, antifungal, antioxidant and sprout suppressant studies on ginger essential oil and its oleoresin. Flavour Fragrance J 2005; 20(1): 1-6. [http://dx.doi.org/10.1002/ffj.1373]
- [10] Mahady GB. Medicinal plants for the prevention and treatment of bacterial infections. Curr Pharm Des 2005; 11(19): 2405-27.
 [http://dx.doi.org/10.2174/1381612054367481] [PMID: 16026296]
- [11] Gupta S, Ravishankar S. A comparison of the antimicrobial activity of garlic, ginger, carrot, and turmeric pastes against Escherichia coli O157:H7 in laboratory buffer and ground beef. Foodborne Pathog Dis 2005; 2(4): 330-40.
 [http://dx.doi.org/10.1089/fpd.2005.2.330] [PMID: 16366855]
- [12] Negi PS, Jayaprakasha GK, Jagan Mohan Rao L, Sakariah KK. Antibacterial activity of turmeric oil: a

byproduct from curcumin manufacture. J Agric Food Chem 1999; 47(10): 4297-300. [http://dx.doi.org/10.1021/jf990308d] [PMID: 10552805]

- Cosentino S, Tuberoso CI, Pisano B, et al. In-vitro antimicrobial activity and chemical composition of [13] Sardinian Thymus essential oils. Lett Appl Microbiol 1999; 29(2): 130-5. [http://dx.doi.org/10.1046/j.1472-765X.1999.00605.x] [PMID: 10499301]
- Nabavi SF, Di Lorenzo A, Izadi M, Sobarzo-Sánchez E, Daglia M, Nabavi SM. Antibacterial effects [14] of cinnamon: From farm to food, cosmetic and pharmaceutical industries. Nutrients 2015; 7(9): 7729-48.

[http://dx.doi.org/10.3390/nu7095359] [PMID: 26378575]

- [15] Singh G, et al. Antioxidant and antimicrobial activities of essential oil and various oleoresins of Elettaria cardamomum (seeds and pods). J Sci Food Agric 2008; 88(2): 280-9. [http://dx.doi.org/10.1002/jsfa.3087]
- Zou L, Hu Y-Y, Chen W-X. Antibacterial mechanism and activities of black pepper chloroform [16] extract. J Food Sci Technol 2015; 52(12): 8196-203. [http://dx.doi.org/10.1007/s13197-015-1914-0] [PMID: 26604394]
- Allahghadri T, Rasooli I, Owlia P, et al. Antimicrobial property, antioxidant capacity, and cytotoxicity [17] of essential oil from cumin produced in Iran. J Food Sci 2010; 75(2): H54-61. [http://dx.doi.org/10.1111/j.1750-3841.2009.01467.x] [PMID: 20492235]
- [18] Hussain AI, Anwar F, Hussain Sherazi ST, Przybylski R. Chemical composition, antioxidant and antimicrobial activities of basil (Ocimum basilicum) essential oils depends on seasonal variations. Food Chem 2008; 108(3): 986-95. [http://dx.doi.org/10.1016/j.foodchem.2007.12.010] [PMID: 26065762]
- [19] Goyal S, et al. Investigating therapeutic potential of Trigonella foenum-graecum L. as our defense mechanism against several human diseases. Journal of Toxicology 2016.
- [20] Tavassoli S, Djomeh ZE. Total phenols, antioxidant potential and antimicrobial activity of methanol extract of rosemary (Rosmarinus officinalis L.). Glob Vet 2011; 7(4): 337-41.
- [21] Li W-R, Shi QS, Dai HQ, et al. Antifungal activity, kinetics and molecular mechanism of action of garlic oil against Candida albicans. Sci Rep 2016; 6: 22805. [http://dx.doi.org/10.1038/srep22805] [PMID: 26948845]
- Beatovic D, et al. Chemical composition, antioxidant and antimicrobial activities of the essential oils [22] of twelve Ocimum basilicum L. cultivars grown in Serbia. Rec Nat Prod 2015; 9(1): 62.
- [23] Singh G. Maurva S. Antimicrobial, antifungal and insecticidal investigations on essential oils; an overview. 2005.
- [24] Dwivedi S, Dwivedi N. Antifungal activity of some plant extracts against guava wilt pathogen. Int J Environ Sci 2012; 3(1): 412-20.
- Serra E, Hidalgo-Bastida LA, Verran J, Williams D, Malic S. Antifungal activity of commercial [25] essential oils and biocides against Candida albicans. Pathogens 2018; 7(1): 15. [http://dx.doi.org/10.3390/pathogens7010015] [PMID: 29370147]
- [26] Satish S, et al. Antifungal activity of some plant extracts against important seed borne pathogens of Aspergillus sp. Agric Technol Thail 2007; 3(1): 109-19.
- [27] Chemical composition and antimicrobial activity of essential oil of Thymus vulgaris from Yemen. Turkish J Biochem 2011; 36: 342-9.
- Girova T, et al. Antimicrobial activity of essential oils from spices against psychrotrophic food [28] spoilage microorganisms. Biotechnology & Biotechnological Equipment, 2010; 24(sup1): 547-52.
- [29] Schmidt E, et al. Antifungal activity of eugenol and various eugenol-containing essential oils against 38 clinical isolates of Candida albicans. J Essent Oil-Bear Plants 2007; 10(5): 421-9. [http://dx.doi.org/10.1080/0972060X.2007.10643575]

Antimicrobial Properties

- [30] Hussien J, et al. Assessment of the antimicrobial effects of some Ethiopian aromatic spice and herb hydrosols. Int J Pharmacol 2011; 7(5): 635-40. [http://dx.doi.org/10.3923/ijp.2011.635.640]
- [31] Ağaoğlu S, et al. Antimicrobial activity of some spices used in the meat industry. Bull Vet Inst Pulawy 2007; 51: 53-7.
- [32] Agnihotri S, Wakode S. Antimicrobial activity of essential oil and various extracts of fruits of greater cardamom. Indian J Pharm Sci 2010; 72(5): 657-9. [http://dx.doi.org/10.4103/0250-474X.78542] [PMID: 21695005]
- [33] Adegoke GO, *et al.* African cardamom (Aframomum danielli) oils, in Essential Oils in Food Preservation, Flavor and Safety. Elsevier 2016; pp. 163-71.
- [34] Singh G, et al. Chemical, antioxidant and antifungal activities of volatile oil of black pepper and its acetone extract. J Sci Food Agric 2004; 84(14): 1878-84. [http://dx.doi.org/10.1002/jsfa.1863]
- [35] Mageed MAAE, et al. Effect of microwaves on essential oils of coriander and cumin seeds and on their antioxidant and antimicrobial activities. J Essent Oil-Bear Plants 2012; 15(4): 614-27. [http://dx.doi.org/10.1080/0972060X.2012.10644096]
- [36] Santoyo S, Cavero S, Jaime L, Ibañez E, Señoráns FJ, Reglero G. Supercritical carbon dioxide extraction of compounds with antimicrobial activity from *Origanum vulgare* L.: determination of optimal extraction parameters. J Food Prot 2006; 69(2): 369-75. [http://dx.doi.org/10.4315/0362-028X-69.2.369] [PMID: 16496578]
- [37] Souza ELd, et al. Interference of heating on the antimicrobial activity and chemical composition of Origanum vulgare L.(Lamiaceae) essential oil. Food Sci Technol (Campinas) 2008; 28(2): 418-22. [http://dx.doi.org/10.1590/S0101-20612008000200023]
- [38] Dharajiya D, et al. Evaluation of antibacterial and antifungal activity of fenugreek (*Trigonella foenum-graecum*) extracts. Int J Pharm Pharm Sci 2016; 8(4): 212-7.
- [39] Sudan P, et al. Antifungal potential of fenugreek seeds (*Trigonella foenum-graecum*) crude extracts against Microsporum gypseum. International Journal of Research in Pharmaceutical Sciences 2020; 11(1): 646-9. [http://dx.doi.org/10.26452/ijrps.v11i1.1870]
- [40] Bozin B, Mimica-Dukic N, Samojlik I, Jovin E. Antimicrobial and antioxidant properties of rosemary and sage (*Rosmarinus officinalis* L. and *Salvia officinalis* L., Lamiaceae) essential oils. J Agric Food Chem 2007; 55(19): 7879-85. [http://dx.doi.org/10.1021/jf0715323] [PMID: 17708648]
- [41] Dehghani F, Merat A, Panjehshahin MR, Handjani F. Healing effect of garlic extract on warts and corns. Int J Dermatol 2005; 44(7): 612-5. [http://dx.doi.org/10.1111/j.1365-4632.2004.02348.x] [PMID: 15985039]
- [42] Fenwick GR, Hanley AB. Allium species poisoning. Vet Rec 1985; 116(1): 28-8.
 [http://dx.doi.org/10.1136/vr.116.1.28] [PMID: 3984170]
- [43] Shoji S, Furuishi K, Yanase R, Miyazaka T, Kino M. Allyl compounds selectively killed human immunodeficiency virus (type 1)-infected cells. Biochem Biophys Res Commun 1993; 194(2): 610-21. [http://dx.doi.org/10.1006/bbrc.1993.1865] [PMID: 8343148]
- [44] Tsai Y, Cole LL, Davis LE, Lockwood SJ, Simmons V, Wild GC. Antiviral properties of garlic: in vitro effects on influenza B, herpes simplex and coxsackie viruses. Planta Med 1985; 51(5): 460-1. [http://dx.doi.org/10.1055/s-2007-969553] [PMID: 17342616]
- [45] Weber ND, Andersen DO, North JA, Murray BK, Lawson LD, Hughes BG. *In vitro* virucidal effects of *Allium sativum* (garlic) extract and compounds. Planta Med 1992; 58(5): 417-23. [http://dx.doi.org/10.1055/s-2006-961504] [PMID: 1470664]

- [46] Rasool A, Khan MU, Ali MA, *et al.* Anti-avian influenza virus H9N2 activity of aqueous extracts of *Zingiber officinalis* (Ginger) and *Allium sativum* (Garlic) in chick embryos. Pak J Pharm Sci 2017; 30 (4): 1341-4.
 [PMID: 29039335]
- [47] Strauch MA, Tomaz MA, Monteiro-Machado M, et al. Antiophidic activity of the extract of the Amazon plant Humirianthera ampla and constituents. J Ethnopharmacol 2013; 145(1): 50-8. [http://dx.doi.org/10.1016/j.jep.2012.10.033] [PMID: 23123799]
- [48] Aboubakr HA, Nauertz A, Luong NT, et al. In vitro antiviral activity of clove and ginger aqueous extracts against feline calicivirus, a surrogate for human norovirus. J Food Prot 2016; 79(6): 1001-12. [http://dx.doi.org/10.4315/0362-028X.JFP-15-593] [PMID: 27296605]
- [49] von Rhein C, Weidner T, Henß L, et al. Curcumin and Boswellia serrata gum resin extract inhibit chikungunya and vesicular stomatitis virus infections in vitro. Antiviral Res 2016; 125: 51-7. [http://dx.doi.org/10.1016/j.antiviral.2015.11.007] [PMID: 26611396]
- [50] Saptawati L, et al. Health Science Journal of Indonesia 2017; 8(1): 63814. [http://dx.doi.org/10.22435/hsji.v8i1.6601.12-18]
- [51] Prasad S, Aggarwal BB. Turmeric, the golden spice: from traditional medicine to modern medicine. 2011.
 [http://dx.doi.org/10.1201/b10787-14]
- [52] Anggakusuma CC, Colpitts CC, Schang LM, et al. Turmeric curcumin inhibits entry of all hepatitis C virus genotypes into human liver cells. Gut 2014; 63(7): 1137-49. [http://dx.doi.org/10.1136/gutjnl-2012-304299] [PMID: 23903236]
- [53] Niranjan A, Prakash D. Chemical constituents and biological activities of turmeric (*Curcuma longa* 1.)-a review. J Food Sci Technol 2008; 45(2): 109.
- [54] Shukla DP, et al. Anticancer and cytotoxic potential of turmeric (Curcuma longa), neem (Azadirachta indica), tulasi (Ocimum sanctum) and ginger (Zingiber officinale) extracts on HeLa cell line. Int J Life Sci Sci Res 2016; 2: 309-15. [http://dx.doi.org/10.21276/ijlssr.2016.2.4.2]
- [55] Son M, Lee M, Sung GH, et al. Bioactive activities of natural products against herpesvirus infection. J Microbiol 2013; 51(5): 545-51.
 [http://dx.doi.org/10.1007/s12275-013-3450-9] [PMID: 24173639]
- [56] Dai J, Gu L, Su Y, *et al.* Inhibition of curcumin on influenza A virus infection and influenzal pneumonia *via* oxidative stress, TLR2/4, p38/JNK MAPK and NF-κB pathways. Int Immunopharmacol 2018; 54: 177-87. [http://dx.doi.org/10.1016/j.intimp.2017.11.009] [PMID: 29153953]
- [57] Marathe SA, Datey AA, Chakravortty D. Herbal cocktail as anti-infective: promising therapeutic for the treatment of viral diseases. Recent Patents Anti-Infect Drug Disc 2012; 7(2): 123-32. [http://dx.doi.org/10.2174/157489112801619692] [PMID: 22630820]
- [58] Wu J, Hou W, Cao B, et al. Virucidal efficacy of treatment with photodynamically activated curcumin on murine norovirus bio-accumulated in oysters. Photodiagn Photodyn Ther 2015; 12(3): 385-92. [http://dx.doi.org/10.1016/j.pdpdt.2015.06.005] [PMID: 26117199]
- [59] Juhua L, Zhongling Z, Chuanju Z. Therapeutic Effect of Aromatic Turmeric Oil (ATO, Oleum Curcumae Aromaticae) on Pneumonia Caused by Respiratory Syncytial Virus (RSV). LIFERATUE AND INFORMATION ON PREVENTINE 2000; p. 3. J
- [60] Ahmad RS, et al. Biochemistry, Safety, Pharmacological Activities, and Clinical Applications of Turmeric: A Mechanistic Review. Evidence-Based Complementary and Alternative Medicine 2020.
- [61] Gupta H, Gupta M, Bhargava S. Potential use of turmeric in COVID-19. Clin Exp Dermatol 2020; 45 (7): 902-3.

[http://dx.doi.org/10.1111/ced.14357] [PMID: 32608046]

- [62] Rocha F, de Assis M. Turmeric against Covid-19: too much of a coincidence? 2020.
- [63] El-Saber Batiha G, Magdy Beshbishy A, El-Mleeh A, Abdel-Daim MM, Prasad Devkota H. Traditional uses, bioactive chemical constituents, and pharmacological and toxicological activities of Glycyrrhiza glabra L.(Fabaceae). Biomolecules 2020; 10(3): E352. [http://dx.doi.org/10.3390/biom10030352] [PMID: 32106571]
- [64] Brochot A, Guilbot A, Haddioui L, Roques C. Antibacterial, antifungal, and antiviral effects of three essential oil blends. MicrobiologyOpen 2017; 6(4): e00459. [http://dx.doi.org/10.1002/mbo3.459] [PMID: 28296357]
- [65] Omer OL. Plant-based therapeutic agent with virustatic and antiviral effect. Google Patents 1998.
- [66] Mair C, *et al.* Antiviral and anti-proliferative *in vitro* activities of piperamides from black pepper. Planta Medica 2016; 82(S 01): P807.
- [67] Motamedifar M, *et al.* The effect of Cumin seed extracts against herpes simplex virus type 1 in Vero cell culture. 2010.
- [68] Gilling DH, Kitajima M, Torrey JR, Bright KR. Antiviral efficacy and mechanisms of action of oregano essential oil and its primary component carvacrol against murine norovirus. J Appl Microbiol 2014; 116(5): 1149-63. [http://dx.doi.org/10.1111/jam.12453] [PMID: 24779581]
- [69] Sharifi-Rad J, Salehi B, Schnitzler P, et al. Susceptibility of herpes simplex virus type 1 to monoterpenes thymol, carvacrol, p-cymene and essential oils of Sinapis arvensis L., Lallemantia royleana Benth. and Pulicaria vulgaris Gaertn. Cell Mol Biol 2017; 63(8): 42-7. [http://dx.doi.org/10.14715/cmb/2017.63.8.10] [PMID: 28886313]
- [70] Pilau MR, Alves SH, Weiblen R, Arenhart S, Cueto AP, Lovato LT. Antiviral activity of the Lippia graveolens (Mexican oregano) essential oil and its main compound carvacrol against human and animal viruses. Braz J Microbiol 2011; 42(4): 1616-24. [http://dx.doi.org/10.1590/S1517-83822011000400049] [PMID: 24031796]
- [71] Geuenich S, Goffinet C, Venzke S, et al. Aqueous extracts from peppermint, sage and lemon balm leaves display potent anti-HIV-1 activity by increasing the virion density. Retrovirology 2008; 5(1): 27. [http://dx.doi.org/10.1186/1742-4690-5-27] [PMID: 18355409]
- Santoyo S, *et al.* Antiviral properties of supercritical CO2 extracts from oregano and sage. Int J Food Prop 2014; 17(5): 1150-61.
 [http://dx.doi.org/10.1080/10942912.2012.700539]
- [73] Nolkemper S, Reichling J, Stintzing FC, Carle R, Schnitzler P. Antiviral effect of aqueous extracts from species of the Lamiaceae family against Herpes simplex virus type 1 and type 2 *in vitro*. Planta Med 2006; 72(15): 1378-82.
 [http://dx.doi.org/10.1055/s-2006-951719] [PMID: 17091431]
- Battistini R, Rossini I, Ercolini C, *et al.* antiviral activity of essential oils against hepatitis A virus in soft fruits. Food Environ Virol 2019; 11(1): 90-5.
 [http://dx.doi.org/10.1007/s12560-019-09367-3] [PMID: 30684236]
- [75] Chiang LC, Ng LT, Cheng PW, Chiang W, Lin CC. Antiviral activities of extracts and selected pure constituents of Ocimum basilicum. Clin Exp Pharmacol Physiol 2005; 32(10): 811-6. [http://dx.doi.org/10.1111/j.1440-1681.2005.04270.x] [PMID: 16173941]
- [76] Ravindran P, Divakaran M. Handbook of herbs and spices Wood Publishing series in Food Science, Technology and nutrition. 2012.
- [77] Bachmann FM. The inhibiting action of certain spices on the growth of microÖrganisms. Ind Eng Chem 1916; 8(7): 620-3.
 [http://dx.doi.org/10.1021/i500007a014]

- [78] López P, Sánchez C, Batlle R, Nerín C. Solid- and vapor-phase antimicrobial activities of six essential oils: susceptibility of selected foodborne bacterial and fungal strains. J Agric Food Chem 2005; 53(17): 6939-46.
 [http://dx.doi.org/10.1021/jf050709v] [PMID: 16104824]
- [79] Soni U, et al. Effect of seasonal variation on secondary metabolites of medicinal plants. Int J Pharm Sci Res 2015; 6(9): 3654-62.
- [80] Pawar VC, Thaker VS. In vitro efficacy of 75 essential oils against Aspergillus niger. Mycoses 2006; 49(4): 316-23.
 [http://dx.doi.org/10.1111/j.1439-0507.2006.01241.x] [PMID: 16784447]
- [81] Alves-Silva JM, et al. Chemical composition and in vitro antimicrobial, antifungal and antioxidant properties of essential oils obtained from some herbs widely used in Portugal. Food Control 2013; 32 (2): 371-8.

[http://dx.doi.org/10.1016/j.foodcont.2012.12.022]

- [82] Tejeswini MG, et al. Antifungal activity of essential oils and their combinations in *in vitro* and *in vivo* conditions. Arch Phytopathol Pflanzenschutz 2014; 47(5): 564-70. [http://dx.doi.org/10.1080/03235408.2013.814235]
- [83] Tongnuanchan P, Benjakul S. Essential oils: extraction, bioactivities, and their uses for food preservation. J Food Sci 2014; 79(7): R1231-49. [http://dx.doi.org/10.1111/1750-3841.12492] [PMID: 24888440]
- [84] Bhavaniramya S, et al. Role of essential oils in food safety: Antimicrobial and antioxidant applications. Grain & Oil Science and Technology 2019; 2(2): 49-55. [http://dx.doi.org/10.1016/j.gaost.2019.03.001]
- [85] Roohinejad S, et al. Extraction methods of essential oils from herbs and spices Essential Oils in Food Processing: Chemistry. Safety and Applications 2017; pp. 21-55. [http://dx.doi.org/10.1002/9781119149392.ch2]
- [86] Nguyen TA, Friedman AJ. Curcumin: a novel treatment for skin-related disorders. J Drugs Dermatol 2013; 12(10): 1131-7. [PMID: 24085048]
- [87] Gull I, Saeed M, Shaukat H, Aslam SM, Samra ZQ, Athar AM. Inhibitory effect of *Allium sativum* and *Zingiber officinale* extracts on clinically important drug resistant pathogenic bacteria. Ann Clin Microbiol Antimicrob 2012; 11(1): 8. [http://dx.doi.org/10.1186/1476-0711-11-8] [PMID: 22540232]
- [88] Kobayashi Y, et al. Inhibition of HIV-1 reverse transcriptase by methanol extracts of commercial herbs and spices. Nippon Shokuhin Kagaku Kogaku Kaishi=. Nippon Shokuhin Kagaku Kogaku Kaishi 2000; 47(8): 642-5. [http://dx.doi.org/10.3136/nskkk.47.642]
- [89] Umesh, *et al.* Identification of new anti-nCoV drug chemical compounds from Indian spices exploiting SARS-CoV-2 main protease as target. Journal of Biomolecular Structure and Dynamics 2020; 1-9.
- [90] Gutierrez J, Barry-Ryan C, Bourke P. Antimicrobial activity of plant essential oils using food model media: efficacy, synergistic potential and interactions with food components. Food Microbiol 2009; 26(2): 142-50. [http://dx.doi.org/10.1016/j.fm.2008.10.008] [PMID: 19171255]
- [91] Stoilova I, et al. Antioxidant activity of a ginger extract (Zingiber officinale). Food Chem 2007; 102(3): 764-70.
 [http://dx.doi.org/10.1016/j.foodchem.2006.06.023]
- [92] Rubió L, Motilva MJ, Romero MP. Recent advances in biologically active compounds in herbs and spices: a review of the most effective antioxidant and anti-inflammatory active principles. Crit Rev

Food Sci Nutr 2013; 53(9): 943-53. [http://dx.doi.org/10.1080/10408398.2011.574802] [PMID: 23768186]

- [93] Ciesarová Z, *et al.* Correlation between acrylamide contents and antioxidant capacities of spice extracts in a model potato matrix. J Food Nutr Res 2008; 47(1)
- [94] Przygodzka M, et al. Comparison of methods for evaluation of the antioxidant capacity and phenolic compounds in common spices. Lebensm Wiss Technol 2014; 58(2): 321-6. [http://dx.doi.org/10.1016/j.lwt.2013.09.019]
- Srinivasan K. Antioxidant potential of spices and their active constituents. Crit Rev Food Sci Nutr 2014; 54(3): 352-72.
 [http://dx.doi.org/10.1080/10408398.2011.585525] [PMID: 24188307]
- [96] Tajkarimi M, et al. Antimicrobial herb and spice compounds in food. Food Control 2010; 21(9): 1199-218.
 [http://dx.doi.org/10.1016/j.foodcont.2010.02.003]
- [97] Souza ELd, Stamford TLM, Lima EO. Sensitivity of spoiling and pathogen food-related bacteria to Origanum vulgare L.(Lamiaceae) essential oil. Braz J Microbiol 2006; 37(4): 527-32. [http://dx.doi.org/10.1590/S1517-83822006000400023]
- [98] Celikel N, Kavas G. Antimicrobial properties of some essential oils against some pathogenic microorganisms. Czech Republic: Czech Journal of Food Sciences-UZPI 2008. [http://dx.doi.org/10.17221/1603-CJFS]
- [99] Rasooli I. Food preservation-a biopreservative approach. Food 2007; 1(2): 111-36.
- [100] Carramiñana JJ, Rota C, Burillo J, Herrera A. Antibacterial efficiency of Spanish Satureja montana essential oil against Listeria monocytogenes among natural flora in minced pork. J Food Prot 2008; 71(3): 502-8.
 [http://dx.doi.org/10.4315/0362-028X-71.3.502] [PMID: 18389692]
- [101] Emiroğlu ZK, Yemiş GP, Coşkun BK, Candoğan K. Antimicrobial activity of soy edible films incorporated with thyme and oregano essential oils on fresh ground beef patties. Meat Sci 2010; 86(2): 283-8.
 [http://dx.doi.org/10.1016/j.meatsci.2010.04.016] [PMID: 20580990]
- [102] Rodriguez-Lafuente A, Nerin C, Batlle R. Active paraffin-based paper packaging for extending the shelf life of cherry tomatoes. J Agric Food Chem 2010; 58(11): 6780-6. [http://dx.doi.org/10.1021/jf100728n] [PMID: 20476770]
- [103] Ramos M, *et al.* Carvacrol and thymol for fresh food packaging. Вестник Казанского технологического университета 2013; 16(3)
- [104] Ramos M, et al. Active Packaging Based on the Release of Carvacrol and Thymol for Fresh Food. Chemistry and Physics of Complex Materials: Concepts and Applications. 2013; p. 1.
- [105] Gutiérrez L, Escudero A, Batlle R, Nerín C. Effect of mixed antimicrobial agents and flavors in active packaging films. J Agric Food Chem 2009; 57(18): 8564-71. [http://dx.doi.org/10.1021/jf901459e] [PMID: 19711918]
- [106] Yazdani D, Mior Ahmad ZA, Yee How T, Jaganath IB, Shahnazi S. Inhibition of aflatoxin biosynthesis in Aspergillus flavus by phenolic compounds extracted of Piper betle L. Iran J Microbiol 2013; 5(4): 428-33. [PMID: 25848517]
- [107] Sindhu S, Chempakam B, Leela NK, Suseela Bhai R. Chemoprevention by essential oil of turmeric leaves (*Curcuma longa* L.) on the growth of Aspergillus flavus and aflatoxin production. Food Chem Toxicol 2011; 49(5): 1188-92.
 [http://dx.doi.org/10.1016/j.fct.2011.02.014] [PMID: 21354246]
- [108] Du W-X, et al. Antimicrobial volatile essential oils in edible films for food safety. Science against

microbial pathogens: communicating current research and technological advances 2011; 2: 1124-34.

- [109] Otoni CG, et al. Trends in antimicrobial food packaging systems: Emitting sachets and absorbent pads. Food Res Int 2016; 83: 60-73. [http://dx.doi.org/10.1016/j.foodres.2016.02.018]
- [110] Jayasena DD, Jo C. Essential oils as potential antimicrobial agents in meat and meat products: A review. Trends Food Sci Technol 2013; 34(2): 96-108. [http://dx.doi.org/10.1016/j.tifs.2013.09.002]

Spices and Herbs in the Treatment of Neurobiological Disorders

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Abstract: Spices and herbs have been used for centuries for treating and preventing many ailments. They have been popularised over the commercial and new drugs due to their purported therapeutic efficacy with lesser side effects, easy availability, and costefficiency. Herbal extracts contain mixtures of phytochemicals, mainly secondary metabolites, which include fatty acids, sterols, alkaloids, flavonoids, glycosides, saponins, tannins, terpenes, and many others. Phytochemicals play a vital role in maintaining chemical balance in the brain and, therefore, can be targeted to treat neural disorders. In recent years, many herbs and spices have gained attention in the treatment of neurological disorders. Although the precise mechanisms of action of herbal medicines have not yet been defined, some of them have been shown to exert antiinflammatory and antioxidant activities. Several herbs and spices have also shown neuroprotective activity, and their extracts have been found to be effective in learning and memory improvement, depression, anxiety, pain, Alzheimer's disease and other neurodegenerative conditions. Based on the investigations on herbal plants and neurological substrates in disease conditions, herbal medicines can be effectively used in the treatment of various neurological disorders.

Keywords: Antioxidant, Depression, Herbs, Neurodegeneration, Reactive oxygen species, Spices.

INTRODUCTION

For centuries, spices and herbs have been used as food adjuncts for various purposes such as seasoning, flavouring, colouring, and sometimes as a preservative. Aside from food ingredients, they have also been used as nutritional supplements as they play a crucial role as supplementary, complementary, and synergistic components [1]. The ancient Indian system of medicine used a large number of herbs and spices, alone or in combination, for treating and/or preven-

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ting many ailments. They contain a number of biologically active constituents that are responsible for their biological effects. The heterogeneous collection of compounds present in spices and herbs inarguably contributes to their therapeutic usage. Natural products obtained from them in pure form or as extracts have been considered to possess medicinal value and have been used in the indigenous systems of medicine [2].

Dietary herbs and spices have been proven to be beneficial for human health through a number of actions, such as antioxidative, chemopreventive, antimutagenic, anti-inflammatory, antibacterial, and immunomodulatory on gastrointestinal, cardiovascular, respiratory, metabolic, reproductive, neural, and other systems of body [1]. Their extracts had been used for a long to cure various disorders, such as cough, bronchitis, laryngitis, tonsillitis, spasmodic, gastricintestinal complaints, carminative, and as diuretic agents. Also, the topical preparations of these extracts were used in the treatment of wounds and disorders of the oral cavity. Herbs and spices exert protective effects in various chronic conditions, including diabetes, cancer, and cardiovascular disease, but the exact mechanisms underlying their action are, however, not very clear.

Today, a growing number of people worldwide have brain disease or disorder such as Alzheimer's disease, Parkinson's disease, depression, schizophrenia, and addiction, which affects the quality of daily life through abnormal behaviors, thoughts, emotions, and social communication. Despite exhaustive research, the aetiologies of these ailments remain poorly understood. Although multiple factors are responsible for the development of neurological diseases, dysregulation in the inflammatory mediators, oxidative imbalance, excitotoxicity, and loss of protective mechanism are key components in the pathogenesis of various neurological conditions such as neuropsychiatric and/or neurodegenerative disorders [3 - 7]. Therefore, for targeted treatment, the agents should be pharmacologically safe, cost-effective, and immediately available with minimal side effects. But one of the major disadvantages of the current treatments for neurological disease with synthetic drugs is that they are associated with multiple side effects.

Extensive research on herbs and spices has taken place, which could provide a new alternative therapeutic approach for diseases of the central nervous system [8]. Generally, herbal extracts contain a number of bioactive phytochemicals that have a wide range of actions, including antioxidant, anti-inflammatory, and neuroprotective, thus use of herbal medicines or natural products was found to be the most commonly used alternative treatment because they can easily cross the blood-brain barrier, exert multiple synergistic effects due to a number of constituents and show less toxicity [9].

Recently, herbs and spices have received attention in their useful physiological functions. More research is required on the usefulness of herbs and spices in disorders related to the brain. The present chapter deals with the involvement of different constituents of herbs and spices in the treatment of neurological disorder and their possible mechanism.

BIOACTIVE PHYTOCHEMICALS IN HERBS AND SPICES:

The biological activities of herbs and spices have been related to the presence of different phytochemicals. The herbal extract contains mixtures of phytochemicals, which are mainly secondary metabolites including fatty acids, polyphenols (phenolic acids, anthocyanins, proanthocyanidins, flavonols, and tannins), isoprenoids (sesquiterpenes, diterpenes, triterpenes, steroids, and saponins), alkaloids, glycosides, and so forth. These phytochemicals are currently used in the pharmaceutical industry for various purposes as they can regulate a variety of enzymes as well as cell receptors [10].

Polyphenols

Polyphenols are a group of plant secondary metabolites that contain phenols, anthocyanins, proanthocyanidins, flavonols, and tannins and are characterized by the presence of phenolic hydroxyl group which is directly linked to the aromatic ring. There has been a growing interest in the beneficial effects of active polyphenols derived from spices and herbs endowed with potent antioxidative and anti-inflammatory activity [11]. Polyphenols have the property to reduce free radical species, which contributes to their neuroprotective effects. Apart from the antioxidant action, this category of compounds can also alter several signaling pathways by acting on a variety of molecular targets [12]. Also, the polyphenolic substance has the potential to inhibit lipid peroxidation, neutralize Reactive Oxygen Species (ROS) and NO-based free radicals (nitric oxide and peroxynitrite) [13, 14].

Flavonoids form the most important group of polyphenols. Numerous studies show that flavanols have beneficial effects on neuronal health. Most common flavonoids include flavones (*e.g.* apigenin, luteolin,), flavanones (*e.g.* hesperetin,), catechins [*e.g.* epicatechin, epigallocatechin-3-gallate (EGCG)], and anthocyanins. They are found in various spices and herbs like oregano, thyme, parsley, coriander, celery, dill weed, onions, spinach, and rosemary [15]. Catechin, epicatechin, and epicatechin gallate found in cumin and cinnamon have been shown to reduce neuroinflammation, protect the brain against injuries produced by neurotoxins and also attenuate the apoptotic mediators of neurons [16, 17]. In addition, they also delay the onset of neurodegenerative disorders *via* iron chelation, radical scavengers, and modulation of prosurvival genes [16]. Chinese medicinal herb

Epimedium revicornum Maxim contains Icariin, a major constituent of flavonoids, is found to improve memory in rats after common carotid artery occlusion [18]. Quercetin has been found in rosemary, oregano, sage, bay, and thyme and is shown to protect against calcium dysregulation during an ischemic injury in neuronal cell death and brain damage. It is also found to be beneficial in attenuating protein oxidation and apoptosis in the hippocampus [19 - 21].

Phenols and flavonoids have antioxidant properties, which may be attributed to their molecular weight, presence of conjugated aromatic rings, and hydroxyl groups, which have the potential to scavenge the free radicals involved in oxidative processes through hydrogenation and complexation with oxidizing species and thus resist oxidative stress [22].

Table 1 enlists herbs and spices including thyme, oregano, rosemary, curry leaves, sage, nutmeg, mace, clove, allspice, ginger, and turmeric which contain antioxidant properties [23]. Anthocyanins have been found to act against oxidative stress, lipid peroxidation and exhibit neuroprotective action in improving cognitive brain function [24].

Common Name	Family	Scientific Name	Total Phenolic Content (g of gae/ 100 g of Dried wt. or Fresh wt.)	Phenolic Compounds
Oregano	Labiatae	Origanum vulgare L.	10.17 ± 0.010^{a}	Phenolic acids (caffeic acid, p-coumaric acid, rosmarinic acid, caffeoyl derivatives), volatile compounds (carvacrol), flavonoids
Rosemary	Labiatae	Rosmarinus officinalis L.	5.07 ± 0.036^{a}	Phenolic acids (caffeic acid, rosmarinic acid, caffeoyl derivatives), phenolic diterpenes (carnosic acid, carnosol, epirosmanol), volatile compounds (carvacrol), flavonoids
Sage	Labiatae	Salvia officinalis L.	5.32 ± 0.006^{a}	Phenolic acids (rosmarinic acid), phenolic diterpenes (carnosic acid), volatile compounds, flavonoids

Table 1. Major phenolic compounds found in herbs	s and spices.
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able 1) cont Common Name	Family	Scientific Name	Total Phenolic Content (g	Phenolic Compounds	
			of gae/ 100 g of Dried wt. or Fresh wt.)		
Thyme	Labiatae	Thymus vulgaris L.	4.52 ± 0.006^{a}	Phenolic acids (gallic acid, caffeic acid rosmarinic acid), volatile compounds (thymol), phenolic diterpenes, flavonoids	
Cinnamon	Lauraceae	Cinnamomum cassia Presl	6.34 ± 0.021^{a}	Phenolic acids, phenolic volatile oils (2 hydroxycinnamaldehyde, cinnamyl aldehyde derivatives), flavan 3-ols	
Nutmeg	Myristicaceae	Myristica fragrans Houtt.	$1.61\pm0.001^{\text{a}}$	Phenolic volatile oils, phenolic acid (caffeic acid), flavanols (catechin)	
Clove	Myrtaceae	Eugenia caryophylata Thunb.	$\begin{array}{c} 14.38 \pm \\ 0.006^a \end{array}$	Phenolic acids (gallic acid), flavonol glucosides, phenolic volatile oils (eugenol, acetyl eugenol), tannins	
Black and white Pepper	Piperaceae	Piper nigrum L.	$\begin{array}{c} 0.30 \pm 0.002^{a} \\ 0.78 \pm 0.004^{a} \end{array}$	Volatile oils, phenolic amides	
Dill	Umbelliferae	Anethum graveolens L.	0.98 ± 0.009^{a}	Phenolic acids (protocatechuic acid), flavonoids (catechin), volatile oils	
Caraway	Umbelliferae	Carum carvi L.	0.61 ± 0.017^{a}	Volatile oils, phenolic acids, flavonoids (kaempferol), coumarins	
Coriander	Umbelliferae	Coriandrum sativum L.	$0.88\pm0.007^{\text{a}}$	Phenolic acids (caffeic acid), flavonoids, volatile oils	
Cumin	Umbelliferae	Cuminum cyminum L.	0.23 ± 0.005^{a}	Volatile oils, phenolic acids, flavonoids (kaempferol), coumarins	
Parsley	Umbelliferae	Petroselinum crispum L.	$0.97\pm0.002^{\rm a}$	Phenolic acids (caffeic acid), flavonoids, volatile oils	
Ginger	Zingiberaceae	Zingiber officinale Rosc.	0.63 ± 0.009^{a}	Phenolic volatile oils (gingerol, shogaol), phenolic acids	
Cardamom	Zingiberaceae	<i>Elettaria car-</i> <i>damomum</i> Maton.	$0.46\pm0.009^{\mathtt{a}}$	Phenolic acids (caffeic acid), volatile oils	

a-Total phenolic content expressed as g of gallic acid (GAE)/100 g of dry weight (DW)(17), b- Total phenolic content expressed as g of gallic acid (GAE)/100 g of fresh weight (FW) [25].

Terpenoids

Terpenoids, also known as isoprenoids, consist of a very large group of two or more branched 5 carbon units synthesized from precursor mevalonic acid. Depending upon the number of isoprene units, they are subdivided into different classes: monoterpenes, sesquiterpenes, diterpenes, triterpenes, and steroids [26]. Herbs and spices mainly contain monoterpenes and diterpenes. These compounds are lipophilic, thus can cross the blood-brain barrier.

The representative molecules among these are monoterpenes formed from the coupling of two isoprene units (C10). These compounds are responsible for the antioxidant and anti-inflammatory, anxiolytic, and anticonvulsant activity of herbs and spices. The presence of phenolic monoterpenes in thyme, thymol, and oregano are identified as the dominant compounds for antioxidant activity [27]. Also, the presence of thymol in oregano showed anti-inflammatory effects by decreasing the levels of proinflammatory cytokines like TNF- α , IL-1b, and IL-6, as well as increasing the production of the anti-inflammatory cytokine IL-10 [27]. Many of the monoterpenes possess its Anti- Cholinesterase activity due to the presence of a hydrocarbon skeleton. The 1,8-cineole and a-pinene, cyclic monoterpenes present in S. lavandulaefolia oil inhibit the striatum and the hippocampus Cholinesterase (ChE) [27].

The Sesquiterpene class of terpenes consists of three isoprene units (C15). Recent reports show that sesquiterpenes also had beneficial effects on neurological conditions. *Ginkgo biloba* contains sesquiterpene, bilobalide is extensively used as an anticonvulsant and cognitive enhancement. Valerian root and rhizome contain sesquiterpenes, valerenic acid, which has beneficial effects in a number of neurological disorders such as epilepsy, insomnia, dizziness, and anxiety. Atractylenolide, isolated from *Atractylodes macrocephala*, is used to treat sleep disorders, as these sesquiterpenes enhance the action of GABA [28].

Diterpenes are formed from the coupling of four isoprene units (C20) and are responsible for the antioxidant activity of many herbs. Carnosic acid, carnosol and rosmarinic acid are antioxidant diterpenes present in aromatic herbs [29]. Ginkgolide in *Ginkgo biloba* leaf and Zerumin isolated from *Curcuma kwangsiensis* rhizomes have GABAA modulatory action [30]. Terpenoids responsible for the biological activities in herbs and spices are shown in Table **2** [29, 30].

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Isoprene Class	Terpenoids	Plant Source	Mechanism	Action
Monoterpenes	Borneol	Rosemary type B	Modulation of GABA receptors	Anticonvulsant
	Carvacrol	Caraway, spearmint, and dill	GABAA Modulation Anti-inflammatory Antioxidant	Anxiety Anticonvulsant Anti-inflammatory
	Epinepetalactone	Nepeta sibthorpii	Agonists on the BZD binding site on GABAA receptors, increasing chloride conductance.	Anticonvulsant
	Isopulegol	Zanthoxylum schinifolium	Activate the GABAA–BZD site	Anxiety Depressant Anticonvulsant
	Menthol	Peppermint and thyme	Suppresses excitability via increasing GABAA receptor- mediated inhibition of hippocampal neurons The modulator of the GABAA receptor	Anticonvulsant
	Thujone	Oregano, sage	Inhibition of the GABAergic system	Convulsion Memory enhancement
	Thymol	Oregano	Positive allosteric modulator of GABAA receptor Antioxidant	Anticonvulsant
	Cineole	Coriander, lavender, rosemary, sage, and thyme	Opioid system Interact with excitatory amino acid	Anticonvulsant
	A-pinene	Caraway, coriander, fennel, juniper berry, rosemary, And thyme	Anti-ache activity	Dementia Alzheimer's disease
Sesquiterpene	Valerenic acid	Valerian (Valeriana officinalis)	an allosteric modulator of GABAA receptors	Anxiolytic Sleep disorder
	Bilobalide	Ginkgo biloba	GABA antagonist	Convulsant Cognition enhancement
	Atractylenolides	Atractylodes macrocephala	potentiate GABA-induced chloride currents	Anticonvulsant

Table 2. Terpenoids in herbs and spices that are responsible for the biological activities of herbs and spices [29, 30].

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Isoprene Class	Terpenoids	Plant Source	Mechanism	Action
Diterpenes	Ginkgolides	Ginkgo biloba	GABA antagonist	Not defined
	Zerumin A	Curcuma kwangsiensis	positive GABAA receptor modululator	Not defined
	Rosmanol	Rosemary, sage	Anti-inflammatory-Inhibits the activation of NF-kB and STAT3	Inflammation

Alkaloids

Alkaloids are naturally occurring nitrogen-containing compounds, usually divided into different classes based on their sources and chemical structures [31, 32]. They are mainly found in certain families of flowering plants such as Solanaceae (nightshades), Papaveraceae (poppies family), Ranunculaceae (buttercups), and Amaryllidaceae (amaryllis) [33]. Alkaloid-containing extracts have been used for the treatment of a variety of ailments.

Capsaicin is the primary capsaicinoid alkaloid found in chili and peppers, which exhibits antioxidant and anti-inflammatory properties. It also ameliorates synaptic damage and tau hyperphosphorylation in stressed mice [34]. Tobacco pyridine alkaloid nicotine belonging to the Solanaceae family found in *Nicotiana tobacum* has been extensively investigated as it enhances cholinergic function [35 - 37].

Berberine is an isoquinoline alkaloid isolated from Chinese herb and several plants, including *Hydrastis canadensis* (Goldenseal), *Berberis vulgaris* (barberry), *Coptis chinensis* (copies or golden thread), and *Berberis aristata* (tree turmeric). It has multiple pharmacological effects like anti-inflammatory, antioxidant, anxiolytic, antidepressant, and anti-amnesic and also has potential for the treatment of drug addiction [36]. It also improves memory by enhancing neurogenesis and inhibiting apoptosis in the hippocampal dentate gyrus [38].

Also, Piperine alkaloid, the main chemical constituents of long pepper (*Piper longum*) and black pepper (*Piper nigrum*), have demonstrated the beneficial effects of piperine which include anti-inflammatory effects, analgesic effect, anticonvulsant, antidepressant effect, cognitive enhancing effect, cytoprotective effect, and antioxidant activity. Piperine also significantly attenuates the memory impairment, the elevation in AChE activity, and neurodegeneration [39]. In combination with curcumin, piperine inhibits the action of Monoamine Oxidase (MAO) enzymes, thus increasing serotonin and dopamine levels which is found to be beneficial for the treatment of depression [40].

Harmine is an indole β -carboline, characterized by indole structure and a pyridine ring, found in *Peganum harmala* belonging to Nitrariaceae family. It has a wide spectrum of activities, including antioxidant, antimicrobial, anti-inflammatory, and neuroprotective activity through various targets AChE, MAO-A, MAO-B, and tyrosine phosphorylation regulated kinase (DYRK1A) inhibition [36]. Harmine is also found to reduce tau protein phosphorylation and thus can be beneficial in the treatment of Alzheimer's Disease (AD) [41].

Some of the alkaloids also produce toxic effects. Pyrrolizidine alkaloids and their N-oxides (PANO) are predominantly present as contaminants in herbal food supplements, as well as in spices. Culinary herbs like oregano and cumin also contain high amounts of PA/PANO [42]. Commonly used alkaloids in herbs and spices for neurological treatment are shown in Table **3**.

Class of Alkaloid	Plant Source	Alkaloid	Mechanism	Disease	References
Isoquinoline alkaloids	Hydrastis Canadensis, Coptis Chinensis, Berberis Aquifolium, Berberis vulgaris, Berberis aristata	Berberine	Anti-oxidant, MAO inhibitor, anti-amyloid	AD, PD, HD, and Epilepsy	[32, 38]
	Papaver somniferum (opium Poppy)	Morphine	Neuroprotective against oxidative stress	AD	[43, 44]
	Hippeastrum vittatum	Montanine	Enhance GABAnergic Neurotransmission	Epilepsy	[45]
Piperidine alkaloids	Black pepper (Piper nigrum) and Long pepper (Piper longum)	Piperine	MAO inhibitor	AD, PD, Epilepsy	[39, 40]
Pyridine alkaloids	Nicotiana tobacum	Nicotine	Nicotinic agonist, anti-amyloid	AD, PD	[35 - 37]
Indole β- carboline	Peganum harmala	Harmine	COMT inhibitor, Tyrosinephosphorylation regulated kinase (DYRK1A) inhibition, MAO inhibitor	AD	[32, 41]

Table 3. Commonly used alkaloids in herbs and spices for neurological	treatment.

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Class of Alkaloid	Plant Source	Alkaloid	Mechanism	Disease	References
Capsaicinoid alkaloid	Capsicum annuum (red pepper)	Capsaicin	Anti-amyloid	AD	[34]

REGULATORY MECHANISM UNDERLYING TREATMENT

Today neurological diseases represent a severe health problem, as the prevalence of neurological conditions, including neurodegenerative and psychiatric disorders, are steadily increasing. The functional methods of treatment should target the major causes underlying the diseases. The major etiological targets for these disorders are oxidative stress, neuroinflammation, and dysregulation in the neurotransmitter system. Though synthetic medicine can show beneficial effects by targeting the etiological factors, one of the major disadvantages of the current treatments for neurological disease with synthetic drugs is that they are associated with multiple side effects.

Possible Mechanism of Herbs and Spices in Neurological Disorders

Herbs and spices have been recognized to have advantageous effects on the body. In recent years, these have garnered special attention in the treatment of neurological degeneration, for their broad spectrum of molecular and cellular actions. Although the precise mechanisms of action of many phytochemicals in herbs and spices are yet to be defined, some of them have been shown to exert anti-inflammatory and antioxidant effects. Also, herbs and spices have neuroprotective activity and their extracts have been found to be effective in learning and memory improvement, antidepressant, anxiolytic, antipain, Alzheimer's, and other neurodegenerative conditions. The possible mechanism of spices and herbs in neurological disorders is shown in Fig. (1).

Antioxidant

Among the theories proposed to explain the mechanisms of pathogenesis at the molecular level, oxidative stress or imbalance is the key component in brainrelated diseases. Oxidative stress can be defined as an imbalance in the cell oxidation/reduction (redox) status that results in the production of partially reduced oxygen intermediates termed as reactive oxygen species (ROS), which are more reactive than molecular oxygen in their ground state. Numerous pieces of evidence suggest that reactive oxygen species (ROS) play an important role in neurological and psychiatric disorders [46, 47].

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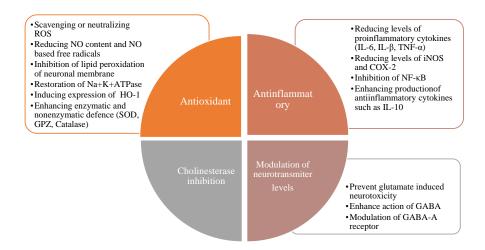


Fig. (1). Possible mechanism of spices and herbs in neurological disorders.

Although the brain weighs less than 2% of the body, it consumes approximately 20% of the oxygen, thus due to its high oxygen demand, the brain is most susceptible to oxidative damage [48]. Aerobic organisms produce ROS following the use of molecular oxygen during oxidative phosphorylation in mitochondria. However, the excessive production of ROS can pose serious neurological damage to biological targets such as DNA, lipids, and proteins, thereby altering cellular responses and pathways. In addition to this, a high amount of polyunsaturated fatty acids (PUFAs) are present in neuronal membranes, which makes the brain tissues more susceptible to lipid peroxidation; resulting in the production of cytotoxic aldehydes, such as malondialdehyde (MDA) and 4-hydroxynonenal (HNE) [49]. Natural antioxidantdefence comprises of Superoxide Dismutase (SODs), catalase, and Glutathione (GSH), which can quench free radicals, and thus counteract the imbalance in the redox homeostasis and keep the ROS levels under the threshold thereby preventing oxidative damage [50].

Therefore, treatment with the appropriate antioxidant agent can show beneficial effects. Unfortunately, with the risks and unwanted side effects associated with drug therapies, natural alternatives are potential solutions to alleviate the condition. The phytochemical constituents in herbs and spices, mainly responsible for the antioxidant activity, are polyphenols. In several studies, herbs and spices including clove, ginger, cinnamon, turmeric, black pepper, cumin, oregano, sage, peppermint, thyme, rosemary, dill, parsley, basil, cinnamon, and saffron, were reported to have the greatest antioxidant capacity [29, 51 - 53].

All of these compounds appear to act on different molecular targets, affecting several signaling pathways. The major mechanisms by which these herbs and spices show antioxidant properties are: scavenging ROS to protect neurons from oxidative damage through hydrogenation and complexation with oxidizing species and thus resist oxidative stress [22, 54]; neutralizing ROS and NO-based free radicals [13, 14]; inhibiting lipid peroxidation of neuronal membranes (54); enhancing enzymatic and nonenzymatic antioxidant defence such as, SOD, Glutathione Peroxidase (GPx), Catalase (CAT), in the brain [55]; inducing HO-1 expression thereby increasing resistance to oxidative injury [56]; reducing nitric oxide content and restoration of Na⁺-K⁺ ATPase [57].

Anti-inflammatory

Neuroinflammation is a prominent pathological feature of neurological disorder, characterized by activated microglia and infiltrating T-lymphocytes at sites of neuronal injury [58]. Injury to the neurons stimulates the microglia, a member of the innate immune system, which further activates other glia such as astrocytes to coordinate a collective response to neuronal injury. In response to alterations induced in the innate immune system, T-lymphocytes, members of the adaptive immune system, infiltrate the CNS at sites of neuronal injury. Thus, active participation of inflammation in neurological disease pathogenesis, as well as its contribution to neurodegenerative pathology and tissue destruction is evident [59, 60].

Like the macrophages, microglia can cause a transition from protective antiinflammatory to a cytotoxic proinflammatory state, thus offering a potential pathway for neurodegeneration [61]. Thus, in response to injury, proinflammatory response mediated by microglia secretes potent ROS such as superoxide radicals and nitric oxide, proinflammatory cytokines such as tumor necrosis factor- α (TNF- α), IL-6, and IL-1 β and also reduces the release of neurotrophic factor. Other inflammatory mediators include the chemokine macrophage inflammatory protein-1 α (MIP-1 α), interferon- γ (IFN- γ), and compounds such as lipopolysaccharide (LPS) [61].

While the mechanisms behind the health benefits from spices and herbs are becoming clearer over time, the vast majority of spices and herbs are found to act *via* its anti-inflammatory effect. The most frequently identified spices that possess anti-inflammatory effects are thyme, oregano, rosemary, sage, basil, mint, turmeric, dill, parsley, cinnamon, clove, nutmeg, lemongrass, ginger, chili pepper, fenugreek, and pepper [29].

The reported mechanism behind the anti-inflammatory effects of herbs and spices in the presence of compounds such as curcumin, gingerol, and capsaicin, which appear to operate by inhibiting one or more of the pro-inflammatory mediators responsible for neurotoxicity [62].

A diet rich in herbs and spices may contribute to the reduction of inflammatory mediators. Several herbs and spices also exhibited anti-inflammatory activity by increasing anti-inflammatory IL-10 production [63]. Quercetin (found in basil, cumin, and fennel), ursolic acid (found in basil and rosemary), gingerol (found in ginger), capsaicin, and curcumin, are found to inhibit the NF– κ B pathway [64]. Curcumin is also reported to significantly decrease the levels of proinflammatory cytokine IL-1 β of oxidized proteins of A β peptide and it also inhibits microglia activation [65, 66]. The highest anti-inflammatory activity has been detected in chili pepper, which enhances the IL-10, IL-6 levels and reduces TNF-a secretion, iNOS expression. The chili pepper compound, capsaicin also modulates NF-jB-and IL-8 pathways [67, 68]. Black pepper also strongly inhibits IL-6 production and the expression of iNOS. Nutmeg extract was reported to be the most potent inhibitor of TNF-a, IL-6, and IL-10 production and COX-2 expression. According to the literature, macelignan, a constituent of nutmeg is found to show anti-inflammatory activity in the hippocampus and primary microglial cells [69].

Cholinesterase Inhibition

In the central nervous system, acetylcholine (Ach) is involved in various physiological functions such as learning, memory, and mood. But once released in the synaptic cleft, it is cleaved by the enzyme acetylcholinesterase (AChE) thereby terminating the synaptic activity (Soreq and Seidman 2001). Inhibitors of AChE have become an important strategy to treat cognitive impairment, orientation, comprehension, learning, thinking, and judgment in AD and PD [70].

The cholinergic hypothesis is the basis for the development of treatment approaches designed to facilitate the survival of the cholinergic system. For this purpose, cholinesterase inhibition (ChEI) has proven to be a preferable therapy, as it amplifies the action of ACh. Several ChEI's have been approved by the Food and Drug Administration (FDA) for the symptomatic treatment of AD. However, these drugs have limitations due to unfavorable side effects. Hence, more studies must be conducted to look for better alternatives for ChEIs from natural sources [71].

The phytochemicals present in herbs and spices such as terpenoids, flavonoids, and steroidal alkaloids have been reported to possess potent ChE inhibitory activity. In in-vitro and in-vivo studies of *Ferula asafoetida* have been reported to have ChE inhibiting activity on the snail nervous system [72]. Two of the main

rosemary constituents, rosmarinic and carnosic acids, were found to exhibit promising AChE inhibitory activity for preventing AD [70]. Berberine has also demonstrated acetylcholinesterase-inhibiting property [72].

Modulation of Amino Acid Neurotransmitter System (GABA)

The brain neurotransmitter system plays an important role in neurological disorders. The principal inhibitory neurotransmitter is gamma-aminobutyric acid (GABA), while the principle excitatory neurotransmitter is glutamate [72]. Coordination between these two neurotransmitters ensures adequate synaptic excitation/inhibition and neural oscillation. Thus dysregulation in the neurotransmitter system has been implicated in a number of neurological and psychiatric diseases including epilepsy and schizophrenia [72]. Several different categories of drugs modulate GABAergic synapses and are used for the treatment of anxiety and sleep disorders, epilepsy, alcohol withdrawal, and induction and maintenance of anesthesia [73]. Also, glutamate dysfunction has been correlated with neurological disorders, such as Alzheimer's disease, schizophrenia, pain disorders, drug addiction [73].

Recent studies have reported that phytoconstituents in herbs and spices act on the neurotransmitter system, indicating theirrole in positive correlation or the treatment of neurological disorders. Gambogic acid present in Kokum spice is found to prevent glutamate-induced neuronal cell death and neurite outgrowth in PC12 cells [74]. Atractylenolide, isolated from *Atractylodes macrocephala*, herb is used to treat sleep disorders, as this sesquiterpene enhance the action of GABA (28). The essential oil of thyme has been suggested to have a neuroprotective effect against toxicity *via* facilitating GABA action and modulates the GABAA receptor [75 - 77]. Flavonoids present in garlic extract are found to have a protective effect on primary neurons from glutamate toxicity and oxidative injuries [74].

SPICES AND HERBS USED IN MAJOR NEUROLOGICAL DISORDERS

Alzheimer's Disease (AD)

Amongst a variety of neurodegenerative diseases, Alzheimer's disease is the most devastating disorder which typically appears after age 60 and is linked to a specific genetic defect. Clinically AD is characterized by progressive and irreversible memory deficits. Memory impairment is the hallmark of disease at the early stage and is followed by a later stage that involves motor and sensory dysfunction. The diagnosis of the disease is based on the presence of one or more cognitive deficits including the impaired ability for motor activities, language impairment, failure to recognize or identify objects, and impairment in executive

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functioning [78]. Over the decade, researchers have tried to discover the pathophysiology and risk factors responsible for AD and possible mechanisms leading to the formation of plaques and tangles in the brain. The primary mechanism underlying the neurodegeneration in AD is attributed to the production and accumulation of β -amyloid (A β) peptides in the specific region in the brain [79]. In addition, other etiological cause for disease progression includes oxidative stress manifested by lipid peroxidation, ROS formation, protein oxidation, nitrotyrosine, and DNA/RNA oxidation [80, 81]. The major risk factor for AD is aging, which leads to loss of free radical scavenging ability by the endogenous defence system [29]. Hence, these contribute to a disturbance in the normal balance between free radical generation and free radical scavenging [78].

Current conventional treatment approved by FDA includes acetylcholinesterase inhibitors (AChEIs) (rivastigmine, galantamine, donepezil) and N-methyl Daspartate (NMDA) receptor antagonist (memantine) [78]. Complex pathophysiological mechanisms of AD provide insight for a potential new therapeutic compound. In recent years, the therapeutic potentials of alternative herbal drugs have become popular in the treatment of many ailments. It has been also implicated for symptomatic treatment of AD based on the mechanisms of action and therapeutic targets of herbal drugs. The therapeutic strategies for the treatment of AD using herbs and spices include anti-amyloid action, β - and γ secretase inhibition, inhibition of Tau hyperphosphorylation, antioxidant and antiapoptotic effect, anti-inflammatory effect, anticholinesterase activity, *etc.* The possible mechanism of herbs and spices in Alzheimer's disease is shown in Fig. (2).

Researchers have also evaluated several constituents in herbs and spices for their anti-amyloidogenic activities, including ellagic acid, garlic acid, ginger, mulberry leaf extract [39]. Recent research has also reported that curcumin improves memory and synaptic functions in animal models of AD [82]. Further, it reduces the levels of oxidized proteins and IL1B in brains [66]. It inhibits peroxidase and binds to redox-active metals, iron, and copper, and suppresses inflammatory damage by preventing metal induction of nuclear transcription factor- κB (NF κB) [83 - 85]. Extract of Ginkgo biloba has been widely used in memory impairment *via* multiple cellular and molecular neuroprotective mechanisms; which includes attenuation of apoptosis, the inhibition of membrane lipid peroxidation, antiinflammatory effects, and the direct inhibition of A β aggregation [39]. The phenolic and flavonoid contents in Atriplex laciniata possess significant antioxidant and anticholinesterase effects, which are effective in the treatment of AD and other neurological disorders [86]. Garlic extract, Rosmarinic acid in sage reduces amyloid-beta induced apoptosis in PC12 cells [87, 88]. Turmeric, ginger, cinnamon, and curcumin extracts have been found to successfully block amyloid-

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beta aggregation and thus, are effective in the treatment of AD [89, 90]. Ursolic acid in basil and linalool in coriander is found to treat AD by inhibiting acetylcholinesterase activity [91].

Piperine present in Black pepper improves memory impairment and neurodegeneration in AD [39]. A list of commonly used herbs and spices in the treatment of AD is shown in Table **4** [91, 92].

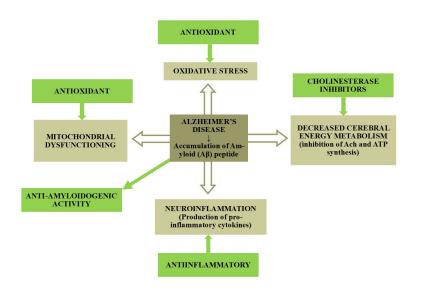


Fig. (2). Possible mechanism of herbs and spices in Alzheimer's disease.

Herbal Drug	Biological Source	Family	Possible Mechanism	References
Ginkgo	Ginkgo biloba	Ginkgoaceae	Antioxidant, Free radical scavengers for free radicals, inhibits lipid peroxidation	[93]
Sage	Salvia officinalis	Lamiaceae	Antioxidant and Acetylcholinesterase- inhibitory effect	[94]
Rosemarry	Rosmarinus officinalis	Lamiaceae	Antioxidant, inhibitors of lipid peroxidation	[95]
Turmeric	Curcuma longa	Zingiberaceae	Inhibition of NFkB and amyloid-beta aggregation	[96]

Table 4. List of commonly used herbs and spices in treatment of AD [91, 92].

Neurobiological Disorders

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Herbal Drug	Biological Source	Family	Possible Mechanism	References
Ginseng	Panax ginseng	Araliaceae	inhibition of amyloid-beta aggregation and enhancement of the removal of Aβ from the neurons, interruption of tau hyperphosphorylation	[97]
Ginger	Zingiber officinale	Zingiberaceae	inhibition of amyloid-beta aggregation	[98]
Basil	Ocimum basilicum	Lamiaceae	Inhibited acetylcholinesterase	[99]
Coriander	Coriandrum sativum	Apiaceae	Inhibited acetylcholinesterase in vitro	[100]
Sage	Salvia officinalis	Lamiaceae	Protected PC12 cells from Abeta-induced neurotoxicity	[101]

Parkinson's Disease (PD)

Parkinson's disease is the second most common neurodegenerative disease that can impair quality of life. It is a chronic and progressive movement disorder characterized by muscular rigidity, tremor, and bradykinesia [3]. The etiology of PD is not clear, but studies have implicated that oxidative stress from exogenous stressors or endogenous neurotoxins can cause dysfunction in the dopaminergic system. The disease is triggered by the reduction of dopaminergic neurons in the substantia nigra pars compacta, along with the presence of Lewy bodies within dopaminergic neuronal populations [102]. Apart from dopaminergic dysfunctioning, chronic neuroinflammation is considered a pathological hallmark of PD. Chronic release of pro-inflammatory cytokines by activated astrocytes and microglia leads to the expression of pro-inflammatory mediators such as TNF- α , IL-1 β , IL-6, and interferon- γ , which leads to the worsening of neuronal degeneration in the SNpc [102].

Most of the current treatment approaches in PD are aimed to replenish the striatal dopamine levels to provide symptomatic relief during the early stage but do not effectively tackle tremor, postural instability, and cognitive deficits. These drugs do not exhibit neuroprotective effects in PD. Thus, novel therapies involving natural antioxidants and plant products with neuroprotective properties are currently being explored as adjunct therapy.

Curcumin is a polyphenol component of *Curcuma longa* that exhibits antioxidant and anti-inflammatory properties. It is also reported to cross the blood-brain barrier and shows neuroprotective effect in neurological disorders. Moreover, studies in different experimental models of PD suggested the clinical application

of curcumin in PD [103]. It also reduces synuclein toxicity, intracellular ROS, and apoptosis in neuroblastoma cells in PD [104].

Zingerone in ginger and eugenol in clove have been reported to prevent 6-OHDA induced dopamine depression in the striatum and increased superoxide scavenging activity in mouse brain [105, 106]. Growing studies indicated that various herbs such as green tea polyphenols (catechins), ginseng (ginsenoside), ginkgo biloba, *etc.* attenuate degeneration of dopamine neurons caused by the neurotoxins 1-methyl-4-phenyl-1,2,3,6- tetrahydropyridine (MPTP) and 6-hydroxydopamine (6-OHDA) and have suggested to promote neuronal survival, and facilitate functional recovery of brain injures through neuroprotective effects such as the antioxidants, dopamine transporter inhibitors, monoamine oxidase inhibitor, free radical scavengers, chelators of harmful metal ions, modulating cell survival genes, signaling and anti-apoptosis activity [107]. Studies also reported a protective effect of ginseng against PD by promoting neuronal cell survival and reducing the neurotoxicity induced by neurotoxin toxins [108]. Commonly used spices and herbs are shown in Table **5**.

Herbal Drug	Biological Source	Family	Possible Mechanism	References
Ginkgo	Ginkgo biloba	Ginkgoaceae	protects apoptosis of PC12 cells by increasing bcl-2 activation, maintaining mitochondrial membrane potential and decreasing caspase-3 activation through the mitochondria-dependent pathway, blockade of lipid peroxidation, reduction of oxidative stress, and prevents neurodegeneration of the nigrostriatal pathway	[93]
Ginseng	Panax ginseng and Panax notoginseng	Araliaceae	Antioxidant effect on hydrogen peroxide (H2O2)-induced oxidative stress to PC12 cells, the release of cytochrome c and activation of caspase-3, elevated Bax/Bcl-2 ratio, decreases the increase of iron influx	[97]
Baicalein	Scutellaria baicalensis	Labiatae	Inhibits the accumulation of ROS, deficiency of ATP, dissipation of mitochondrial membrane potential, and activation of caspase-3/7	[109]
Curcumin	Curcuma longa	Zingiberaceae	Decrease intracellular ROS generation and inhibit caspase-3 activation, inhibits MPTP- induced hyperphosphorylation of c-Jun N- terminal kinase (JNK), prevents the degeneration of nigrostriatal neurons by inhibiting the dysfunction of mitochondria, neuroprotection	[66]

Table 5. List of comm	only used Spices	and Herbs in PD.
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Neurobiological Disorders

Herbal Drug	Biological Source	Family	Possible Mechanism	References
Gastrodin	Gastrodia elata	Orchidaceae	Reduces the proportion of apoptotic cells, ROS, and Bax/Bcl-2 ratio	[110]
Tenuigenin	Polygala tenuifolia	Polygalaceae	Prevents the abnormal shrinking of dendrites and promotes the survival of mesencephalic dopaminergic neurons, downregulate caspase-3 activity, neuroprotective effects <i>via</i> its antioxidant and antiapoptotic profile	[111]

Epilepsy

Epilepsy is a group of heterogeneous disorders characterized by the propensity to experience spontaneous recurrent seizures. The condition can be genetic or acquired, and the underlying mechanisms of disease initiation, propagation, and comorbid conditions are incompletely understood. Reports suggested that metabolic changes, including the production of reactive species, are known to result from prolonged seizures and may further contribute to epilepsy development [112].

Bilobalide in *Ginkgo biloba* is extensively used as an anticonvulsant and cognitive enhancement. Valerenic acid present in Valerian root and rhizome has beneficial effects in a number of neurological disorders such as epilepsy, insomnia, dizziness, and anxiety. Ginkgolide and Zerumin isolated from *Curcuma kwangsiensis* rhizomes have GABAA modulatory action [30]. Black pepper extract has been found to prolong anticonvulsant activity against audiogenic seizures in different animal models [113]. Eugenol in clove suppresses epileptiform field potentials in neocortical and hippocampal tissues [114]. Anethole in tarragon exhibited dosedependent antiseizure activity in maximal electroshock and pentylenetetrazole models of experimental seizures [112]. Horseradish and Celery seed reduces seizure phenotype attributed to the presence of kaempferol and apigenin [115].

Curcumin is reported to have a beneficial effect in ameliorating seizures, oxidative stress, and cognitive impairment in pentylene-tetrazole-treated animals [116].

Depression

Major depression is a commonly occurring multifactorial disorder classified under life-threatening neuropsychiatric conditions. Several mechanisms for the pathophysiology of depression are known. Traditionally, decreased monoamine function in the specific region of the brain is considered to be the primary cause of depression [117]. The imbalance between neuroprogressive and neuroprotective

factors is observed in major depression. Other factors include proinflammatory cytokines and lipid peroxidation. Also, Mitochondrion dysfunctioning plays a significant role in the pathophysiology of depression *via* actions of free radicals, nonradical molecules, and reactive oxygen and nitrogen species [118]. Stress and inflammatory processes are known to contribute to the development of depression. Several studies also suggested that stress leads to the activation of microglia, which increases the production of inflammatory mediators such as IL-1 β , IL-6, and TNF- α [3]. Further studies have shown that major depressive disorders are associated with the dysregulation of the hypothalamic-pituitary-adrenal (HPA) axis, which leads to elevated levels of glucocorticoids and impaired glucocorticoid receptor-mediated negative feedback on the function of the HPA axis [117]. Recent reports also suggest that neurotrophic factors play a critical role in the pathogenesis of depression. Analysis revealed that depression is associated with lower levels of BDNF [118].

The secondary metabolites present in herbs and spices, such as polyphenols, flavonoids, and alkaloids possess antidepressant activity through a different mechanism. Alkaloids present in *Areca catechu* (Arecaidine, Arecoline, Guvacine) and *Withania somnifera* (Withanolides and withanols) show mood-stabilizing action through inhibition of MAO-A. Curcumin, Hyperforin, and Rutin also possess inhibitory action against MAO-A. Flavonoid present in herb *Apocynum venetum* is found to modulate the central monoaminergic system and thus can be effective in the treatment of depression. Several herbs also act against oxidative stress and thus can be potential molecules against depression, such as bacoside-A in *Bacopa monniera* and warifteine in *Cissampelos sympodialis* [119].

Although several spices are found to be effective in the treatment of depression, saffron has been one of the first to be tested as a treatment of mild depression in clinical trials [64]. Garlic extract also possesses anti-depression properties *via* inhibition of monoamine oxidases and thereby increasing levels of monoamine neurotransmitters such as norepinephrine, dopamine, and serotonin, and also by decreasing GABA levels [120]. Chronic treatment of piperine significantly ameliorates behavioral deficits of CUMS-treated animals, associated with significantly increased BDNF protein expression in the hippocampus and frontal cortex [121]. Curcumin also alle*via*tes symptoms of depression by enhancing neurogenesis in the hippocampus and frontal cortex of the brain [122]. Traditionally Ginkgo biloba has been prescribed in the treatment of depression [123]. Eugenol in Cloves also acts as an antidepressant by inducing the expression of metallothionein-III in the hippocampus. Eugenol also induces upregulation of BDNF in the hippocampus of mice [124].

Schizophrenia

Schizophrenia is a neuropsychiatric condition characterized by positive symptoms such as the presence of abnormal feelings or behaviors, including hallucinations, delusions, and negative symptoms such as lack of interest and indifference in daily life. Two major hypotheses that explain the etiology of schizophrenia include dopamine dysfunction and glutamatergic hypofunction. More recently, increased cytokine levels, including pro-inflammatory cytokines IL-6, IL-1 β , TNF- α , have been found to be associated with schizophrenia [3]. Apart from neuroinflammation, oxidative stress has been suggested to contribute to the pathophysiology of the disease. Experimental findings demonstrated that oxidative stress is responsible for behavioral and molecular anomalies associated with schizophrenia [125].

Quercetin rutoside, an antioxidant glycoside present in several herbs and spices, can quench the superoxide production without interfering with the electron transfer activity of the reductase. It is also reported that higher concentrations of quercetin in red onions may be beneficial in neuropsychiatric conditions [126]. The anthraquinone derivative emodin suppressed the acoustic startle response and abolished prepulse inhibition associated with psychiatric conditions. The application of emodin also attenuated the phosphorylation of ErbB1, ErbB2, and EGF receptor signaling and ameliorated behavioral deficits [125].

CONCLUSION

Herbs and spices are gifts of nature to mankind, as they are inherited with numerous components which possess several curative properties. In this chapter, several spices and herbs which are beneficial in treating various neurobiological disorders and their probable mechanisms of action are described. Though various drugs for treating neurological disorders like depression, AD, PD, Schizophrenia and epilepsy are available, treatment with spices and herbs described here may overcome the adverse effects of regular treatment.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The authors declared that there is no conflict of interest.

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REFERENCES

- [1] Kochhar KP. Dietary spices in health and diseases: I Article in Indian journal of physiology and pharmacology. 2008; 106-22.
- [2] Nadkarni KM. Indian materia medica. 1996.
- [3] Hong H, Kim BS, Im HI. Pathophysiological role of neuroinflammation in neurodegenerative diseases and psychiatric disorders. Int Neurourol J 2016; 20 (Suppl. 1): S2-7. [http://dx.doi.org/10.5213/inj.1632604.302] [PMID: 27230456]
- [4] Patel M. Targeting Oxidative Stress in Central Nervous System Disorders. Trends Pharmacol Sci 2016; 37(9): 768-78.

[http://dx.doi.org/10.1016/j.tips.2016.06.007] [PMID: 27491897]

- [5] Geronzi U, Lotti F, Grosso S. Oxidative stress in epilepsy. Expert Rev Neurother 2018; 18(5): 427-34.
 [http://dx.doi.org/10.1080/14737175.2018.1465410] [PMID: 29651881]
- [6] Lee GT, Ha H, Lee HC, Cho YD. Agmatine reduces hydrogen peroxide in mesangial cells under high glucose conditions. J Biochem Mol Biol 2003; 36(3): 251-7. [PMID: 12787478]
- [7] Dong XX, Wang Y, Qin ZH. Molecular mechanisms of excitotoxicity and their relevance to pathogenesis of neurodegenerative diseases. Acta Pharmacol Sin 2009; 30(4): 379-87. [http://dx.doi.org/10.1038/aps.2009.24] [PMID: 19343058]
- [8] de Almeida RN, Agra M de F, Maior FN, de Sousa DP. Essential oils and their constituents: anticonvulsant activity. Molecules 2011; 16(3): 2726-42. [http://dx.doi.org/10.3390/molecules16032726] [PMID: 21441872]
- [9] Anekonda TS, Reddy PH. Can herbs provide a new generation of drugs for treating Alzheimer's disease? Brain Res Brain Res Rev 2005; 50(2): 361-76.
 [http://dx.doi.org/10.1016/j.brainresrev.2005.09.001] [PMID: 16263176]
- [10] Facchini PJ. Alkaloid biosynthesis in plants: Biochemistry, Cell Biology, Molecular Regulation, and Metabolic Engineering Applications. Annu Rev Plant Physiol Plant Mol Biol 2001; 52(1): 29-66. [http://dx.doi.org/10.1146/annurev.arplant.52.1.29] [PMID: 11337391]
- [11] Sun AY, Wang Q, Simonyi A, Sun GY. Botanical phenolics and brain health. Neuromolecular Med 2008; 10(4): 259-74.
 [http://dx.doi.org/10.1007/s12017-008-8052-z] [PMID: 19191039]
- Kim J, Lee HJ, Lee KW. Naturally occurring phytochemicals for the prevention of Alzheimer's disease. J Neurochem 2010; 112(6): 1415-30.
 [http://dx.doi.org/10.1111/j.1471-4159.2009.06562.x] [PMID: 20050972]
- Priyadarsini KI, Guha SN, Rao MNA. Physico-chemical properties and antioxidant activities of methoxy phenols. Free Radic Biol Med 1998; 24(6): 933-41.
 [http://dx.doi.org/10.1016/S0891-5849(97)00382-1] [PMID: 9607603]
- [14] Sreejayan, Rao MN. Nitric oxide scavenging by curcuminoids. J Pharm Pharmacol 1997; 49(1): 105--. [http://dx.doi.org/10.1111/j.2042-7158.1997.tb06761.x] [PMID: 9120760]
- [15] Hendrich AB. Flavonoid-membrane interactions: possible consequences for biological effects of some polyphenolic compounds. Acta Pharmacol Sin 2006; 27(1): 27-40.
 [http://dx.doi.org/10.1111/j.1745-7254.2006.00238.x] [PMID: 16364208]
- [16] Wu D, Guo Z, Ren Z, Guo W, Meydani SN. Green tea EGCG suppresses T cell proliferation through impairment of IL-2/IL-2 receptor signaling. Free Radic Biol Med 2009; 47(5): 636-43. [http://dx.doi.org/10.1016/j.freeradbiomed.2009.06.001] [PMID: 19501156]
- Shan B, Cai YZ, Sun M, Corke H. Antioxidant capacity of 26 spice extracts and characterization of their phenolic constituents. J Agric Food Chem 2005; 53(20): 7749-59.
 [http://dx.doi.org/10.1021/jf051513y] [PMID: 16190627]

- [18] Xu R-X, Wu Q, Luo Y, *et al.* Protective effects of icariin on cognitive deficits induced by chronic cerebral hypoperfusion in rats. Clin Exp Pharmacol Physiol 2009; 36(8): 810-5. [http://dx.doi.org/10.1111/j.1440-1681.2009.05149.x] [PMID: 19215241]
- [19] Pandey AK, Hazari PP, Patnaik R, Mishra AK. The role of ASIC1a in neuroprotection elicited by quercetin in focal cerebral ischemia. Brain Res 2011; 1383: 289-99. [http://dx.doi.org/10.1016/j.brainres.2011.01.085] [PMID: 21281608]
- [20] Ansari MA, Abdul HM, Joshi G, Opii WO, Butterfield DA. Protective effect of quercetin in primary neurons against Abeta(1-42): relevance to Alzheimer's disease. J Nutr Biochem 2009; 20(4): 269-75. [http://dx.doi.org/10.1016/j.jnutbio.2008.03.002] [PMID: 18602817]
- [21] Hossain MB, Rai DK, Brunton NP, Martin-Diana AB, Barry-Ryan C. Characterization of phenolic composition in Lamiaceae spices by LC-ESI-MS/MS. J Agric Food Chem 2010; 58(19): 10576-81. [http://dx.doi.org/10.1021/jf102042g] [PMID: 20825192]
- [22] Thatoi HN, Patra JK, Das SK. Free radical scavenging and antioxidant potential of mangrove plants: A review. Acta Physiol Plant 2014; 36(3): 561-79. [http://dx.doi.org/10.1007/s11738-013-1438-z]
- [23] Nakatani N. Phenolic antioxidants from herbs and spices. Biofactors 2000; 13(1-4): 141-6. [http://dx.doi.org/10.1002/biof.5520130123] [PMID: 11237173]
- [24] Talavéra S, Felgines C, Texier O, *et al.* Anthocyanin metabolism in rats and their distribution to digestive area, kidney, and brain. J Agric Food Chem 2005; 53(10): 3902-8. [http://dx.doi.org/10.1021/jf050145v] [PMID: 15884815]
- [25] Wojdyło A, Oszmiański J, Czemerys R. Antioxidant activity and phenolic compounds in 32 selected herbs. Food Chem 2007; 105(3): 940-9. [http://dx.doi.org/10.1016/j.foodchem.2007.04.038]
- [26] Trease and Evans. Trease and Evans Pharmacognosy, International Edition E-Book William Charles Evans - Google Books..
- [27] Ocaña-Fuentes A, Arranz-Gutiérrez E, Señorans FJ, Reglero G. Supercritical fluid extraction of oregano (Origanum vulgare) essentials oils: anti-inflammatory properties based on cytokine response on THP-1 macrophages. Food Chem Toxicol 2010; 48(6): 1568-75. [http://dx.doi.org/10.1016/j.fct.2010.03.026] [PMID: 20332013]
- [28] Saki K, Bahmani M, Rafieian-Kopaei M, Hassanzadazar H, Dehghan K, Bahmani F, et al. The most common native medicinal plants used for psychiatric and neurological disorders in Urmia city, northwest of Iran. Asian Pac J Trop Dis 2014; 4(S2): S895-901. [http://dx.doi.org/10.1016/S2222-1808(14)60754-4]
- [29] Rubió L, Motilva MJ, Romero MP. Recent advances in biologically active compounds in herbs and spices: a review of the most effective antioxidant and anti-inflammatory active principles. Crit Rev Food Sci Nutr 2013; 53(9): 943-53. [http://dx.doi.org/10.1080/10408398.2011.574802] [PMID: 23768186]
- [30] Manayi A, Nabavi SM, Daglia M, Jafari S. Natural terpenoids as a promising source for modulation of GABAergic system and treatment of neurological diseases. Pharmacol Rep 2016; 68(4): 671-9. [http://dx.doi.org/10.1016/j.pharep.2016.03.014] [PMID: 27110875]
- [31] McNaught A D, Wilkinson A. Compendium of Chemical Terminology The Gold Book. 2nd Edition
- [32] Girdhar S, Girdhar A, Kumar Verma S, Lather V, Pandita D. Plant derived alkaloids in major neurodegenerative diseases: from animal models to clinical trials. J Ayurvedic Herb Med 2015; 1(3): 91-100.
- [33] Ng YP, Or TCT, Ip NY. Plant alkaloids as drug leads for Alzheimer's disease. Neurochem Int 2015; 89: 260-70.
 [http://dx.doi.org/10.1016/j.neuint.2015.07.018] [PMID: 26220901]

- [34] Jiang X, Jia LW, Li XH, et al. Capsaicin ameliorates stress-induced Alzheimer's disease-like pathological and cognitive impairments in rats. J Alzheimers Dis 2013; 35(1): 91-105. [http://dx.doi.org/10.3233/JAD-121837] [PMID: 23340038]
- [35] Barreto GE, Iarkov A, Moran VE. Beneficial effects of nicotine, cotinine and its metabolites as potential agents for Parkinson's disease. Front Aging Neurosci 2015; 6(JAN): 340. [PMID: 25620929]
- [36] Hussain G, Rasul A, Anwar H, et al. Role of plant derived alkaloids and their mechanism in neurodegenerative disorders. Int J Biol Sci 2018; 14(3): 341-57. [http://dx.doi.org/10.7150/ijbs.23247] [PMID: 29559851]
- [37] Salomon AR, Marcinowski KJ, Friedland RP, Zagorski MG. Nicotine inhibits amyloid formation by the β-peptide. Biochemistry 1996; 35(42): 13568-78.
 [http://dx.doi.org/10.1021/bi9617264] [PMID: 8885836]
- [38] Huang M, Jiang X, Liang Y, Liu Q, Chen S, Guo Y. Berberine improves cognitive impairment by promoting autophagic clearance and inhibiting production of β-amyloid in APP/tau/PS1 mouse model of Alzheimer's disease. Exp Gerontol 2017; 91: 25-33. [http://dx.doi.org/10.1016/j.exger.2017.02.004] [PMID: 28223223]
- [39] Chonpathompikunlert P, Wattanathorn J, Muchimapura S. Piperine, the main alkaloid of Thai black pepper, protects against neurodegeneration and cognitive impairment in animal model of cognitive deficit like condition of Alzheimer's disease. Food Chem Toxicol 2010; 48(3): 798-802. [http://dx.doi.org/10.1016/j.fct.2009.12.009] [PMID: 20034530]
- [40] Kulkarni SK, Bhutani MK, Bishnoi M. Antidepressant activity of curcumin: involvement of serotonin and dopamine system. Psychopharmacology (Berl) 2008; 201(3): 435-42. [http://dx.doi.org/10.1007/s00213-008-1300-y] [PMID: 18766332]
- [41] Frost D, Meechoovet B, Wang T, Gately S, Giorgetti M, Shcherbakova I, et al. β-Carboline Compounds, Including Harmine, Inhibit DYRK1A and Tau Phosphorylation at Multiple Alzheimer's Disease-Related Sites. Skoulakis EMC, editor. PLoS One. 2011; 6: p. (5)e19264.
- [42] Origins G. Occurrence and Risk Assessment of Pyrrolizidine Geographical Origins. 2020.
- [43] Almeida MB, Costa-Malaquias A, Nascimento JLM, Oliveira KR, Herculano AM, Crespo-López ME. Therapeutic concentration of morphine reduces oxidative stress in glioma cell line. Braz J Med Biol Res 2014; 47(5): 398-402. [http://dx.doi.org/10.1590/1414-431X20143697] [PMID: 24728211]
- [44] Cui J, Wang Y, Dong Q, et al. Morphine protects against intracellular amyloid toxicity by inducing estradiol release and upregulation of Hsp70. J Neurosci 2011; 31(45): 16227-40. [http://dx.doi.org/10.1523/JNEUROSCI.3915-11.2011] [PMID: 22072674]
- [45] Pagliosa LB, Monteiro SC, Silva KB, et al. Effect of isoquinoline alkaloids from two Hippeastrum species on *in vitro* acetylcholinesterase activity. Phytomedicine 2010; 17(8-9): 698-701. [http://dx.doi.org/10.1016/j.phymed.2009.10.003] [PMID: 19969445]
- [46] Schmidt AJ, Krieg JC, Vedder H. Antioxidative and steroid systems in neurological and psychiatric disorders. World J Biol Psychiatry 2005; 6(1): 26-35. [http://dx.doi.org/10.1080/15622970510029759] [PMID: 16097403]
- [47] Sayre LM, Moreira PI, Smith MA, Perry G. Metal ions and oxidative protein modification in neurological disease. Ann Ist Super Sanita 2005; 41(2): 143-64. [PMID: 16244388]
- [48] Halliwell B. Reactive oxygen species and the central nervous system. J Neurochem 1992; 59(5): 1609-23.
 [http://dx.doi.org/10.1111/j.1471-4159.1992.tb10990.x] [PMID: 1402908]
- [49] Smith KJ, Kapoor R, Felts PA. Demyelination: the role of reactive oxygen and nitrogen species. Brain

Pathol 1999; 9(1): 69-92.

[http://dx.doi.org/10.1111/j.1750-3639.1999.tb00212.x] [PMID: 9989453]

[50] Yu BP. Cellular defenses against damage from reactive oxygen species. Physiol Rev 1994; 74(1): 139-62.

[http://dx.doi.org/10.1152/physrev.1994.74.1.139] [PMID: 8295932]

- [51] Ninfali P, Mea G, Giorgini S, Rocchi M, Bacchiocca M. Antioxidant capacity of vegetables, spices and dressings relevant to nutrition. Br J Nutr 2005; 93(2): 257-66. [http://dx.doi.org/10.1079/BJN20041327] [PMID: 15788119]
- [52] Brabin BJ, Hakimi M, Pelletier D. Several Culinary and Medicinal Herbs Are Important Sources of Dietary Antioxidants. J Nutr 2001; 131(2): 604S-15S. [http://dx.doi.org/10.1093/jn/131.2.604S] [PMID: 11160593]
- [53] Zheng W, Wang SY. Antioxidant activity and phenolic compounds in selected herbs. J Agric Food Chem 2001; 49(11): 5165-70.
 [http://dx.doi.org/10.1021/jf010697n] [PMID: 11714298]
- [54] Kim YO, Leem K, Park J, et al. Cytoprotective effect of Scutellaria baicalensis in CA1 hippocampal neurons of rats after global cerebral ischemia. J Ethnopharmacol 2001; 77(2-3): 183-8. [http://dx.doi.org/10.1016/S0378-8741(01)00283-5] [PMID: 11535362]
- [55] Bhattachajya SK. Kalkunte •, Satyan S, Ghosal S. Antioxidant activity of glycowithano lides from Withania somnifera. IBdian J Exp Biol 1997; 35: 236-9.
- [56] Scapagnini G, Foresti R, Calabrese V, Giuffrida Stella AM, Green CJ, Motterlini R. Caffeic acid phenethyl ester and curcumin: a novel class of heme oxygenase-1 inducers. Mol Pharmacol 2002; 61(3): 554-61.
 [http://dx.doi.org/10.1124/mol.61.3.554] [PMID: 11854435]
- [57] Saini N, Singh D, Sandhir R. Neuroprotective effects of Bacopa monnieri in experimental model of dementia. Neurochem Res 2012; 37(9): 1928-37.
 [http://dx.doi.org/10.1007/s11064-012-0811-4] [PMID: 22700087]
- [58] Skaper SD, Facci L, Zusso M, Giusti P. An inflammation-centric view of neurological disease: Beyond the neuron. Front Cell Neurosci 2018; 12: 72. [http://dx.doi.org/10.3389/fncel.2018.00072] [PMID: 29618972]
- [59] Simon E, Obst J, Gomez-Nicola D. The Evolving Dialogue of Microglia and Neurons in Alzheimer's Disease: Microglia as Necessary Transducers of Pathology. Neuroscience 2019; 405: 24-34. [http://dx.doi.org/10.1016/j.neuroscience.2018.01.059] [PMID: 29427657]
- [60] Kempuraj D, Thangavel R, Selvakumar GP, et al. Brain and peripheral atypical inflammatory mediators potentiate neuroinflammation and neurodegeneration. Front Cell Neurosci 2017; 11: 216. [http://dx.doi.org/10.3389/fncel.2017.00216] [PMID: 28790893]
- [61] Appel SH, Beers DR, Zhao W. Neurobiology of Brain Disorders. Elsevier Inc. 2015; pp. 380-95.
- [62] Tapsell LC, Hemphill I, Cobiac L, et al. Health benefits of herbs and spices: the past, the present, the future. Med J Aust 2006; 185(S4) (Suppl.): S1-S24. [http://dx.doi.org/10.5694/j.1326-5377.2006.tb00548.x] [PMID: 17022438]
- [63] Mueller M, Hobiger S, Jungbauer A. Anti-inflammatory activity of extracts from fruits, herbs and spices. Food Chem 2010; 122(4): 987-96. [http://dx.doi.org/10.1016/j.foodchem.2010.03.041]
- [64] Kurian A. Health benefits of herbs and spices. Handb Herbs Spices Second Ed. 2012. [http://dx.doi.org/10.1533/9780857095688.72]
- [65] Cole GM, Morihara T, Lim GP, Yang F, Begum A, Frautschy SA. NSAID AND ANTIOXIDANT PREVENTION OF AD. Ann N Y Acad Sci 2004; 1035: 68-84. [http://dx.doi.org/10.1196/annals.1332.005] [PMID: 15681801]

- [66] Lim GP, Chu T, Yang F, Beech W, Frautschy SA, Cole GM. The curry spice curcumin reduces oxidative damage and amyloid pathology in an Alzheimer transgenic mouse. J Neurosci 2001; 21(21): 8370-7.
 [http://dx.doi.org/10.1523/JNEUROSCI.21-21-08370.2001] [PMID: 11606625]
- [67] Lee IO, Lee KH, Pyo JH, Kim JH, Choi YJ, Lee YC. Anti-inflammatory effect of capsaicin in Helicobacter pylori-infected gastric epithelial cells. Helicobacter 2007; 12(5): 510-7. [http://dx.doi.org/10.1111/j.1523-5378.2007.00521.x] [PMID: 17760719]
- [68] Park J-Y, Kawada T, Han I-S, *et al.* Capsaicin inhibits the production of tumor necrosis factor α by LPS-stimulated murine macrophages, RAW 264.7: a PPARgamma ligand-like action as a novel mechanism. FEBS Lett 2004; 572(1-3): 266-70. [http://dx.doi.org/10.1016/j.febslet.2004.06.084] [PMID: 15304360]
- [69] Ma J, Hwang YK, Cho W-H, Han S-H, Hwang JK, Han J-S. Macelignan attenuates activations of mitogen-activated protein kinases and nuclear factor kappa B induced by lipopolysaccharide in microglial cells. Biol Pharm Bull 2009; 32(6): 1085-90. [http://dx.doi.org/10.1248/bpb.32.1085] [PMID: 19483320]
- [70] Taylor P. Development of acetylcholinesterase inhibitors in the therapy of Alzheimer's disease. Neurology 1998; 51(1) (Suppl. 1): S30-5.
 [http://dx.doi.org/10.1212/WNL.51.1_Suppl_1.S30] [PMID: 9674760]
- Schulz V. Ginkgo extract or cholinesterase inhibitors in patients with dementia: what clinical trials and guidelines fail to consider. Phytomedicine 2003; 10 (Suppl. 4): 74-9.
 [http://dx.doi.org/10.1078/1433-187X-00302] [PMID: 12807348]
- [72] Kumar P, Singh VK, Singh DK. Kinetics of enzyme inhibition by active molluscicidal agents ferulic acid, umbelliferone, eugenol and limonene in the nervous tissue of snail *Lymnaea acuminata*. Phytother Res 2009; 23(2): 172-7.
 [http://dx.doi.org/10.1002/ptr.2578] [PMID: 18814203]
- [73] Janko S, Dubravka SS. Benzodiazepines and Anxiety disorders: From laboratory to clinic. In New Developments in Anxiety Disorders. 2016; pp. 23-45.
- [74] Jang SW, Okada M, Sayeed I, *et al.* Gambogic amide, a selective agonist for TrkA receptor that possesses robust neurotrophic activity, prevents neuronal cell death. Proc Natl Acad Sci USA 2007; 104(41): 16329-34.
 [http://dx.doi.org/10.1073/pnas.0706662104] [PMID: 17911251]
- [75] Delgado Marin L, Sanchez-Borzone M. A. Garcia D. Comparative Antioxidant Properties of Some Gabaergic Phenols and Related Compounds, Determined for Homogeneous and Membrane Systems. Med Chem (Los Angeles) 2011; 7(4): 317-24.
- [76] Waliwitiya R, Belton P, Nicholson RA, Lowenberger CA. Effects of the essential oil constituent thymol and other neuroactive chemicals on flight motor activity and wing beat frequency in the blowfly *Phaenicia sericata*. Pest Manag Sci 2010; 66(3): 277-89. [http://dx.doi.org/10.1002/ps.1871] [PMID: 19890946]
- [77] El-Nekeety AA, Mohamed SR, Hathout AS, Hassan NS, Aly SE, Abdel-Wahhab MA. Antioxidant properties of Thymus vulgaris oil against aflatoxin-induce oxidative stress in male rats. Toxicon 2011; 57(7-8): 984-91.
 [http://dx.doi.org/10.1016/j.toxicon.2011.03.021] [PMID: 21477612]
- [78] Calabrese V, Scapagnini G, Colombrita C, *et al.* Redox regulation of heat shock protein expression in aging and neurodegenerative disorders associated with oxidative stress: a nutritional approach. Amino Acids 2003; 25(3-4): 437-44.
 [http://dx.doi.org/10.1007/s00726-003-0048-2] [PMID: 14661103]
- [79] Murpy M, Dis HLI-JA. undefined. Alzheimer's disease and the β-amyloid peptide. 2019; 1(19): 311-23.

- [80] Varadarajan S, Yatin S, Aksenova M, Butterfield DA. Review: Alzheimer's amyloid β-peptidassociated free radical oxidative stress and neurotoxicity. J Struct Biol 2000; 130(2-3): 184-208. [http://dx.doi.org/10.1006/jsbi.2000.4274] [PMID: 10940225]
- [81] Butterfield DA, Drake J, Pocernich C, Castegna A. Evidence of oxidative damage in Alzheimer's disease brain: Central role for amyloid β-peptide. Trends in Molecular Medicine. Elsevier Current Trends 2001; Vol. 7: pp. 548-54.
- [82] Reddy PH, Manczak M, Yin X, et al. Protective effects of Indian spice curcumin against Amyloid-β in Alzheimer's disease. J Alzheimers Dis 2018; 61(3): 843-66. [http://dx.doi.org/10.3233/JAD-170512] [PMID: 29332042]
- [83] Baum L, Ng A. Curcumin interaction with copper and iron suggests one possible mechanism of action in Alzheimer's disease animal models. J Alzheimers Dis 2004; 6(4): 367-77. [http://dx.doi.org/10.3233/JAD-2004-6403] [PMID: 15345806]
- [84] Atamna H, Boyle K. Amyloid-β peptide binds with heme to form a peroxidase: relationship to the cytopathologies of Alzheimer's disease. Proc Natl Acad Sci USA 2006; 103(9): 3381-6. [http://dx.doi.org/10.1073/pnas.0600134103] [PMID: 16492752]
- [85] Fiala M, Liu PT, Espinosa-Jeffrey A, *et al.* Innate immunity and transcription of MGAT-III and Tolllike receptors in Alzheimer's disease patients are improved by bisdemethoxycurcumin. Proc Natl Acad Sci USA 2007; 104(31): 12849-54. [http://dx.doi.org/10.1073/pnas.0701267104] [PMID: 17652175]
- [86] Kamal Z, Ullah F, Ayaz M, et al. Anticholinesterase and antioxidant investigations of crude extracts, subsequent fractions, saponins and flavonoids of atriplex laciniata L.: potential effectiveness in Alzheimer's and other neurological disorders. Biol Res 2015; 48(1): 21. [http://dx.doi.org/10.1186/s40659-015-0011-1] [PMID: 25889712]
- [87] Iuvone T, De Filippis D, Esposito G, D'Amico A, Izzo AA. The spice sage and its active ingredient rosmarinic acid protect PC12 cells from amyloid-β peptide-induced neurotoxicity. J Pharmacol Exp Ther 2006; 317(3): 1143-9. [http://dx.doi.org/10.1124/jpet.105.099317] [PMID: 16495207]
- [88] Peng Q, Buz'Zard AR, Lau BHS. Neuroprotective effect of garlic compounds in amyloid-β peptideinduced apoptosis *in vitro*. Med Sci Monit 2002; 8(8): BR328-37. [PMID: 12165737]
- [89] Guo JP, Yu S, McGeer PL. Simple *in vitro* assays to identify amyloid-β aggregation blockers for Alzheimer's disease therapy. J Alzheimers Dis 2010; 19(4): 1359-70. [http://dx.doi.org/10.3233/JAD-2010-1331] [PMID: 20061605]
- [90] Ryu EK, Choe YS, Lee KH, Choi Y, Kim BT. Curcumin and dehydrozingerone derivatives: Synthesis, radiolabeling, and evaluation for β-amyloid plaque imaging. Journal of Medicinal Chemistry. American Chemical Society 2006; pp. 6111-9. [http://dx.doi.org/10.1021/jm0607193]
- [91] Kannappan R, Gupta SC, Kim JH, Reuter S, Aggarwal BB. Neuroprotection by spice-derived nutraceuticals: you are what you eat! Mol Neurobiol 2011; 44(2): 142-59. [http://dx.doi.org/10.1007/s12035-011-8168-2] [PMID: 21360003]
- [92] Agarwal P, Fatima A, Singh PP. Herbal Medicine Scenario in India and European Countries. Vol. 1. J Pharmacogn Phytochem 2012.
- [93] Sierpina VS, Wollschlaeger B, Blumenthal M. Ginkgo biloba. Am Fam Physician 2003; 68(5): 923-6.[PMID: 13678141]
- [94] Hamidpour M, Hamidpour R, Hamidpour S, Shahlari M. Chemistry, Pharmacology, and Medicinal Property of Sage (Salvia) to Prevent and Cure Illnesses such as Obesity, Diabetes, Depression, Dementia, Lupus, Autism, Heart Disease, and Cancer. J Tradit Complement Med 2014; 4(2): 82-8.http://www.ncbi.nlm.nih.gov/pubmed/24860730 [Internet].

[http://dx.doi.org/10.4103/2225-4110.130373] [PMID: 24860730]

- [95] De Oliveira JR, Camargo SEA, De Oliveira LD. Rosmarinus officinalis L. (rosemary) as therapeutic and prophylactic agent. Journal of Biomedical Science BioMed Central Ltd 2019; Vol. 26
- [96] Prasad S, Aggarwal BB. Turmeric, the golden spice: From traditional medicine to modern medicine. In: Herbal Medicine: Biomolecular and Clinical Aspects: Second Edition [Internet]. CRC Press 2011. Available from: https://www.ncbi.nlm.nih.gov/books/NBK92752/
- [97] Lu J-M, Yao Q, Chen C. Ginseng Compounds: An Update on their Molecular Mechanisms and Medical Applications. Curr Vasc Pharmacol 2009; 7(3): 293-302. Available from: /pmc/articles/PMC2928028/
- [98] Mao QQ, Xu XY, Cao SY, Gan RY, Corke H, Beta T, et al. Bioactive compounds and bioactivities of ginger (zingiber officinale roscoe). Foods MDPI Multidisciplinary Digital Publishing Institute 2019; Vol. 8 Available from: /pmc/articles/PMC6616534/
- [99] Joshi R. Chemical composition and antimicrobial activity of the essential oil of Ocimum basilicum L. (sweet basil) from Western Ghats of North West Karnataka, India Anc Sci Life [Internet] 33(3): 149.2014; Available from: /pmc/articles/PMC4264302/
- [100] Önder A. Coriander and Its Phytoconstituents for the Beneficial Effects. Potential of Essential Oils. InTech 2018. Internet [http://dx.doi.org/10.5772/intechopen.78656]
- [101] Ghorbani A, Esmaeilizadeh M. Pharmacological properties of Salvia officinalis and its components. J Tradit Complement Med [Internet] 2017; 7(4): 433-0. Available from: /pmc/articles/PMC5634728/
- [102] Mandir AS, Vaughan C. Pathophysiology of Parkinson's disease. Int Rev Psychiatry 2000; 12(4): 270-80.
 [http://dx.doi.org/10.1080/09540260020002497]
- [103] Mythri RB, Bharath MM. Curcumin: a potential neuroprotective agent in Parkinson's disease. Curr Pharm Des 2012; 18(1): 91-9. [http://dx.doi.org/10.2174/138161212798918995] [PMID: 22211691]
- [104] Wang MS, Boddapati S, Emadi S, Sierks MR. Curcumin reduces α-synuclein induced cytotoxicity in Parkinson's disease cell model. BMC Neurosci 2010; 11(1): 57. [http://dx.doi.org/10.1186/1471-2202-11-57] [PMID: 20433710]
- [105] Kabuto H, Tada M, Kohno M. Eugenol [2-methoxy-4-(2-propenyl)phenol] prevents 6hydroxydopamine-induced dopamine depression and lipid peroxidation inductivity in mouse striatum. Biol Pharm Bull 2007; 30(3): 423-7.
 [http://dx.doi.org/10.1248/bpb.30.423] [PMID: 17329831]
- [106] Kabuto H, Nishizawa M, Tada M, Higashio C, Shishibori T, Kohno M. Zingerone [4-(4-hydrox--3-methoxyphenyl)-2-butanone] prevents 6-hydroxydopamine-induced dopamine depression in mouse striatum and increases superoxide scavenging activity in serum. Neurochem Res 2005; 30(3): 325-32. [http://dx.doi.org/10.1007/s11064-005-2606-3] [PMID: 16018576]
- [107] Chen L-W, Wang Y-Q, Wei L-C, Shi M, Chan YS. Chinese herbs and herbal extracts for neuroprotection of dopaminergic neurons and potential therapeutic treatment of Parkinson's disease. CNS Neurol Disord Drug Targets 2007; 6(4): 273-81.
 [http://dx.doi.org/10.2174/187152707781387288] [PMID: 17691984]
- [108] Iriti M, Vitalini S, Fico G, Faoro F. Neuroprotective herbs and foods from different traditional medicines and diets. Molecules 2010; 15(5): 3517-55. [http://dx.doi.org/10.3390/molecules15053517] [PMID: 20657497]
- [109] Liang S, Deng X, Lei L, et al. The Comparative Study of the Therapeutic Effects and Mechanism of Baicalin, Baicalein, and Their Combination on Ulcerative Colitis Rat. Front Pharmacol 2019; 10: 1466.https://www.frontiersin.org/article/10.3389/fphar.2019.01466/full [Internet]. [http://dx.doi.org/10.3389/fphar.2019.01466] [PMID: 31920656]

- [110] Liu Y, Gao J, Peng M, Meng H, Ma H, Cai P, et al. A review on central nervous system effects of gastrodin [Internet]. Frontiers in Pharmacology Frontiers Media SA 2018; Vol. 9: 24.
- [111] Lv J, Jia H, Jiang Y, et al. Tenuifolin, an extract derived from tenuigenin, inhibits amyloid- β secretion in vitro. Acta Physiol (Oxf) 2009; 196(4): 419-25. [http://dx.doi.org/10.1111/j.1748-1716.2009.01961.x] [PMID: 19208093]
- [112] Pearson-Smith JN, Patel M. Metabolic Dysfunction and Oxidative Stress in Epilepsy. Int J Mol Sci 2017; 18(11): 2365. [http://dx.doi.org/10.3390/ijms18112365] [PMID: 29117123]
- [113] Abila B, Richens A, Davies JA. Anticonvulsant effects of extracts of the west African black pepper, Piper guineense. J Ethnopharmacol 1993; 39(2): 113-7. [http://dx.doi.org/10.1016/0378-8741(93)90026-2] [PMID: 8412244]
- [114] Müller M, Pape HC, Speckmann EJ, Gorji A. Effect of eugenol on spreading depression and epileptiform discharges in rat neocortical and hippocampal tissues. Neuroscience 2006; 140(2): 743--. [http://dx.doi.org/10.1016/j.neuroscience.2006.02.036] [PMID: 16563641]
- [115] Song J, Parker L, Hormozi L, Tanouye MA. DNA topoisomerase I inhibitors ameliorate seizure-like behaviors and paralysis in a Drosophila model of epilepsy. Neuroscience 2008; 156(3): 722-8. [http://dx.doi.org/10.1016/j.neuroscience.2008.07.024] [PMID: 18703119]
- [116] Mehla J, Reeta KH, Gupta P, Gupta YK. Protective effect of curcumin against seizures and cognitive impairment in a pentylenetetrazole-kindled epileptic rat model. Life Sci 2010; 87(19-22): 596-603. [http://dx.doi.org/10.1016/j.lfs.2010.09.006] [PMID: 20840851]
- [117] Krishnan V, Nestler EJ. The molecular neurobiology of depression. Nature 2008; 455(7215): 894-902. [http://dx.doi.org/10.1038/nature07455] [PMID: 18923511]
- [118] Vaváková M, Trebatická J. Markers of Oxidative Stress and Neuroprogression in Depression Disorder. 2015

[http://dx.doi.org/10.1155/2015/898393]

- [119] Kumar GP, Anilakumar KR, Naveen S. Phytochemicals having neuroprotective properties from dietary sources and medicinal herbs. Pharmacogn J 2015; 7(1): 1-17. [http://dx.doi.org/10.5530/pj.2015.1.1]
- [120] Dhingra D, Goyal PK. Evidences for the involvement of monoaminergic and GABAergic systems in antidepressant-like activity of Tinospora cordifolia in mice. Indian J Pharm Sci 2008; 70(6): 761-7. [http://dx.doi.org/10.4103/0250-474X.49118] [PMID: 21369437]
- [121] Mao QQ, Huang Z, Zhong XM, Xian YF, Ip SP. Brain-derived neurotrophic factor signalling mediates the antidepressant-like effect of piperine in chronically stressed mice. Behav Brain Res 2014; 261: 140-5. [http://dx.doi.org/10.1016/j.bbr.2013.12.020] [PMID: 24361910]
- [122] Kulkarni S, Dhir A, Akula KK. Potentials of curcumin as an antidepressant. ScientificWorldJournal 2009; 9: 1233-41. [http://dx.doi.org/10.1100/tsw.2009.137] [PMID: 19882093]
- [123] Kleijnen J, Knipschild P. Ginkgo biloba for cerebral insufficiency. Br J Clin Pharmacol 1992; 34(4): 352-8. [http://dx.doi.org/10.1111/j.1365-2125.1992.tb05642.x] [PMID: 1457269]
- [124] Irie Y, Itokazu N, Anjiki N, Ishige A, Watanabe K, Keung WM. Eugenol exhibits antidepressant-like activity in mice and induces expression of metallothionein-III in the hippocampus. Brain Res 2004; 1011(2): 243-6.

[http://dx.doi.org/10.1016/j.brainres.2004.03.040] [PMID: 15157811]

[125] Bitanihirwe BKY, Woo TUW. Oxidative stress in schizophrenia: an integrated approach. Neurosci Biobehav Rev 2011; 35(3): 878-93.

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[http://dx.doi.org/10.1016/j.neubiorev.2010.10.008] [PMID: 20974172]

[126] Marchbanks RM, Ryan M, Day INM, Owen M, McGuffin P, Whatley SA. A mitochondrial DNA sequence variant associated with schizophrenia and oxidative stress. Schizophr Res 2003; 65(1): 33-8. [http://dx.doi.org/10.1016/S0920-9964(03)00011-2] [PMID: 14623372]

Spices and Herbs in Bacterial and Fungal Resistance

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Abstract: A plethora of pathogenic microorganisms is responsible for several infectious diseases. For many decades, the treatment of microbial infections includes the use of various antimicrobial agents. However, the extensive use of antibiotics has been found to develop multiple drug resistance (MDR) in many pathogens. Furthermore, the rate of MDR is higher in developing countries because of indiscriminate use and self-medication. The emergence of MDR limits the effectiveness and therapeutic options for common infections. As a result, much attention is given to naturally derived products that can be used as potential, with better efficacy, less expensive alternative, and safe antimicrobials for the treatment of common infections.

Herbal medicines have always been used as an alternative to treat diseases due to toxicity and associated side effects of allopathic medicines. In recent years, the use of herbs and spices in therapy has been gradually increasing in many developing countries because of their safety, efficacy, and other beneficial effects. Spices and herbs have been used for thousands of years for flavouring and preserving foods. Many of these herbs and spices, such as thyme, cinnamon, clove, oregano, cardamom, nutmeg, mint, and cumin, are known to exert a range of therapeutic activities, including antioxidant, anti-inflammatory, and anticancerogenic. These are also useful for preventing lipid oxidation and free radical scavenging agents in living organisms.

Spices and herbs demonstrate antimicrobial activity due to the presence of some of the important phytochemicals or essential oils, which are naturally toxic to microbes. The phytochemical screening disclosed the presence of a number of secondary metabolites such as resins, phenols, alkaloids, flavonoids, sterols, reducing sugars, tannins, glycosides, *etc.* and various essential oils which act on a wide range of microorganisms such as fungi, viruses, bacteria, protozoa, *etc.* The antimicrobial activity of the spices depends on the composition, the type of the spices, and various other environmental factors. The mechanism of its action by which these phytoconstituents act is generally complex and mostly depends on the presence of chemical constituents. It is also affected by the different cell wall components, such as its composition and the presence

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of different proteins on the cell envelope, as these are the major components that interact with the molecules. Some essential oils in plants cause partial disintegration due to the disruption of the lipopolysaccharide in the outer layer. It also inhibits the synthesis of nucleic acids, polysaccharides, and proteins in bacterial and fungal cells.

Thus, the use of spices and herbs presents a great potential to be used as an alternative or in addition with the allopathic medicine to decrease the side effects and progressively increase the resistance of pathogens induced by the use of allopathic drugs.

Keywords: Alkaloids, Spices, Herbs, Bacterial resistance, Essential oils, Cell wall synthesis inhibition, Protein synthesis.

INTRODUCTION

For centuries, spices and herbs have been used to improve the flavour and aroma of foods. Early cultures also reported the importance of using herbs and spices in the preservation of foods and their medicinal value. Since the late 19th century, as per scientific experiments, the antimicrobial properties or application of some spices, herbs, and their active components have been documented [1, 2]. There are many spices and herbs that are known for their antioxidant activity and importance in the prevention of lipid oxidation in living organisms as well as in foods. For instance, turmeric, oregano, cinnamon, cumin, parsley, garlic, mustard seed, ginger, basil, pepper, and cardamom are stated to possess antioxidant activity. For thousands of years and throughout the world, spices and herbs have been used for different purposes. Specifically, extracts of these crude spices and oils extracted from them have various applications, including alternative medicine, pharmaceutical, raw, and processed food preservation, and natural therapies. In addition, the antimicrobial action of different spices and herb extracts has been studied and reported against many microorganisms. Spices have also been used to combat snakebites, stomach disorders, poor eyesight, poor circulation, sleeping problems, colds, sores, motion sickness, lumbago, muscular aches, gout, and hangover [3 - 5].

Spices possess antibacterial and antifungal activity. Many microbiologists and food-product developers or specialists have conducted laboratory studies that involve diverse and challenging food-borne bacteria, fungi, and yeasts with extracted phytochemicals from spice plants. Several techniques have been used to examine antimicrobial activity, and the primary data vary considerably in quantity and quality among different sp0ices. Hence, it strongly demonstrates potent antimicrobial properties [6 - 8].

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Various bacteria show resistance towards many antimicrobial agents, which is called antibiotic resistance. In food contaminated by microbial pathogens, spoilage may occur and risk of foodborne diseases, and the development of multidrug-resistant and disinfectant resistant bacteria like Escherichia coli (E. coli), Pseudomonas aeruginosa (P. aeruginosa), and Staphylococcus aureus (S. aureus) may increase rapidly, causing an increase in the rate of mortality and morbidity [9]. Weak acids such as sorbic acids and benzoic acids [10] are commonly used in the food industry as preservatives to increase the safety. stability, and overall shelf-life of manufactured foods by controlling pathogenic and food-related spoilage causing microorganisms, that in turn may contribute to the development of microbiological resistance [11, 12]. However, chemical preservatives fail to eliminate several pathogenic bacteria in food products or extend the occurrence of microbial spoilage. Naturally obtained spices can be used as a preservative and are tolerated by the human body compared with synthetic products. The antimicrobial activities of natural products are applied in the food industry [13, 14].

The World Health Organization (WHO) has statistically reported that of the 55 million people who died in 2011 worldwide, one-third of the deaths were due to infectious diseases [15]. Antibiotic-resistant microorganisms can increase the death rate as they are not easily killed by antimicrobial agents [16]. Such situations increase the need for modifications in the structure of synthetic antibiotics that have been marketed [17]. Hence, much attention and importance must be paid to natural products like spices that can be used for treating various infectious diseases, with high efficacy against pathogens and minimal side effects [18]. Various spices such as oregano, clove, turmeric, cinnamon, ginger, cumin, and garlic have been used to prevent and treat infectious diseases and protect food. These spices were experimentally confirmed to exhibit antimicrobial activities against pathogenic fungi and bacteria [19, 20]. The secondary metabolites found in spices are antimicrobial agents that generally have negligible adverse effects [21]. Therefore, spices and herbs could be novel antimicrobial agents against food-derived and human pathogens.

ANTIBACTERIAL AGENTS

Antibiotics are compounds that are 'against life; are typically antibacterial agents, which interfere with some process or structure essential for bacterial growth or survival without any harm to the eukaryotic host harboring the infecting bacteria. There are two types of antibacterial agents, bactericidal (can kill bacteria) and bacteriostatic (nullify the growth). Mechanism of antibiotic resistance is shown in Fig. (1) while targets of spices and herbs for antibacterial activity are shown in Table 1.

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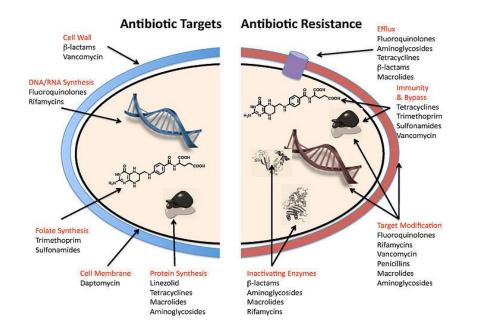


Fig. (1). Mechanism of Antibiotic Resistance.

DEVELOPMENT OF RESISTANCE

Pump Out The Antibiotic

To be effective, antibiotics must specifically target bacteria and get collected in specific effective concentrations that can act in the same sensible time frame. For instance, the protein synthesis machinery is situated in the cytoplasm, so antibacterial agents that are inhibitors of protein synthesis must pass across the cell membrane (inner membrane barriers for Gram-positive bacteria; outer and inner permeability barriers for Gram-negative bacteria) and then accumulate to adequate concentrations to block the target step during protein assembly. Both Gram-positive and negative bacteria that develop resistance to antibiotics (like tetracyclines) usually overproduce related membrane proteins (with comparative molecular masses of 42,000) that can act as an efflux or export pump for the drugs. A drug is pumped out of the cell faster than it can diffuse in, so intrabacterial concentration decreases and becomes ineffective, and bacterial protein synthesis continues at largely unhindered rates [22, 23].

Destroy the Antibiotic Warhead

A second approach to developing resistance is through the destruction of the specific chemical warhead in the synthetic or naturally obtained antibiotic. The classic example is hydrolytic deactivation of the β -lactam ring in the penicillins and cephalosporins through amplification of the hydrolytic enzyme β -lactamase by resistant bacteria. Due to strain, the four-membered lactam ring is the chemically activated functionality in the drugs that acylates and irreversibly modifies the cell wall-crosslinking. The lactamase-producing bacteria discharge this enzyme into the periplasm to destroy β -lactam antibiotics before they can reach PBP targets located in the cytoplasmic membrane [22, 24].

Reprogramme the Target Sructure

The possible third approach of developing resistance against antibiotics does not focus on the destruction or removal of the antibiotic but on reprogramming or masking of the target that is responsible for resistance in bacteria [22, 25].

Mechanism of Action of Spices as an Antibacterial Agent

The major phytoconstituent found in cloves is eugenol, which destroys the cell walls of microorganisms, and permeates the cytoplasmic membranes, and further inhibits the normal protein synthesis [26]. Eugenol can inhibit the production of amylase and proteases and thus causes cell wall deterioration and cell lysis [27]. Thyme is another example of a spice that contains an active compound called thymol, which has the ability to bind to the membrane protein by hydrogen bonding or hydrophobic bonding leading to a change in the permeability of the membrane. It also decreases the intracellular ATP content of E. coli and increases extracellular ATP, which disrupts the function of plasma membranes and leads to the death of bacteria [26, 28]. Cinnamon contains cinnamaldehyde as a major component or an active component, which inhibits membrane function, cell wall biosynthesis, and specific enzyme activities. Cinnamon shows antimicrobial activities in a wide range of its species, like MRSA and A. niger [29]. Garlic possesses' ability to penetrate the cellular membranes of bacteria and results in the death of different species of bacteria [30]. Fennel seeds can break the permeability of the cell membrane of S. dysenteriae and causes leakage of electrolytes, loss of reducing sugars, proteins, etc., and further causes decomposition and death of cells. Coriander has the capability to permeate bacterial cell walls, resulting in the loss of all cellular functions [31]. Black pepper causes physical changes in the cell walls of E. coli and further leads to leakage of proteins, electrolytes, ATP, and DNA materials, which results in the death of bacteria [32].

S. No	Targets	Spices and Herbs
1	Cellular Metabolism	Cinnamon, Fennel
2	Bacterial Cell wall	Clove
3	DNA and Protein synthesis	Oregano, Black pepper, Nutmeg, Rosemary, Clove
4	Cell apoptosis	Oregano
5	Affects bacterial cellular morphology	Fennel, Mustard, Galangal
6	Cell Proliferation Process	Turmeric
7	Bacterial Cell Membrane	Oregano, Turmeric, Thyme, Cumin, Ginger, Nutmeg, Galangal, Cardamom, Dill, Anise
8	Bacterial Enzymes	Nutmeg, Garlic, Mustard, Asafoetida
9	Biofilm formation	Cumin, Ginger, Cardamom
10	Energy Metabolism Process	Oregano, Black Pepper, Rosemary
11	Bacterial Respiration	Black Pepper, Basil
12	Membrane Potential	Coriander
13	Efflux Pump	Coriander
14	Synthesis of fatty acid	Rosemary
15	Leakage of essential constituents	Cinnamon, Fennel, Rosemary, Basil

ANTIFUNGAL AGENTS

A drug that selectively removes fungal cells from a host with minimal toxicity to the host is known as an antifungal agent. Targets of spices and herbs for antifungal activity are shown in Table 2.

Table 2. Targets of Spices and herbs for antifungal activity.

S. No	Targets	Spices and Herbs
1	Mycelium Growth	Cinnamon, Thyme, Cumin, Basil, Asafoetida, Anise
2	Destroys Cell Membrane	Cinnamon, Clove, Turmeric, Black Pepper, Coriander, Dill
3	Energy Metabolism	Cinnamon, Clove, Garlic
4	Ergosterol Synthesis	Cinnamon, Clove, Ginger, Thyme
5	Alters the Morphology of Fungal cells	Oregano, Fennel
6	DNA and Protein Synthesis	Black Pepper
7	Fungal Cell Proliferation	Black Pepper

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S. No	Targets	Spices and Herbs
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9	Targets Biofilm	Coriander
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11	Affects Fungal Cell integrity and leakage of constituents	Garlic, Ginger, Black Pepper, Dill, Nutmeg
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DEVELOPMENT OF RESISTANCE

Resistance to Azoles

Decrease drug concentration

The development of active efflux pumps results in decreased drug concentration at the site of action. Efflux pumps are encoded by Candida species by two gene families of transporters: multi drug resistance (MDR) genes of the major facilitators class and the Candida drug resistance (CDR) genes of the ATP binding cassette superfamily [33]. Upregulation of CDR1, CDR2, and MDR1 has been proven in azole-resistant C. albicans. Other transporter genes have been noticed in Candida species, like CdCDR1 and CdMDR1 in *Candida dubliniensis* and CgCDR1 and CgCDR2 in *C.glabrata*. While CDR gene up-regulation confers resistance to approximately all azoles, the MDR-encoded efflux pumps are limited for fluconazole [34, 35].

Target site alteration

ERG1 is the gene encoding for the target enzyme lanosterol C14a-demethylase, which prevents the binding of azoles to the enzymatic site. Mutations in ERG11 lead to the development of intrinsic resistance to fluconazole in *C. krusei*, which decreases the affinity of ERG11p to the drug [36]. More than 80 amino acid substitutions in ERG11p have been detected. Similarly, different mutation coexists in the same gene, which leads to the development of resistance against the azole class of drugs [37].

Up-regulation of the target enzyme

Target enzyme up-regulation can be attained through gene amplification, which increases the transcription rate or decreases degradation of the gene product. As compared to other mechanisms described above, this mechanism contributes less to the overall resistance burden, as previously observed in Candida species [38].

Resistance to Polyene

Defect in the ERG3 gene, which is involved in ergosterol biosynthesis, leads to the accumulation of other sterols in the fungi membrane. Therefore, polyeneresistant Candida and Cryptococcus isolates have comparatively low ergosterol content than polyene-susceptible isolates [39]. This indicates the development of resistance against polyene like amphotericin B caused by the defect in the ERG3 gene. Resistance to amphotericin B is also facilitated by increased activity of enzyme catalase, which causes decreased susceptibility to oxidative damage [40].

Resistance to Echinocandin

Fks1 is the gene of β -1, 3-D-glucan synthase complex, that accounts for synthesizing 1, 3- β -glucan chains, one of the major structural polymers of the Saccharomyces cerevisiae cell wall [41]. Mutation in the Fks1 gene, mostly at Ser645 position, leads to the reduction of glucan content of the cell wall and increase in chitin content and also activates the expression of CWP1, which encodes a glycosylphosphatidylinositol (GPI)-dependent cell wall protein. This causes echinocandin resistance in Candida species as well as in other species [42].

Resistance to Flucytosine

Flucytosine is a base pyrimidine analog that is responsible for inhibiting nucleic acid synthesis. Mutation in cytosine permease results in the impairment in cellular uptake, which causes resistance development against flucytosine [43]. Furthermore, a mutation in enzyme-like uracil phosphoribosyltransferase or cytosine deaminase results in a defect in flucytosine metabolism and leads to the development of resistance.

MECHANISM OF ACTION OF SPICES AS AN ANTIFUNGAL AGENT

Oregano contains carvacrol that binds to the sterols, which is the main component of the fungal cell wall and causes the change in the permeability for small cations. By changing the permeability of the fungal cell wall, it causes the death of Candida species [44]. In addition to this, cinnamaldehyde, a vital constituent of cinnamon, inhibits the fungal cell wall biosynthesis and causes the death of different fungal species. Furthermore, garlic has the ability to penetrate the Bacterial & Fungal Resistance

cellular membrane as well as a mitochondrial membrane which results in the death of fungi like *C. albicans* [29].

DETAILED ACCOUNT OF SOME SPICES AND HERBS HAVING ANTIBACTERIAL AND ANTIFUNGAL POTENTIAL:

Cinnamon

• **Biological source:** The biological source of cinnamon is *Cinnamomum zeylanicum* Nees, which belongs to the family Lauraceae. It comprises the dried inner bark of the shoots of trees [45, 46].



• **Phytoconstituents and phytochemistry:** The cinnamon barks mainly consist of volatile oil in the range of 0.5-1%. Along with this, it also contains mucilage and phlobatannins. In its essential oil, about 4-10% of eugenol and 60-70% of cinnamaldehyde is present, along with it some traces of phellandrene, caryophyllene, pinene, limonene, (-) linalool is also found [46]. The spicy taste and smell of cinnamon are due to the presence of cinnamaldehyde, and it appears brown due to the uptake of oxygen. The presence of a wide range of essential oils has been documented, such as cinnamyl acetate, trans-cinnamaldehyde, eugenol, terpinolene, and α -thujene L-borneol, caryophyllene oxide, L-bornyl acetate, b-caryophyllene, E-nerolidol, α -cubebene, α -terpineol [45].

• Antibacterial mode of action of cinnamon: The cinnamaldehyde causes the leakage of phosphate ions in a time and -dose-dependent manner. Exposure to cinnamon oil or cinnamaldehyde decreases the amount of intracellular ATP. Along with this, cinnamaldehyde also liberates intracellular components like proteins, nucleic acid when subjected to microbes. Due to the lipophilic nature of cinnamon essential oil, it can modify the monolayer structure of the membrane by getting incorporated into it. Also, it enhances the fluidity of the microbe's membrane by decreasing the effectiveness of packing lipid molecules [47].

• Antifungal mode of action of cinnamon: It is also active against fungi. In *Rhizopus nigricans*, cinnamon oil suppresses the growth of mycelia as well as modifies the morphology of mycelia so that flattened, empty, shriveled, and collapsed hyphae are generated. Cinnamon oil reduces the activities of succinate dehydrogenase and malate dehydrogenase enzymes in the Krebs cycle, due to which energy metabolism gets altered in *Rhizopus nigricans*. In addition to this, cinnamon oil causes the outflow of proteins and essential ions by damaging the structure of the cell membrane *via* hampering ergosterol biosynthesis. The activities of beta-1, 3-glucan, and chitin synthase get obstructed by cinnamaldehyde which affects the production of fungal cell wall components [48].

• Other uses of cinnamon: It is used as an antiseptic, flavoring agent, stimulant, and aromatic. Along with this, cinnamon bark is also used as a mild astringent, stomachic, carminative. Cinnamon is also used commercially in the preparation of candy, perfumes, and dentifrices. Chiefly cinnamon barks are used as spices and condiments [49]. Cassia cinnamon is used most frequently in diabetes [50]. In addition, the main constituents in cinnamon are responsible for numerous therapeutic potentials, which include antiviral, antifungal, anti-microbial, antiinflammatory, reducing CVS disease, cognition-enhancing, antioxidant, anticancer, gastro-protective and blood pressure, cholesterol, and lipid-lowering [51]. It is also used for prediabetes, gas (flatulence), obesity, and many other disorders, although no clear medical evidence is sufficient to support these applications. Cassia cinnamon is used as a flavoring agent in food and beverages. Recent reports have shown that cinnamon can ameliorate organ toxicity induced by chemical toxins through its antioxidant, radical scavenging, anti-inflammatory activities via modulation of numerous targets such as mitogen-activated protein kinase (MAPK), tumor necrosis factor (TNF- α), interleukin-6 (IL-6), and nuclear factor-*k*B (NF-*k*B) signalling [52].

Clove

• **Biological source:** The biological source of clove is *Eugenia caryophyllus* which belongs to the family Myrtaceae and comprises dried flower buds [49].



• **Phytoconstituents and phytochemistry:** Clove has been considered as the major source of phenolic molecules such as hydroxybenzoic acids, flavonoids, hydroxyphenyl propane, hydroxycinnamic acids, and eugenol ($C_{10}H_{12}O_2$), which is the largest biologically active molecule, and gallic acid derivatives such as hydrolysable tannins contained in the fresh plant in large amounts. In addition, clove includes flavonoids such as quercetin and kaempferol, as well as phenolic acids such as ferulic, caffeic, ellagic, and salicylic acids. Clove blossoms contain up to 18% of essential oil consisting of eugenol, eugenol acetate, and β -cariofileno. The presence of 36 elements screened by hydro-distillation in the Clove essential oil comprises eugenol, eugenyl acetate, ethyl hexanoate, calacorene, 2-heptanone, β -caryophyllene, α -humulene, humulene, and calamenene [53].

• Antibacterial mode of action of clove: Clove belongs to a class of volatile oils, and eugenol is the major chemical substance present in it. It is seen that essential oil present in clove buds exhibits antibacterial activity. It could destroy bacterial cell walls. Due to the lipophilic nature of the essential oil, it can easily interact with the cell wall. Also, it causes the liberation of intracellular materials from microbial cells, which are crucial for the survival of microorganisms; hence it results in bacterial cell death. Clove essential oil enters inside the cell by penetrating through the cytoplasmic membrane, damages the structure of the cell, and inhibits the process of DNA and protein synthesis, which are essential for bacterial growth [53].

• Antifungal mechanism of action of clove: Clove suppresses the growth of the various strains of fungi. Eugenol causes metabolic arrest in conidia. It shows the dose-dependent fungicidal action. The mode of action of clove behind fungal cell death comprises - occurrence of primary lesions on the cell membrane, which

results in indirect damage to the membrane and further leads to secondary damage. Also, eugenol hampers the ergosterol biosynthesis and affects the growth of fungi, as ergosterol is necessary for maintaining cell integrity and functions. The properties of clove essential oil, such as hydrophobicity and its ability to partition into the lipid bilayer, are accountable for its antifungal mode of action. In addition to the inhibition of dermatophytes, *Aspergillus*, and *Candida* species, the eugenol is also effective against the fluconazole-resistant *Candida albicans*, *Candida krusei*, and *Candida glabrata* [54].

• Other uses of clove: Clove is being used to give hot drinks aromatic and flavoring characteristics and is often paired with other ingredients such as lemon and sugar. The eugenol-containing clove oil is also useful for toothache and other forms of pain [49, 53]. The alcoholic and aqueous extract of clove in higher concentrations inhibit the larvae stage with a 100% mortality rate [55]. Furthermore, nanocomposite formed from clove essential oil has been used for the preservation of muscle fillets of *Thunnus thynnus* during cold storage [56]. Clove extract containing zinc nanoparticles has been used as an alternative product for its anti-cancer, anti-inflammatory, and antioxidant property [57].

Oregano

• **Biological source:** The biological source of oregano is a flowering plant *Origanum vulgare*, belonging to the mint family *i.e.*, Lamiaceae.



• **Phytoconstituents and phytochemistry:** Oregano contains essential oil. The vital component of its oil, carvacrol is a phenol, which is a natural monoterpene derivative of cymene and thymol. It also contains α - and β -pinene, linalool, myrcene, cineole, and terpinene in a small amount [58].

• Antibacterial mechanism of action of oregano: Oregano shows antibacterial action, especially against the methicillin-resistant staphylococcus aureus (MRSA) through various mechanisms. It destroys the cell membrane that leads to the leakage of small molecules such as Na⁺, K⁺. Oregano affects the normal physiological metabolism of microbial cells by inhibiting the action of intracellular enzymes like beta-galactosidase and others. It also inhibits the Krebs cycle, by which the concentrations of citric acid in MRSA increase and accumulate inside bacterial cells. In MRSA, it decreases the rate of metabolism of some amino acids and also decreases the bacterial content of fumaric acid, succinic acid, malic acid. The basic requirement for the survival of an organism is the energy that is given by ATP. Oregano shows the inhibitory effect on energy metabolism in MRSA, due to which the amount of ATP decreases, eventually reducing the activity of the Na⁺ K⁺ ATPase enzyme. The genetic material of bacterial cells gets affected by oregano as it decreases the synthesis and causes leakage of DNA and nucleic acid by destructing the cell membrane. It also obstructs the process of DNA replication, transcription, and translation. The extracellular toxin pvl gene, which is an important pathogenic factor in MRSA, is present. The oregano shows the bactericidal effect in MRSA by promoting its apoptosis, by lowering the production of the *pvl* gene [58].

• Antifungal mode of action of oregano: In addition to this, oregano essential oil suppresses the growth of fungi. It forms the shrunken hyphae and degrades them. The cytoplasm present in the cell and its organelles depletes due to the presence of essential oils in oregano. It alters the morphology of hyphae and promotes its lysis. Oregano forms crinkled and dehydrated sclerotia and also destroys it [59].

• Other uses of oregano: The antioxidants terpinene, limonene, thymol, ocimene, carvacrol, and caryophyllene give the herb its flavor and scent. Oregano is widely used as herbal medicine to treat many ailments, including skin sores, aching muscles, asthma, cramping, diarrhea, indigestion, colds, and to boost up overall health [58, 59]. *in vivo* studies have shown that oregano extract has beneficial effects on gastrointestinal health, specifically by lowering inflammation [60]. Recent studies also highlighted the anti-inflammatory potential of oregano against endotoxemia-induced toxicity [61]. Ileum transcriptomics revealed that 2% oregano extract altered gut-brain axis signaling and also altered other drug targeted gene expression [62].

Fennel

• **Biological source:** The biological source of fennel is *Foeniculum vulgare* Miller, belonging to the family Umbelliferae. Fennel comprises dried, ripe fruits [49].

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• **Phytoconstituents and phytochemistry**: The fenchone, *trans*-anethole, and limonene are the main bioactive components of fennel essential oil. The seeds of *Foeniculum vulgare contain* isolated triterpenes, flavanoid glycosides, smaller terpenes (monoterpenoids, sesquiterpenoids, and diterpenoids), and reduced sugars. The phenolics found in the fruit of this plant are ferulic acid-7--,

neochlorogenic acid, gallic acid, chlorogenic acid, p-coumaric acid, chlorogenic acid, quercetin-7-o-glucoside, ferulic acid, 1,5 di-caffeoylquinic acid, caffeic acid, quercetin, and apigenin [63].

• Antibacterial mode of action of fennel: Although a culinary spice, the antibacterial activity of fennel is also quite prominent. Because of the hydrophobic nature of fennel essential oil and its components, it can easily penetrate the bacterial cell membrane to accumulate inside it and affect the permeability of bacterial cells, which results in the leakage of intracellular ingredients like Na⁺, K⁺, Ca^{2+,} and proteins. The bacterial cell death occurs due to irreversible damage of the cytoplasmic membrane and loss of some vital molecules. It also causes morphological alterations in bacterial cells. It was seen that when bacterial cells were treated with fennel essential oil, the cells became pitted, shriveled, deformed, and got stuck to each other. After that, there occurs leaching of genetic materials and nutrients. All these effects of fennel essential oil increase with an increase in concentration and duration of treatment. Even very small modifications in the structural integrity of cell membrane can affect the metabolism of cells, and this may result in cell death [59, 63].

• Antifungal mode of action of fennel: Fennel essential oil suppresses the

growth of fungi and also damages the morphological structure of the hyphae in the same way and by the same mechanism as that of oregano essential oil. It forms shriveled hyphal aggregates and causes degenerative changes to the sclerotia [59].

Turmeric

• **Biological source:** The biological source of turmeric is *Curcuma longa* Linn, the family Zingiberaceae. Turmeric comprises dried and fresh rhizomes.

• Phytoconstituents and phytochemistry: Curcumin, demethoxy curcumin, and bisdemethoxy curcumin, classified commonly as curcuminoids (3-6 percent), are major polyphenolic components in turmeric rhizomes. Many phenolic compounds in the turmeric rhizome include 1-(4-hydroxy-3-methoxyphenyl)-7-(3, 4dihydroxyphenyl)-1, 6-heptadiene-3,5-dione and 1, 7-bis (4-hydroxyphenyl)-1,4, 6-heptatrien-3-one; 1-(4-hydroxy-3, 5-dimethoxyphenyl)-7-(4-hydrxy-3-methoxyphenyl)-(1E, 6E)-1, 6-heptadiene-3, 4-dione; 1, 5-bis (4-hydroxy-3-methoxyphenyl)-penta-(1E, 4-dien-3-one: 1-(4-hydrox-4E)-1, -3-methoxyphenyl-5-(4-hydroxyphenyl)-penta-(1E, 4E)-1, 4-dien-3-one; 1hvdroxy-1,7-bis (4-hydroxy-3-methoxyphenyl)-(6E)-6-heptene-3,5-dione. А variety of mono- and sesquiterpenes make up the pale yellow to orange-yellow volatile oil (4-6%) acquired from turmeric. Curcumenone was the name of (4 S, 5 S)-germacrone 4, 5-epoxide: bisabola 3, 10-diene 2-one; sesquiterpenes; dehydrocurdione; ar-turmerone [49, 64].

• Antibacterial mode of action of curcumin: Curcumin is active against the growth of both gram-positive and gram-negative bacteria. Almost in all bacteria, *FtsZ* (Filamenting temperature-sensitive mutant Z), which is the FtsZ protein, is necessary for the recruitment of other proteins that create a new cell wall (septum) between dividing cells and z-ring formation. Curcumin inhibits the formation of z-ring, which in turn suppresses the cytokinesis and bacterial cell proliferation through inhibiting *FtsZ* protofilaments assembly by binding into *FtsZ* proteins. The role of *FtsZ* protein in cell division resembles that of tubulin in the eukaryotic cell division. Again, curcumin increases the sensitivity of MRSA (Methicillinresistant staphylococcus aureus) towards the beta-lactam antibiotic's antibacterial action. This is done through the mechanism in which it reduces the expression of PBP-2 alpha protein by inhibiting the transcription of the *mec-A* gene. Curcumin causes the death of bacterial cells by triggering cell membrane damage and cell lysis as it has an affinity for the peptidoglycan. It shows the synergism with various antibiotics against MRSA and MSSA. Glycopeptides, cephalosporins, fluoroquinolones, aminoglycosides, and beta-lactams are the antibiotics classes with which curcumin shows synergistic activity. Along with antibacterial action,

it is also seen that curcumin decreases the resistance of oxaliplatin, 5-fluorouracil, cisplatin, and doxorubicin [65].

• Antifungal mode of action of curcumin: Curcumin also possesses antifungal activity. It induces the outflow of various essential components such as K^+ ions, proteins, *etc.* As K^+ ions are important for the survival of the cell by regulating the pH of the cell and maintaining homeostasis, due to its loss, the death of the fungal cell occurs. This outflow of K^+ ions suggests that curcumin causes the lysis of the membrane because of membrane damage, fungi losses 'their cell membrane integrity. It shows the membrane-targeted mode of action against the fungi and increases membrane permeability [49, 66].

• Other uses of turmeric: It is used as a spice or condiment and is also used commercially in ointment and creams manufacturing as a coloring agent. Turmeric is also known for having potent antioxidant and anti-inflammatory properties. As inflammation is linked to tumor growth, anti-inflammatory compounds such as curcumin may play a role in treating and preventing a variety of cancer types, including colorectal, pancreatic, prostate, breast, and gastric cancers [66].

Garlic

• **Biological source:** The biological source of garlic is *Allium sativum* Linn. belonging to the family Liliaceae. It comprises bulbs of the plant.



• **Phytoconstituents and phytochemistry:** A. Sativum bulb is believed to contain hundreds of bioactive compounds such as sulfur-containing compounds like ajoenes (E-ajoene, Z-ajoene), thiosulfinates (allicin), vinyldithins (2-vinyl-(4H)

--,3-dithiin, 3-vinyl-(4H)-1,2-dithiin), sulfides (diallyl disulfide (DADS), diallyl trisulfide (DATS) and many others that accounted for 82% of the total garlic sulfur content. The key odor molecules in freshly milled garlic homogenates are S-propyl-cysteine-sulfoxide (PCSO), allicin, and S-methyl cysteine-sulfoxide (MCSO) [67].

• Antibacterial mode of action of garlic: Garlic is used as an antibacterial and antifungal agent because of the presence of allicin. Garlic has the same antibacterial action as that of penicillin and chloramphenicol. Tuberculosis disease caused due to the *Mycobacterium tuberculosis* develops resistance against various antibiotics at a very great speed, and this can also be treated with garlic because of the Sulphur containing compound in it. In *Entamoeba histolytica*, two enzyme groups are present; these are *alcohol dehydrogenase* and *cysteine proteinase*. From these two, the enzyme which is responsible for infection is the *cysteine proteinase*, whereas *alcohol dehydrogenase* is simply important for the metabolism and survival of the microorganisms. Allicin acts by reacting with the sulfhydryl (SH) or thiol group of these enzymes and blocks both the enzymes [67, 68].

• Antifungal mode of action of garlic: When "Caposele" and "Rosato" these two different varieties of garlic were studied for their activities; it was found that Rosato had suppressed the growth of the *Penicillium expansum* while *Aspergillus versicolor* and *Penicillium citrinum* had been inhibited by the Caposel variety. The mechanism by which it shows the antifungal action involves the destruction of the structure of the cell *via* piercing or entering inside cells and their organelles, causing the leakage of essential molecules and cytoplasm. In *Candida albicans*, garlic oil alters the normal metabolic process due to which cell cycle, protein modification process, and oxidative phosphorylation also suffer [67, 68].

• Other uses of garlic: Garlic is widely used for several conditions linked to the blood system and heart. Garlic is also used by some people for the prevention of different types of cancers [60]. Also, recent studies reported the anti-obesity property of garlic oil which counteract the effects of an HFD on adipose tissue weight, body weight, and serum lipid profiles *via* preventing excessive adipogenesis [69, 70]. Garlic also possesses a neuroprotective effect *via* altering the caspase-12-dependent pathway and improving spatial acquisition learning in mouse models [71 - 73]. Garlic activates pain-sensing neurons *via* acting on transient receptor potential (TRP) family of cation channels, specifically on TRPA1 and TRPV1 through allicin, showing its involvement in mechanical and inflammatory pain and visceral hypersensitivity [74, 75].

Ginger

• **Biological source:** The biological source of ginger is *Zingiber officinale* Roscoe belonging to the family Zingiberaceae. It comprises of whole or cut and also scrapped or unscraped dried rhizomes of the ginger plant.



• **Phytoconstituents and phytochemistry:** The volatile oil of ginger is responsible for characteristic fragrance and flavor that comprise 1-3% of the weight of fresh ginger. Primarily volatile oil consists of zingerone, shogaols, and gingerol, which are phenolic ketones of oleoresin. Along with that, it contains 6-gingerol (1-[4'-hydroxy-3'-methoxyphenyl]-5-hydroxy-3-decanone), the principal pungent compound [76].

• Antibacterial mode of action of ginger: Ginger is one of the most commonly used spices and is employed in the treatment of some medical conditions. It is effective against fungi, bacteria, and viruses. It strongly inhibits the growth of Pseudomonas aeruginosavia hampering the biofilm formation process and thus affects the membrane formation. Ginger reduces the level of bis-(3'-5')-cyclic dimeric guanosine monophosphate (c-di-GMP) in Pseudomonas aeruginosa PA14 and blocks the biofilm formation. Ginger contains a wide range of active constituents, and from all these, 6-shogoal and gingerenone-A restrict the 6hydroxymethyl-7, 8-dihydropterin pyrophosphokinase activity in pathogen and inhibit the growth of Staphylococcus aureus [77]. The nanofibers from ginger had good microbial activity. Thus, many formulations have been prepared using different methods. One such formulation is transparent cellulose film prepared with chemical treatment, and ultrasonication has been shown to possess good antimicrobial properties [77]. Also. polyvinyl alcohol and ginger bionanocomposites had microbial activity thus can be used as an alternative packaging material for food [78].

• Antifungal mode of action of ginger: In fungi, ginger alters the membrane integrity by penetrating inside the cell wall and cytoplasmic membrane by making

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it permeable due to the lipophilic nature of ginger essential oil. Also, it obstructs the growth of fungi Fusarium verticillioides by means of affecting membrane integrity, reducing biosynthesis of ergosterol, and decreasing the synthesis of fumonisin-B1 and fumonisin-B2. The powerful antifungal constituents from the ginger essential oil are citral and gamma-terpinene; as they obstruct the ergosterol biosynthesis, the expression of the genes which are associated with aflatoxin biosynthesis also gets hampered, which further results in the suppression of *Aspergillus flavus* growth. In addition to this, it is also effective against the virus as it suppresses the growth of the human respiratory syncytial virus (HRSV) in the respiratory tract cell lines and causes inhibition of plaque formation. The viral attachment and internalization are also affected by ginger. It depletes the level of liver function relevant markers such as *Aspartate aminotransferase* (AST) and *Alanine aminotransferase* (ALT) also alpha fetoproteins (AFP) in the people who are suffering from hepatitis C [76].

• Other uses of ginger: Ginger has strong antibacterial activity and can also be used to treat bacterial infections. This is used in colic and atonic dyspepsia and is also used as a stimulant in traditional Chinese medicine. Ginger is described as spicy and sweet rhizome, helping to warm up the body and treat cold extremities, improving a slow and late heart, treating pale skin, and strengthening the body after loss of blood. In Traditional Chinese Medicine it is also used as herbal medicine for several cardiovascular diseases. In traditional medicine, the antiemetic effect of ginger was recognized as a treatment option. Ginger has a significant role to play in treating headaches, nausea, colds, arthritis, rheumatism, muscle pain, and inflammation. Ginger could be used as an anti-edema drug which is used to cure various ailments, which include nausea, gastrointestinal respiratory disorders, atherosclerosis, migraine. depression. disorders. gastrointestinal ulcer, cholesterol, and other ginger benefits include its use for pain reduction, and to treat rheumatoid arthritis [79]. In addition, regular consumption of ginger has been shown to delay the progression of diabetes and cancer due to the effective antioxidant action of its constituents [80]. A bioactive compound present in the ginger act by a different mechanism that shows anticancer activity, and these molecules can be used as a lead for further drug discovery [81]. Recent studies also revealed that ginger supplementation could alter the composition of gut microbiota and increase survival of Bifidobacterium genus and short-chain fatty acid-producing bacteria, thereby showing anti-obesity activity in mice [82].

Black Pepper

• Biological source: The biological source of black pepper is Piper nigrum Linn

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belonging to the family Piperaceae. It comprises of the fruits which are dried unripe fruits of climbing vine of black pepper.



• **Phytoconstituents and phytochemistry:** Piperine is an alkaloid that is commonly present in plants relating to the Piperaceae family. This is also categorized as piperidine (E, E)-1- and piperoylpiperidine (E, E)-1-[5-(1, 3-benzodioxol-5-yl)-1-oxo-2, 4-pentdienyl].Piperine, along with chavicine (a piperine isomer), is the alkaloid accountable for the pungency of long pepper and black pepper. This is also used as an insecticide and in certain types of traditional medicine. Piper nigrum leaf extract has shown the presence of tannins, alkaloids, and flavonoids [83].

• Antibacterial mode of action black pepper: Black pepper inhibits bacterial growth in various ways. It outrages the cell membrane of bacteria so that the intracellularly present enzymes like transaminase comes into the extracellular space. It elevates the permeability of the cell membrane of bacteria by restricting the activity of the Na^+/K^+ ATPase enzyme. The cellular energy metabolisms of bacteria also get affected through the inhibition of *Hexokinase* and *Pyruvate* kinase enzymes of the glycolysis. This may lead to bacterial cell death as black pepper blocks the production of essential substances and energetic components. In addition to this, black pepper obstructs the aerobic respiration, Krebs cycle, so that bacterial cell undergoes the anaerobic respiration. Due to the anaerobic respiration, the level of lactic acid increases, and it accumulates in the bacterial cell. Hence, bacterial cell death occurs due to a high concentration of lactic acid [84]. Black pepper essential oil (BPEO) has been shown to produce physical and morphological alterations in E. coli by modulating permeability of cell membrane via breaks and then leaking electrolytes, proteins, and nucleic acids, which eventually results in the death of E. coli, and thus BPEO can be used as a natural antibacterial agent in food practices [32]. Also, the biocomposite film prepared from BPEO and ginger essential oil (GEO) fabricated by casting technology has been shown to inhibit microbial growth thus can be used as alternatives to wound dressing [85].

• Antifungal mode of action of black pepper:Black pepper contains the active constituents which are extremely fat-soluble and volatile, because of these two properties it exerts antifungal action mainly through the cell membrane. By damaging the cell membrane and inhibiting its synthesis it enters inside the fungal cell, afterward, it invokes the outflow of various ions and molecules from the cell, which results in fungal cell death. In addition to this, black pepper also affects the production of polysaccharides, RNA protein, and DNA. Fungal cell proliferation and spore germination are also gets inhibited by black pepper. It provokes the rupturing of the chitin polymer, beta-1, 3-glycan, and beta-1, 6-glycan [86].

• Other uses of black pepper: Dried P. nigrum fruit is considered as "The King of the Genus" because it gives a unique taste to the dish together with its medicinal properties to treat many diseases as well. Herb is widely used for the treatment of gastrointestinal diseases, malaria, respiratory disorders, cold and cough infections, skin cancer, scabies, nerve pain, and other illnesses [87]. Also, recent data has shown that consumption of herbal formulation prepared from piperine along with turmeric extract and ginger for 1 month has beneficial effects in patients with chronic knee osteoarthritis through anti-inflammatory action *via* altering levels of prostaglandin E_2 [88]. Due to antioxidants, the anti-inflammatory activity of piperine can attenuate cardiac injury, hypertension, and cardiac fibrosis, thus showing the cardioprotective effect [89].

Coriander

• **Biological source:** The biological source of the coriander is *Coriandrum sativum* Linn belonging to the family Umbelliferae. It comprises completely dried, ripe fruits.



• **Phytoconstituents and phytochemistry:** Coriander fruits include essential oil and vegetable oil (fixed oil). Coriander contains monoterpenoid linalool, limonene, camphor, and geraniol. The main component described in most studies is linalool (60-70%), often up to 87.54%. Frequently recognized other key elements are α -pinene, camphor which geraniol, and are responsible for the character of the plant's scent and aroma [90].

• Antibacterial mode of action of coriander: In coriander essential oil, the active chemical constituent linalool shows the antibacterial action by accelerating penetrability inside the cell. Linalool, naturally present in S (+) enantiomer form, is highly penetrable to the bacterial cell membrane, which is negatively charged, *i.e.*, gram-negative bacteria. Due to its alcohol-like properties, the peptidoglycan shrinks and tightens due to dehydration of peptidoglycan. After penetrating inside the cell, coriander oil shows various secondary effects, such as alteration of membrane potential. It also works by restricting the activity of the efflux pump, which is one of the reasons responsible for antibiotic resistance [31, 91].

• Antifungal mode of action of coriander: Coriander shows the antifungal activity by causing damage to the cell membrane. Coriander essential oil binds with ergosterol which is the important component of the fungal cell wall and promotes the outflow of essential ions from the cell, which results in fungal cell death. In addition to this, coriander also affects the integrity of biofilm, which is one of the mechanisms through which microbes develop resistance. At the modest inhibitory concentration of coriander essential oil, the reduction in proteolytic activity takes place in *Candida albicans* [31].

Other uses of coriander: Coriander is useful in treating problems associated with the GI tract, nausea, loss of appetite, *etc.* It is used in the treatment of certain bacterial and fungal infections associated with joint pain. It also increases the flow of milk in breastfeeding women [31, 91].

Thyme

• **Biological source:** The biological source of the thyme is *Thymus vulgaris* Linn, belonging to the family Labiatae. It comprises leaves that may be dried or partially dried and also of flowering tops.

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• **Phytoconstituents and phytochemistry:** The chemical makeup of the thyme oil is distinguished by significant concentrations of thymol, p-cymene, and ÿ-terpinene [92].

• Antibacterial mode of action of thyme: The vital chemical constituent of the thyme essential oil is thymol which has antibacterial efficacy. It acts by rupturing the outward membrane of the gram-negative bacteria. Because of the ruptured cell membrane, the enhancement in the outflow of ATP and liberation of lipopolysaccharides also occurs. This could result in a disturbance in plasma membrane function [93].

• Antifungal mode of action thyme: Thyme suppresses the growth of fungi by interacting with ergosterol and affects the normal function and growth of fungal cell membrane. In addition to this, sporangiospores germination and development of mycelia get obstructed because of thymol. It acts as both fungistatic and fungicidal [94].

• Other uses of thyme: Thyme, along with other herbs, is used for various ailments of the throat such as bronchitis, whooping cough, and sore throat. Thyme is also taken in dyspraxia (a movement disorder in children), GIT, and skin disorders. It is also used as an appetite stimulant. Other than these, thyme is also used in laryngitis, tonsillitis, sore mouth, and bad breath. In foods industries, thyme is used as a flavoring agent [49, 94].

Cumin

• **Biological source:** The biological source of cumin is the *Cuminum cyminum* belonging to the family Umbelliferae. It comprises dried, ripe fruits of the cumin plant.



• **Phytoconstituent and phytochemistry:** Cumin seeds are extremely nutritious. The predominant volatile components of cumin are cumin aldehyde, cymene, and terpenoids. Cumin has a strong, distinctive flavor. Its warm fragrance is a consequence of its essential oil content. Cuminaldehyde and cuminic alcohol are the chief components of the aromatic compound. The substituted pyrazines, 2-methoxy-3-methyl pyrazine, and 2-methoxy-3-sec-butylpyrazine2-et-oxy-3-isopropylpyrazine are other important aroma compounds of roasted cumin. Certain constituents include terpinene, saffron, p-cymene, and β -pinene [95].

• Antibacterial mode of action of cumin: Cumin is effective against both gram bacteria. It acts by targeting biofilm and decreases its production. Also, it causes the liberation of proteins and DNA from the cells of bacteria by destructing the cell membrane [96].

• Antifungal mode of action cumin: Cumin essential oil is also effective against fungi. It inhibits the growth of *Aspergillus flavus*, as cumin has anti-aflatoxigenic activity. Cumin essential oil inhibits mycelial growth completely. It decreases the expression of the fumonisin biosynthetic gene in *Fusarium verticillioides* which is significant for the FUM1 gene [97].

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• Other uses of cumin: The cumin plant seeds are used for indigestion problems, as a diuretic, as an aphrodisiac. Cumin is used as a flavoring agent in the food industry and as a fragrance in cosmetics [49, 97].

Nutmeg

• **Biological source:** The biological surface of the nutmeg is *Myristica fragrans* Houtt. of family Myristicaceae. It comprises dried kernels of nutmeg seeds.



•Phytoconstituents and phytochemistry: The volatile oil portion of nutmeg comprise phenylpropanoids and terpenes, including d-borneol, l-terpineol, d-pinene, geraniol, limonene, safrol, and myristicin. Myristicin in its pure form acts as a poison and is toxic, and the administration of higher amounts of nutmeg can develop myristicin poisoning. The oil is colorless or may be light yellow, and smells and tastes are similar to that of the nutmeg [98].

• Antibacterial mode of action of nutmeg: The gram-negative bacteria are less sensitive to the antibacterial action of nutmeg. This contrast inactivity towards these types of bacteria is due to the difference in their composition of the cell wall. The gram-negative bacteria contain many proteins, lipoproteins, and less concentration of peptidoglycan in its cell membrane than gram-positive bacteria, which reduces their susceptibility towards nutmeg essential oil [98, 99].

• Antifungal mode of action of nutmeg: Nutmeg essential oil is also used as an antifungal agent. Its antifungal mode of action is that it causes the outflow of ions and molecules from cells by increasing the porosity of the cell membrane. The multiple cellular processes of fungi get inhibited by nutmeg essential oil [98].

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• Other uses of nutmeg: Nutmeg is used for GIT disorders and nausea. It is also used to treat insomnia, different types of cancer, and kidney disorder; increase menstrual flow, causing a miscarriage; nutmeg is used as spice and flavoring agent. Nutmeg oil is used in soaps and cosmetics as a fragrance [49, 99].

Rosemary

• **Biological source:** It is a woody, perennial herb with fragrant, evergreen, needle-like leaves and white, pink, purple, or blue flowers belonging to the plant *Rosmarinus officinalis,* family-Labiatae.



• **Phytoconstituents and phytochemistry:** Rosemary plants have a polyphenolic profile, distinguished by the presence of carnosol, carnosic acid, and the principal components are rosmarinic acid and hesperidin. The essential oil of rosemary consists primarily of 1, 8-cineole (46.4%), camphor (11.4%), and pinene. The *Rosmarinus officinalis* is a rich source of phenolic compounds. Rosemary extracts also contain many antioxidants, in addition to the volatile constituents, that mostly belong to the class of phenolic acids, flavonoids, and diterpenoids [100].

• Antibacterial mode of action of rosemary: Rosemary's inhibitory effect results from the action of carnosol, rosmaridiphenol; rosmarinic acid, carnosic acid, rosmanol, epirosmanol, and isorosmanol. They alter the transportation of electrons, cellular leakage components, and changes in fatty acid production [79].

• Antifungal mode of action of rosemary: High proportion of antifungal activity of rosemary oil is correlated with monoterpenes α -pinene as a principle compound. The essential oil from rosemary has a medium anticandidal action. Aflatoxin B1 (AFB1) is potentially a carcinogenic and poisonous metabolite made by the species

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Aspergillus and rosemary acts by strongly inhibiting the aflatoxin synthesis. The antifungal effects of Rosemary essential oil can be due to monoterpenes composite, and especially α -Pinene [79].

• Other uses of rosemary: The rosemary leaf and its oil are used for making medicine. Rosemary is used to improve memory, indigestion (dyspepsia), joint pain associated with arthritis, hair loss, and other disorders, although there is no clear medical evidence to support any of these applications. The rosemary is used as a seasoning in foods. Rosemary oil is used as a fragrant component in the manufacture of soaps and perfumes [100].

Mustard

• **Biological source:** The mustard consists of the dried ripe seeds of the plant *Brassica nigra* belonging to the family Cruciferae.



• **Phytoconstituents and phytochemistry:** Mustard seeds have the compound allyl isothiocyanate (AIT) as their primary pungency ingredient. Isothiocyanates (R - N = C = S) are glucosinolate derivatives in Cruciferae plant cells or the mustard family. Such compounds are formed through the action of the enzyme myrosinase glucosinolates as it injures or mechanically disrupts the plant tissue. Certain side groups of isothiocyanate include ethyl, methyl, benzyl, and phenyl, in addition to the allyl side group. Such compounds were known as potent antimicrobial agents [101].

• Antibacterial mode of action of mustard: The isothiocyanates inhibit bacterial cells may be due to enzyme inhibition by direct reaction with disulfide bonds or by anion reaction with thiocyanate (SCN-) to mollify sulfhydryl enzymes [101,

102]. Also, it can be used as a probiotic as it enhances the growth of some microbes except E. coli and thus can be used in livestock nutrition [103]. It can be used for antimicrobial packaging based on its release of allyl isothiocyanate from its seed in a controlled manner [104].

• Antifungal mode of action of mustard: Mustard essential oil significantly causes the fungal degradation of shelled groundnuts. Mustard essential oil inhibits growth in molds [101, 105].

• Other uses of mustard: Black mustard oil is used to treat severe cold, sore and swollen joints (rheumatism) and arthritis. Black mustard leaves are used in foodstuffs. Black mustard seed is also used as a seasoning in foods and for flavoring mustard condiments. There are about 40 distinct mustard plant species. The mustard condiment is usually made of three different varieties. The most pungent one is the black mustard (*Brassica nigra*) [49].

• The high (200 mg/kg) doses of the crude aqueous extract of the black seeds produce neuronal insult at prefrontal cortex microarchitecture along with alteration of the expression and activities of certain enzymes in the brain due to the production of oxidative stress [106]. Also, mustard seed extract shows anticancer activity against lung cancer *via* stimulation and regulation of proliferation, apoptosis, cell and DNA damage, and invasion [107].

Basil

• **Biological source:** It consists of the flowering shoots of the plant *Ocimum bacsilicum* belonging to the family- Labiatae.



• **Phytoconstituents and phytochemistry:** The essential European basil oil contains high linalool and methyl chavicol (estragole) concentrations in a ratio of about 3:1. 1,8-cineole, eugenol, and myrcene are some of the constituents. The sweet basil clove-scent is derived from eugenol. Basil's fragrance profile contains 1, 8-cineole, and methyl eugenol [108].

• Antibacterial mode of action of basil: It was found that the essential oil of *O. bacsilicum* has moderate antibacterial activity. Bacterial strains considered to be gram-positive are more responsive to basic basil oils as compared to gram-negative bacteria. The inhibition may be accompanied due to enhanced plasma membrane permeability by essential oils and restriction of bacterial respiration. This causes bacterial cell death because of the massive leaching of ions [108].

• Antifungal mode of action of basil: The mycelial development of the pathogenic fungus *Botrytis fabae* was substantially decreased by the chemotype oil of methyl chavicol and the linalool. All the components of the oil reduce fungal growth. *Botrytis fabae* and the rust fungus *Uromyces fabae* were also managed *in vivo*, with all oils of both chemotypes and pure methyl chavicol and linalool, greatly reducing the infection of broad bean leaves [109].

• Other uses of basil: The parts of the plant rising above the ground are used for medicinal purposes. Basil is widely used for problems with the stomach, such as spasms, lack of appetite, digestive gas, vomiting, constipation, and many other disorders, although there is no sufficient medical evidence to support this usage. Basil is used for flavor in foods [108, 109].

Asafoetida

• **Biological source:** Asafoetida is an oleo-gum resin obtained as an exudation by incision of the decapitated rhizome and roots of *Ferula asafoetida* L, *F. foetida, Royel, F. rubricaulis* Boiss, and some other species of Ferula, family-Apiaceae.



• **Phytoconstituents and phytochemistry**: Asafoetida comprises volatile oil, resin, and gum. Owing to the presence of sulfur compounds, it has the oil's garlic-like odor. The resin has three sulfur compounds extracted, such as 1-methyl propyl-1-propenyl disulfide, l-(methylthio)-propyl-1-propenyl disulfide, and l-methyl-propyl 3-(methylthio)-2-propenyl disulfide; the latter two have pesticidal properties. Resin is made up of asaresinotannol ester and ferulic acid, pinene, vanillin and ferulic acid free [110].

• Antibacterial mode of action of asafoetida: The preliminary phytochemical analysis of asafoetida extracts showed the presence of terpenoids, tannins, glycosides, alkaloids, flavonoids, and polyphenolic compounds. Tannins inhibit microbial enzymes in distilled forms. It is reported that the astringent property of tannins is due to their complex formation with enzymes and metal ions. Polyphenolic components are found to have antimicrobial activity probably due to suppression of enzymes in the oxidized forms or *via* more nonspecific protein interactions. Many specific secondary plant metabolites are believed to have an antibacterial effect [110].

• Antifungal mode of action of asafoetida: Asafoetida oil has shown inhibitory activity against all fungal strains, including *Aspergillus Niger*, *Candida blanki*, *Candida krusei*, *Candida glabrata*, *Candida cylindracea*, *Candida tropicalis*, *Candida Albicans*, and *Saccharomyces cerevisiae*, but it shows heavy activity against *Candida Tropicalis*, and the MTCC-227 *Candida albicans*, *Saccharomyces Cerevisiae*, *A. Niger while mild to C. blanki*, *C. cylindracea*, *C. glabrata*, *C. krusei*, and *C. albicans*. The antifungal and allelopathic effects of methanol extract from concentrations of asafoetida oleo-gum resin against Pleurotus spp. and the harzianum and trichoderma were examined. It exhibited fungicidal activity against *T. Pleurotus* and *harzianum* species at greater concentrations. Asafoetida oleo-gum-resin imposes a semispecific antifungal action on the growth of *T. harzianum mycelium* at the minimum concentrations. The inhibition of mycelium production of the asafoetida oleo gum resin towards the fungi has also been reported [111].

• Other uses of asafoetida: People use asafoetida resin, a substance similar to gum, as a medicine. Asafoetida has been used by women to restart their menstrual cycles after menstruation has stopped for some reason, for conditions such as breathing or throat problems, digestive problems. Asafoetida is used in the production as a fragrance in cosmetics and as a flavoring agent in foods and beverages. Asafoetida is also used in materials for the repelling of dogs, cats, and wildlife [49, 111].

Galangal

• **Biological source:** Galangal consists of a rhizome used as an herb, *Alpinia galangal* belonging to the family Zingiberaceae.



• **Phytoconstituents and phytochemistry:** The chemical constituents of the different parts of the medicinal plant Galangal (*Alpinia galanga*) are Monoterpene and sesquiterpene derivatives. β -farnesene and β -caryophyllene (40.5% leaf oil) along with fenchyl acetate (20.7% leaf oil), 1, 8-cineole, and caryophyllene oxide are the main constituents of leaf oils. The key constituent of stem oil is cubenol followed by humulene, germacrene, and cadine. Galangal rhizome oil contains cineole, carotol, 1–8, fenchyl acetate, and rhizome oil contains 1–8 cineole, β -pinene, β -caryophyllene, camphor, methyl cinnamate, fenchyl acetate, limonene, camphor, α -terpineol, and cubenol. The root oil with 1, 8 cineole, and limonene are found to have fenchyl acetate [112].

• Antibacterial mode of action of galangal: The galangal extract has the most potent inhibitory effect on *S. aureus*. The galangal extract damages the outer and the inner membrane and causes coagulation of the cytoplasm. Destruction of the cytoplasmic membrane properties causes the discharge of cellular materials, namely nucleic acid, that results in bacterial cell death [113].

• Antifungal mode of action of galangal: (E)-8 beta, 17-epoxylabd-12-ene-15, 16-dial is a bioactive component from galangal which lyses the *Candida albicans* protoplast. These findings reveal that antifungal action of (E)-8 beta, 17-epoxylabd-12-ene-15, 16-dial is owing to a shift in membrane porosity caused by the variation of membrane lipids [114].

• Other uses of galangal: Alpinia is a ginger-based herb. The rhizome (underground stem) is used to produce the drug. For fevers, muscle spasms,

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digestive gas, bacterial infections, and swelling (inflammation), people have been using Alpinia, although there is no clear medical evidence to support such uses [113, 114].

Cardamom

• **Biological source:** Cardamom comprises the dried, nearly ripe fruits of the plant *Elettaria cardamomum* Var. Minuscule belonging to the family Zingiberaceae. The cardamom seeds contain about 3-6% of the volatile oil.



• **Phytoconstituents and phytochemistry:** The composition of essential oil in the cardamom seeds relies greatly on storage conditions, and it can be as high as 8%. In the crude oil, 45% of α -terpineol was found; others include myrcene, 1, 8-cineol, menthone, β -phellandrene, sabinene, limonene, and heptane [115].

• Antibacterial mode of action of cardamom: Cardamom essential oil inhibits growth and triggers dissociation of biofilms in the *Streptococcus mutans*, and it is noted that it could be of interest to periodontal disease. Extracts of cardamom demonstrated good antibacterial activity against major gram-negative periodontal pathogens, including *Aggregatibacter actinomycetemcomitans*, *Fusobacterium nucleatum*, *Porphyromonas gingivalis*, *intermedia*, and *Prevotella intermedia*. Bacterial membrane disturbance was caused by CFE and CSE, affecting the membrane integrity of bacteria. These findings are consistent with previous research that bacteria treated with 1, 8-cineole, one of its key components of cardamom extracts, has shown shrinking cells and weakened membranes [115].

• Antifungal mode of action of cardamom: The key component 1, 8-cineole, substantially increased elastase and protease output and dose-dependent motility in *Pseudomonas aeruginosa* PAO1. About 50% of essential oils suppressed elastolytic and proteolytic activity in *P. aeruginosa* PAO1. These oils had also hindered the synthesis of violacein in C. A strain of the violaceum.

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Similarly, in large concentrations, the A. corrorima essential oil greatly prevented radish germination [116].

• **Other uses of cardamom:** The cardamom seeds and seed oil are used for preparing medicine. Cardamom is often used in people who consume little to no alcohol (nonalcoholic fatty liver disease) and diabetes. Cardamom is used as a seasoning in foods. It is used even in soaps, creams, and scents [49].

Dill

• **Biological source:** Dill consists of the dried ripe fruits of *Anethum graveolens* Linn., belonging to the family Umbelliferae.



• **Phytoconstituents and phytochemistry:** The chemical constituents of the volatile dill oil differed according to the parts of the plant. The α -phellandrene and limonene percentage differed in leaves and flowers. Dill is available in either leaves or flowers but is not present in fruit oil. Carvone and limonene are the essential elements of the dill fruit oil, while α -phellandrene occupies the herb oil. Dill apiole and myristicin were other key components of herbal oil. Contents and compositions of dill fruit oil differed with the extraction process [117].

• Antibacterial mode of dill: The extract obtained containing carvone as a major component from dill suppresses the strains but is significantly less effective than D-limonene. Elevated membrane permeability of tested microorganisms by exposure to essential oil may be related to proteolytic cleavage and bacterial death induced due to intracellular component leakage, notably electrolyte losses like K⁺,

Ca^{2+,} and Na⁺. The release may be due to the interaction of antimicrobials and the cytoplasmic membrane. A decline in cellular components such as proteins and other important molecules suggests injury to the cytoplasmic membrane [118].

• Antifungal mode of action of dill: The antifungal destinations for dill oil are the plasma membrane and the mitochondria. Dill oil shows dose-dependent antifungal activity; the specific harm to the plasma membrane occurs in *A. flavus* cells with a heavy lesion on the membrane, instead of metabolic disability, which destroys the secondary membrane. Dill oil can modify the architecture of the plasma membrane, suggesting that the fungal cells lack integrity. Dill oil can produce substantial ergosterol biosynthesis inability with *Aspergillus flavus* [119].

• Other uses of dill: Dill is an herb that is used in soaps and cosmetics, cooking spice, and a fragrance. People also use dill seeds and plant parts that grow just above the ground as medicine. Individuals use dill for issues with digestion, liver disease, urinary tract disorders, infections, and several other illnesses, although there is no clear medical evidence to support such uses [29, 119].

Anise

• **Biological source**: Anise consists of dried ripe fruits of *Illicium verum*, *Pimpinella anisum* Linn., belonging to the family Umbelliferae.



• **Phytoconstituents and phytochemistry:** Anethole is the main constituent of anise. Other components comprise p-anisaldehyde, 2-methylbutyrate, Y-himachalene, cis-pseudoisoeugenyl methylchavicol, and trans-pseudoisoeugenyl 2-methylbutyrate. Flavonoids separated from the anise comprise of quercetin 3-glucuronide, rutin, 7-glucoside luteoline, isoorientin, isovitexin, 7-glucoside apigenin and luteolin glycoside. Sesquiterpenes segregated from the essential anise oil (fruit and shoots) entail gamma-himachalene and neophytadiene diterpene [120].

• Antibacterial mode of action of anise: Phenolic substances function as antimicrobials through various mechanisms involving microbial membrane disruption. The exposure of the bacteria to anise extracts has shown that Grampositive bacteria appear to be more sensitive than Gram-negative bacteria ones. Such action may be due to variations in the arrangements in their cell walls. Gram-negative bacteria possess an additional external hydrophilic membrane that works as a protective shield against hydrophobic substances and prohibits phenolic compounds from accumulating in the goal cell membrane. It usually makes the Gram-negative highly resistant to herb extracts than that the grampositive bacteria. Anise is a potential antimicrobial for gram-positive and negative bacteria, filamentous fungi, and yeast [121].

• Antifungal mode of action of anise: The key component of the oil extracted act against spore germination of *M. oryzae* is trans-anethole. Anise also hinders radial mycelial development. The suppression of spore germination is similar to that for mycelial development reduction [96].

• Other uses of anise: Anise is a flowering plant. Seed and oil are used for preparing medicines. Anise is being used to treat stomach upset, intestinal gas, nasal congestion, and increase productive cough as an expectorant. It is often used to boost urination and to boost the desire to eat. Women take anise while nursing to improve milk flow, start the menstrual cycle, alleviate menstrual discomfort or pain, ease labor and boost sexual desire. Other applications involve epilepsy control, nicotine dependency, sleeping disorder (insomnia), asthma, diabetes, and constipation. Some people treat lice, scabies, and psoriasis by applying anise directly to the skin. Anise is used as aromatherapy for nausea. Anise is used as a flavoring agent in foods. It has a sweet, spicy taste which is similar to the taste of black liqueur. It is most commonly used in liqueurs and alcohols, such as anisette and ouzo. Anise is also used in dairy products, gelatins, foods, candy, and fresheners for breathing. Anise in soap, creams, perfumes, and sachets is often used as a scent in production (Table 3) [122, 123].

USE OF HERBS AND SPICES AGAINST CORONAVIRUS

Since December 2019, the novel coronavirus (COVID-19) has spread over the world and has infected more than 2 million people causing more than 100 thousand deaths. People with pre-existing infectious and non-infectious diseases of the lungs are prone to be at more risk of contracting COVID-19 infection owing to their compromised immune system. On the basis of Ayurvedic and scientific literature, the Ministry of AYUSH (Ayurveda, Yoga and Naturopathy, Unani, Siddha, and Homeopathy), India, has issued an advisory in which it has suggested the use of Kadha, *i.e.*, herbal tea or decoction made from *Ocimum*

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tenuiorum (Tulsi), *Piper nigrum* (Kalimirch), *Cinnamomum verum* (Dalchini), *Vitis vinifera* (Munakka) and *Zingiber ocinale* (Shunthi) as a preventive measure which enhances immunity against severe infection caused by COVID-19. It has also been recommended to drink golden milk which is made up of a half teaspoon of turmeric (*Curcuma longa*) powder in 150 ml hot milk once or twice daily [142].

	Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
1	Cinnamon	<i>Cinnamomum</i> <i>zeylanicum Nees.</i> Family-Lauraceae	Cinnamaldehyde $\downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow$ Cinnamate $\downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow$ Cinnamic acid $\downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow$	bacterial agent: disturbance in cellular metabolism, leakage of cellular constituents.	stimulant, aromatic, astringent, stomachic, carminative, preparation of candy, perfumes, dentifrices, spices, and condiments.	

Table 3. Summarization of commonly used antibacterial and antifungal spices and herbs.

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S. No	Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
2	Clove	Eugenia caryophyllus Family- Myrtaceae	Eugenol \downarrow_{HO}° Eugenol acetate \downarrow_{O}° \downarrow_{O}	*As an antibacterial agent: destroy bacterial cell wall, inhibition of DNA and protein synthesis. *As an antifungal agent: metabolic arrest, damage to the membrane hampers the ergosterol biosynthesis.	Flavoring agent, as a spice, repellent for ant, for toothache.	[124]
3	Oregano	Origanum vulgare Family- Lamiaceae	Carvacrol $\downarrow \qquad \alpha$ and β pinene, Linalool $H_{0} \rightarrow H_{3}$ $H_{3} \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow G \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow G \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow G \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow G \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow G \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow G \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow G \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow G \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow G \rightarrow H_{2}$ $H_{3} \rightarrow G \rightarrow H_{3}$ $H_{3} \rightarrow H_{3}$ H_{3	antibacterial agent: destroys cell membrane, affects the normal physiological metabolism, inhibits the Krebs cycle,	Bleeding disorders, Wound healing, Asthma, Bronchitis, Cough, Flu, Painful menstrual periods, Arthritis, Headaches, Cardiovascular disorders, Antioxidant, as an insect repellent	[125]

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S. No	Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
4	Fennel	Foeniculum vulgare Miller Family- Umbelliferae	trans-Anethole Limonene H_{3C} Fenchone H_{3C} H_{3}	antibacterial agent: leakage of intracellular ingredients, morphological alterations,	•	[126]
5	Turmeric	<i>Curcuma longa Linn.</i> Family- Zingiberaceae	Curcuminoids, which is a mixture of curcumin, demethoxycurcumin, and bisdemethoxycurcumin $ \underset{\mu \in f \in f \in f \in G^{H}}{ (f \in G^{H})^{H}} Curcuminoids$	antibacterial agent: suppresses cytokinesis and bacterial cell proliferation, cell membrane damage. *As an antifungal	condiment, as a colouring agent, potent anti- inflammatory and antioxidant, Alzheimer's disease, depression, preventing a variety	[127]
6	Garlic	<i>Allium sativum Linn.</i> Family- Liliaceae	diallyl thiosulfonate(allicin)	*As an antibacterial agent: block the enzymes. *As an antifungal agent: destroys the structure of the cell, alters the normal metabolic process.	high cholesterol, heart attack, coronary heart disease, hypertension, as stimulant, expectorant,	[67]

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S.	3) cont Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
7	Ginger	Zingiber officinale Family- Zingiberaceae	Zingerone $H_{O} = \int_{O}^{O} H_{O}$ Shogaols $H_{O} = \int_{O}^{O} H_{O}$ Gingerol $H_{O} = \int_{O}^{O} H_{O}$	antifungal agent: blocks the biofilm formation, affects the membrane formation,	bacterial infections, as a stimulant, an antiemetic agent, headaches, nausea, colds, arthritis, rheumatism, muscle pain, and inflammation, as an anti-edema, as an antioxidant.	[128]

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No s	Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
	Black pepper	Piper nigrum Linn. Family- Piperaceae	Piperine	antibacterial agent: inhibits the process of protein synthesis, affects energy metabolism, reduces ATP biosynthesis, and obstructs aerobic respiration. *As an antifungal agent: damages	nerve pain, as anti- apoptotic, anti- microbial, anti- pyretic, anti- analgesic, anti- tumor, anti- depressant, anti- inflammatory, anti- arthritic, anti- thyroid, antifibrinolytic, anti-fungal, anti- diarrheal, immunomodulatory,	[129]

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S. No	Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
9	Coriander	Coriandrum sativum Linn. Family- Umbelliferae	Linalool $H_{H_{2}}^{H_{3}} H_{3}^{H_{3}} H_{4}^{H_{4}} H_{2}^{H_{4}}$ $H_{3}^{H_{3}} H_{3}^{H_{4}} H_{4}^{H_{4}} H_{2}^{H_{4}}$ Limonene $G_{H_{3}}^{H_{3}} H_{3}^{H_{4}} H_{4}^{H_{4}}$ Camphor $A_{H_{3}}^{H_{3}} H_{4}^{H_{4}} H_{4}^{H_{4}}$ Geraniol $A_{H_{3}}^{H_{3}} H_{4}^{H_{4}} H_{4}^{H_{4}}$	antibacterial agent: alters the membrane potential, restricts the activity of efflux pump. *As an antifungal agent: causes	hemorrhoids, toothaches, worms, and joint pain, as well as infections caused by bacteria and fungus, as a culinary spice and to prevent food poisoning, flavoring	
10	Thyme	Thymus vulgaris Linn. Family- Labiatae	Thymol	antibacterial agent: ruptures the outward membrane. *As an antifungal agent: interacts with ergosterol and affects the normal function and growth of	stomach, gastritis, diarrhoea, bedwetting, dyspraxia, intestinal gas (flatulence), parasitic worm infections, and skin disorders, as a diuretic, flavoring agent.	[131]

122 The Chemistry Inside Spices and Herbs, Vol. 1 (Table 3) cont.....

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(Table	3) cont				i	ı
S. No	Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
11	Cumin	<i>Cuminum</i> <i>cyminum</i> , Family- Umbelliferae	Cuminaldehyde V Cymene H_3C CH_3 CH_3 Terpenoids	antibacterial agent : targets biofilm and	Digestion problems including diarrhoea, colic, bowel spasms, gas, as a diuretic; to start menstruation, as an aphrodisiac, as a flavoring component.	[132]
12	Nutmeg	<i>Myrestica fragrans Houtt.</i> Family- Myristicae	Myristicin $\downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow \downarrow$ d-pinene Limonene $\downarrow \downarrow \downarrow \downarrow \downarrow$ $H_3C \leftarrow CH_2$ d-borneol, 1-terpineol, Geraniol, Safrol	antibacterial agent: damages the cell	mouth sores, and toothache, like spices and	[133]

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S. No	Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
13	Rosemary	<i>Rosmarinus</i> officinalis, Family-Labiatae	Carnosic acid $H_{HOOC} \rightarrow H_{HOOC} \rightarrow H_{HO$	antibacterial agent: changes in genetic	pain associated with arthritis, hair loss, as a seasoning in foods, as a fragrant	[134]
14	Mustard	<i>Brassica nigra</i> Family- Cruciferae	Allyl isothiocyanate	antibacterial agent: targets the active enzyme, promotes the misfolded protein aggregation. *As an antifungal	Antineoplastic drugs, as a food flavoring, as an emetic, and diuretic, topical treatment, arthritis, and rheumatism. It has shown encouraging pharmacological effects in cancer, cardiovascular disease, and diabetes.	[135]

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S. No		Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
15	Basil	<i>Ocimum</i> <i>bacsilicum</i> Family- Labiatae	Linalool $H_{2C}^{CH_3}$ $H_{3C}^{H_3}$ OH $H_{2C}^{CH_3}$ $H_{3C}^{CH_2}$ $H_{3C}^{CH_3}$ $H_{3C}^{CH_2}$ methyl chavicol (estragole) $CH_{3O}^{CH_3}$ Eugenol O H_{0}^{O}	antibacterial agent: restriction of bacterial respiration	Spasms, lack of appetite, digestive gas, vomiting, constipation, flavor in foods.	[136]
16	Asafoetida	<i>Ferula asafoetida,</i> Family- Apiaceae		antibacterial agent: inhibit several microbial enzymes. *As an antifungal agent:	Breathing or throat problems, digestive problems, for corns and calluses, as a fragrance in cosmetics and as a flavoring agent in foods and beverages, used in materials for the repelling of dogs, cats, and wildlife.	[137]
17	Galangal	<i>Alpinia galangal</i> Family- Zingiberaceae	Fenchyl acetate Fenchyl acetate $\mu_{1,C}$ $\mu_{2,C}$ β -caryophyllene $\mu_{2,C}$ $\mu_{2,C}$ $\mu_{3,3}$ β -farnesene, Caryophyllene oxide, and 1,8-cineole	antibacterial agent: damage to the outer and	infections, and swelling (inflammation)	[138]

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S. No	Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
18	Cardamom	Elettaria cardamomum Var. minuscule Family- Zingiberaceae	a-terpineol	antibacterial agent: inhibits growth and triggers dissociation of biofilms, affects the membrane integrity. *As an antifungal agent:	Diabetes, and high cholesterol, as a seasoning in foods, in soaps, creams, and scents.	[139]
19	Dill	Anethum graveolens Linn. Family- Umbelliferae	Carvone $\begin{array}{c} \circ \hspace{-0.1cm} + \hspace{-0.1cm} + \hspace{-0.1cm} + \hspace{-0.1cm} + \hspace{-0.1cm} - \hspace{-0.1cm} \circ \hspace{-0.1cm} + \hspace{-0.1cm} + \hspace{-0.1cm} + \hspace{-0.1cm} - \hspace{-0.1cm} \circ \hspace{-0.1cm} + \hspace{-0.1cm} + \hspace{-0.1cm} + \hspace{-0.1cm} - $	*As an antibacterial agent: Rise in the membrane permeability, destroys membrane integrity. *As an antifungal agent: modify the architecture of the plasma membrane, inhibit mitochondrial ATPase, and hinder Mitochondrial dehydrogenase functions, Deposition of ROS.	cosmetics, cooking spice, as a fragrance,	[140]

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	Name of spices / herbs	Biological Source	Phyto-constituents	Mechanism of Action	Uses	References
20	Anise	Illicium verum, Pimpinella anisum Family- Umbelliferae.	Anethole H ₃ CO	antibacterialagent:microbialmembranedisruption.*Asantifungal	Stomach upset, intestinal gas, nasal congestion, as an expectorant, to boost urination, alleviate menstrual discomfort or pain, ease labour and boost sexual desire, epilepsy control, nicotine dependency, sleeping disorder (insomnia), asthma, diabetes, constipation, flavoring agent, in dairy products, gelatins, foods, candy and fresheners for breathing.	

CONCLUSION

Bacterial and fungal infections have been a common reason for several diseases in humans. Thus, a lot of antibiotics have been employed for treatment purposes. However, the overuse of antibiotics has led to the emergence of resistant strains of bacteria and fungi. To combat/reduce this problem, some of the spices and herbs can be really helpful. In this chapter, we have discussed the various reasons for the development of resistant strains in bacteria and fungi. Furthermore, the details of various spices and herbs that can be beneficial in the treatment of infections and different possible mechanisms by which they act are described.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The authors declared that there is no conflict of interest.

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REFERENCES

- [1] Shelef LA. Antimicrobial effects of spices. J Food Saf 1984; 6(1): 29-44. [http://dx.doi.org/10.1111/j.1745-4565.1984.tb00477.x]
- Zaika LL. Spices and herbs: their antimicrobial activity and its determination. J Food Saf 1988; 9(2): 97-118.
- [3] Baytop T. Health treatment in Turkey using plant extracts [Internet]. Publication of the Istanbul University. 1984 [cited 2021 Mar 16]; 3255 .https://scholar.google.com/scholar?hl=en&as_sdt =0%2C5&q =ealth+treatment+ in+Turkey+using+ plant+ extracts&btnG=
- Hamburger M, Hostettmann K. 7. Bioactivity in plants: the link between phytochemistry and medicine. Phytochemistry 1991; 30(12): 3864-74.
 [http://dx.doi.org/10.1016/0031-9422(91)83425-K]
- [5] Deans SG, Ritchie G. Antibacterial properties of plant essential oils. Int J Food Microbiol 1987; 5(2): 165-80.
 [1] Hard Hard and Control 101 (2016) 1605 (27) 00024 11

[http://dx.doi.org/10.1016/0168-1605(87)90034-1]

- [6] Hirasa K, Takemasa M. Spice science and technology [Internet]. 1998 [cited 2021 Mar 16]. https://books.google.com/books?hl=en&lr=&id=jCYovj2TIa8C&oi=fnd&pg=PP11&dq=%5B6 %5D%09Hirasa+K,+Takemasa+M.+Spice+science+and+technology.+CRC+Press+1998.&ots=qZ0B GsJK9Y&sig=86TJocW7OR4zGulqkRuQdBeWiNQ
- [7] Nakatani N. Antioxidative and antimicrobial constituents of herbs and spices 1994. https://ci.nii.ac.jp/naid/ 10018787936/
- [8] Miladi H, Zmantar T, Chaabouni Y, *et al.* Antibacterial and efflux pump inhibitors of thymol and carvacrol against food-borne pathogens. Microb Pathog 2016; 99: 95-100. [http://dx.doi.org/10.1016/j.micpath.2016.08.008] [PMID: 27521228]
- [9] Brul S, Coote P. Preservative agents in foods: Mode of action and microbial resistance mechanisms. Vol. 50, International Journal of Food Microbiology. Elsevier; 1999. p. 1–17.
- [10] De Souza EL, Stamford TLM, De Oliveira Lima E, Trajano VN, Barbosa Filho JM. Antimicrobial effectiveness of spices: An approach for use in food conservation systems. Braz Arch Biol Technol 2005; 48(4): 549-58.http://www.scielo.br/scielo.php?script=sci_arttext &pid=S1516-8913200500007&lng=en&nrm=iso&tlng=en [Internet].
 [http://dx.doi.org/10.1590/S1516-89132005000007]
- Silva F, Domingues FC. Antimicrobial activity of coriander oil and its effectiveness as food preservative. Crit Rev Food Sci Nutr 2017; 57(1): 35-47.https://www.tandfonline.com /doi/abs/10.1080/10408398.2013.847818 [Internet].
 [http://dx.doi.org/10.1080/10408398.2013.847818] [PMID: 25831119]
- Tajkarimi MM, Ibrahim SA, Cliver DO. Antimicrobial herb and spice compounds in food. Food Control 2010; 21(9): 1199-218.
 [http://dx.doi.org/10.1016/j.foodcont.2010.02.003]
- [13] Nabavi SM, Marchese A, Izadi M, Curti V, Daglia M, Nabavi SF. Plants belonging to the genus Thymus as antibacterial agents: from farm to pharmacy. Food Chem 2015; 173: 339-47. [http://dx.doi.org/10.1016/j.foodchem.2014.10.042] [PMID: 25466031]
- [14] Marchese A, Barbieri R, Sanches-Silva A, et al. Antifungal and antibacterial activities of allicin: A review. Trends Food Sci Technol 2016; 52: 49-56. [http://dx.doi.org/10.1016/j.tifs.2016.03.010]

- [15] Paphitou NI. Antimicrobial resistance: action to combat the rising microbial challenges. Int J Antimicrob Agents 2013; 42 (Suppl.): S25-8. [http://dx.doi.org/10.1016/j.ijantimicag.2013.04.007] [PMID: 23684003]
- [16] Högberg LD, Heddini A, Cars O. The global need for effective antibiotics: challenges and recent advances. Trends Pharmacol Sci 2010; 31(11): 509-15.
 [http://dx.doi.org/10.1016/j.tips.2010.08.002] [PMID: 20843562]
- [17] Lai PK, Roy J. Antimicrobial and chemopreventive properties of herbs and spices. Curr Med Chem 2004; 11(11): 1451-60.
 [http://dx.doi.org/10.2174/0929867043365107] [PMID: 15180577]
- [18] Nabavi SF, Di Lorenzo A, Izadi M, Sobarzo-Sánchez E, Daglia M, Nabavi SM. Antibacterial Effects of Cinnamon: From Farm to Food, Cosmetic and Pharmaceutical Industries. Nutrients 2015; 7(9): 7729-48. http://www.mdpi.com/2072-6643/7/9/5359 [Internet]. [http://dx.doi.org/10.3390/nu7095359] [PMID: 26378575]
- [19] Zheng J, Zhou Y, Li Y, Xu D-P, Li S, Li H-B. Spices for Prevention and Treatment of Cancers. Nutrients 2016; 8(8): 495.http://www.mdpi.com/2072-6643/8/8/495 [Internet]. [http://dx.doi.org/10.3390/nu8080495] [PMID: 27529277]
- [20] De M, Krishna De A, Banerjee AB. Antimicrobial screening of some indian spices 1999.https://onlinelibrary.wiley.com/doi/10.1002/(SICI)1099-1573(199911)13:7%3C616:AID-PTR475%3E3.0.CO;2-V [http://dx.doi.org/10.1002/(SICI)1099-1573(199911)13:7<616::AID-PTR475>3.0.CO;2-V]
- [21] Arora DS, Kaur J. Antimicrobial activity of spices. Int J Antimicrob Agents 1999; 12(3): 257-62.
 [http://dx.doi.org/10.1016/S0924-8579(99)00074-6] [PMID: 10461845]
- [22] Walsh C. Molecular mechanisms that confer antibacterial drug resistance. Nature 2000; 406(6797): 775-81.https://www.nature.com/articles/35021219 [Internet]. [http://dx.doi.org/10.1038/35021219] [PMID: 10963607]
- [23] Lister PD, Wolter DJ, Hanson ND. Antibacterial-resistant Pseudomonas aeruginosa: clinical impact and complex regulation of chromosomally encoded resistance mechanisms. Clin Microbiol Rev 2009; 22(4): 582-610. [http://dx.doi.org/10.1128/CMR.00040-09] [PMID: 19822890]
- [24] Sharma R, Sharma CL, Kapoor B. Antibacterial resistance: current problems and possible solutions. Indian J Med Sci 2005; 59(3): 120-9. [http://dx.doi.org/10.4103/0019-5359.15091] [PMID: 15805685]
- [25] Alekshun MN, Levy SB. Molecular mechanisms of antibacterial multidrug resistance. Cell 2007; 128 (6): 1037-50.
 [http://dx.doi.org/10.1016/j.cell.2007.03.004] [PMID: 17382878]
- [26] Xu JG, Liu T, Hu QP, Cao XM. Chemical composition, antibacterial properties and mechanism of action of essential oil from clove buds against staphylococcus aureus. Molecules 2016; 21(9): 1-13. [http://dx.doi.org/10.3390/molecules21091194] [PMID: 27617990]
- [27] Burt S. Essential oils: their antibacterial properties and potential applications in foods--a review. Int J Food Microbiol 2004; 94(3): 223-53. [http://dx.doi.org/10.1016/j.ijfoodmicro.2004.03.022] [PMID: 15246235]
- [28] Tiwari BK, Valdramidis VP, O'Donnell CP, Muthukumarappan K, Bourke P, Cullen PJ. Application of natural antimicrobials for food preservation. J Agric Food Chem 2009; 57(14): 5987-6000.https://pubs.acs.org/doi/abs/10.1021/jf900668n [Internet]. [http://dx.doi.org/10.1021/jf900668n] [PMID: 19548681]
- [29] Shreaz S, Wani WA, Behbehani JM, et al. Cinnamaldehyde and its derivatives, a novel class of antifungal agents. Fitoterapia 2016; 112: 116-31. [http://dx.doi.org/10.1016/j.fitote.2016.05.016] [PMID: 27259370]

Bacterial & Fungal Resistance

- [30] Li W, Shi Q, Dai H, Liang Q, Xie X. Antifungal activity, kinetics and molecular mechanism of action of garlic oil against Candida albicans. nature.com [Internet]. [cited 2021 Mar 16]; https://www.nature.com/articles/srep22805.
- [31] Freires IdeA, Murata RM, Furletti VF, et al. Coriandrum sativum L. (Coriander) essential oil: antifungal activity and mode of action on Candida spp., and molecular targets affected in human whole-genome expression. PLoS One 2014; 9(6)e99086 [http://dx.doi.org/10.1371/journal.pone.0099086] [PMID: 24901768]
- [32] Zhang J, Ye KP, Zhang X, Pan DD, Sun YY, Cao JX. Antibacterial activity and mechanism of action of black pepper essential oil on meat-borne escherichia coli. Front Microbiol 2017; 7(JAN): 2094. [http://dx.doi.org/10.3389/fmicb.2016.02094] [PMID: 28101081]
- [33] Albertson GD, Niimi M, Cannon RD, Jenkinson HF. Multiple efflux mechanisms are involved in Candida albicans fluconazole resistance. Antimicrob Agents Chemother 1996; 40(12): 2835-41. [http://dx.doi.org/10.1128/AAC.40.12.2835] [PMID: 9124851]
- [34] Sanglard D, Kuchler K, Ischer F, Pagani JL, Monod M, Bille J. Mechanisms of resistance to azole antifungal agents in Candida albicans isolates from AIDS patients involve specific multidrug transporters. Antimicrob Agents Chemother 1995; 39(11): 2378-86.http://aac.asm.org/ [Internet]. [http://dx.doi.org/10.1128/AAC.39.11.2378] [PMID: 8585712]
- [35] Sanglard D, Ischer F, Monod M, Bille J. Cloning of Candida albicans genes conferring resistance to azole antifungal agents: characterization of CDR2, a new multidrug ABC transporter gene. Microbiology 1997; 143(Pt 2): 405-16.https://www.microbiologyresearch.org/content/journal/micro/10.1099/00221287-143-2-405 [Internet].
 [http://dx.doi.org/10.1099/00221287-143-2-405] [PMID: 9043118]
- [36] White TC. Increased mRNA levels of ERG16, CDR, and MDR1 correlate with increases in azole resistance in Candida albicans isolates from a patient infected with human immunodeficiency virus. Antimicrob Agents Chemother 1997; 41(7): 1482-7.http://aac.asm.org/ [Internet]. [http://dx.doi.org/10.1128/AAC.41.7.1482] [PMID: 9210670]
- [37] SANGLARD D. Current understanding of the modes of action of and resistance mechanisms to conventional and emerging antifungal agents for treatment of Candida infections. Candida and Candidiasis [Internet]. 2002 [cited 2021 Mar 16]; https://ci.nii.ac.jp/naid/10018350289.
- [38] Löffler J, Kelly SL, Hebart H, Schumacher U, Lass-Flörl C, Einsele H. Molecular analysis of cyp51 from fluconazole-resistant Candida albicans strains. FEMS Microbiol Lett 1997; 151(2): 263-8.https://academic.oup.com/femsle/article-lookup/doi/10.1111/j.1574-6968.1997.tb12580.x [Internet].
 [http://dx.doi.org/10.1016/S0378-1097(97)00172-9] [PMID: 9228762]
- [39] Orozco AS, Higginbotham LM, Hitchcock CA, et al. Mechanism of fluconazole resistance in Candida krusei. Antimicrob Agents Chemother 1998; 42(10): 2645-9.http://aac.asm.org/ [Internet]. [http://dx.doi.org/10.1128/AAC.42.10.2645] [PMID: 9756770]
- [40] Sanglard D, Ischer F, Koymans L, Bille J. Amino acid substitutions in the cytochrome P-450 lanosterol 14α-demethylase (CYP51A1) from azole-resistant Candida albicans clinical isolates contribute to resistance to azole antifungal agents. Antimicrob Agents Chemother 1998; 42(2): 241-53.http://aac.asm.org/ [Internet].
 [http://dx.doi.org/10.1128/AAC.42.2.241] [PMID: 9527767]
- [41] Lopez-Ribot JL, McAtee RK, Lee LN, et al. Distinct patterns of gene expression associated with development of fluconazole resistance in serial candida albicans isolates from human immunodeficiency virus-infected patients with oropharyngeal candidiasis. Antimicrob Agents Chemother 1998; 42(11): 2932-7.http://aac.asm.org/ [Internet]. [http://dx.doi.org/10.1128/AAC.42.11.2932] [PMID: 9797228]
- [42] Sokol-Anderson ML, Brajtburg J, Medoff G. Amphotericin B-induced oxidative damage and killing of

Candida albicans. J Infect Dis 1986; 154(1): 76-83.https://academic.oup.com/jid/articlelookup/doi/10.1093/infdis/154.1.76 [Internet]. [http://dx.doi.org/10.1093/infdis/154.1.76] [PMID: 3519792]

- [43] Hernandez S, López-Ribot JL, Najvar LK, McCarthy DI, Bocanegra R, Graybill JR. Caspofungin resistance in Candida albicans: correlating clinical outcome with laboratory susceptibility testing of three isogenic isolates serially obtained from a patient with progressive Candida esophagitis. Antimicrob Agents Chemother 2004; 48(4): 1382-3.http://aac.asm.org/ [Internet]. [http://dx.doi.org/10.1128/AAC.48.4.1382-1383.2004] [PMID: 15047549]
- [44] Lima IO, De Oliveira Pereira F, De Oliveira WA, De Oliveira Lima E, Menezes EA, Cunha FA, et al. Antifungal activity and mode of action of carvacrol against Candida albicans strains. J Essent Oil Res 2013; 25(2): 138-42.https://www.tandfonline.com/doi/abs/10.1080/10412905.2012.754728 [Internet]. [http://dx.doi.org/10.1080/10412905.2012.754728]
- [45] Rao PV, Gan SH. Cinnamon: A multifaceted medicinal plant. Evidence-based Complement Altern Med 2014.
- [46] Rangari V. Pharmacognosy And Phytochemistry Vinod Rangari (PDF) credit by Okkonen M Katja archived 21 November 2015. ID 94cd58bb53 eBook PDF File: Pharmacognosy And Phytochemistry PHARMACOGNOSY AND PHYTOCHEMISTRY BY VINOD RANGARI. 2009.
- [47] Nowotarska SW, Nowotarski K, Grant IR, Elliott CT, Friedman M, Situ C. Mechanisms of Antimicrobial Action of Cinnamon and Oregano Oils, Cinnamaldehyde, Carvacrol, 2,5-Dihydroxybenzaldehyde, and 2-Hydroxy-5-Methoxybenzaldehyde against Mycobacterium avium subsp. paratuberculosis (Map). Foods 2017; 6(9): 72.http://www.mdpi.com/2304-8158/6/9/72 [Interne t].

[http://dx.doi.org/10.3390/foods6090072] [PMID: 28837070]

- [48] Zhang Y, Liu X, Wang Y, Jiang P, Quek SY. Antibacterial activity and mechanism of cinnamon essential oil against Escherichia coli and Staphylococcus aureus. Food Control 2016; 59: 282-9. [http://dx.doi.org/10.1016/j.foodcont.2015.05.032]
- [49] Kokate CK, Purohit AP, Gokhale SB. 2008.
- [50] Ranasinghe P, Journal PG-CM. 2016.https://www.researchgate.net/profile/Priyanga_ Ranasinghe/publication/299357322_Health_benefits_of_Ceylon_cinnamon_Cinnamomum_zeylanicu m_a_summary_of_the_current_evidence/links/570e1f2508aec783ddd063a1.pdf
- [51] Bandara T, Uluwaduge I, Jansz ER. Bioactivity of cinnamon with special emphasis on diabetes mellitus: a review. Int J Food Sci Nutr 2012; 63(3): 380-6.https://www.tandfonline. com/doi/abs/10.3109/09637486.2011.627849 [Internet].
 [http://dx.doi.org/10.3109/09637486.2011.627849] [PMID: 22007625]
- [52] Dorri M, Hashemitabar S, Hosseinzadeh H. Cinnamon (Cinnamomum zeylanicum) as an antidote or a protective agent against natural or chemical toxicities: a review. Drug Chem Toxicol 2018; 41(3): 338-51.https://www.tandfonline.com/doi/abs/10.1080/01480545.2017.1417995 [Internet]. [http://dx.doi.org/10.1080/01480545.2017.1417995] [PMID: 29319361]
- [53] Saeed S, Tariq P. *in vitro* ANTIBACTERIAL ACTIVITY OF CLOVE AGAINST GRAM NEGATIVE BACTERIA. Pak J Bot 2008; 40(5): 2157-60.
- [54] Pinto E, Vale-Silva L, Cavaleiro C, Salgueiro L. Antifungal activity of the clove essential oil from Syzygium aromaticum on Candida, Aspergillus and dermatophyte species. J Med Microbiol 2009; 58 (Pt 11): 1454-62.https://www.microbiologyresearch. org/content/journal/jmm/ 10.1099/jmm.0.010538-0 [Internet]. [http://dx.doi.org/10.1099/jmm.0.010538-0] [PMID: 19589904]
- [55] Ahmed SS. Alwaan AL-jubori, Abd-Alkhaliq. GARLIC (ALLIUM SATIVUM) AND CLOVE (SYZYGIUM AROMATICUM) AS ALTERNATIVE TREATMENTS FOR THE CONTROL OF HAEMONCHUS CONTORTUS IN SHEEP. Plant Arch 2020; 20(1): 106-9.http://www. plantarchives.org /SPECIAL ISSUE 20-1/19_106-109_.pdf [Internet].

- [56] Echeverría I, López-Caballero ME, Gómez-Guillén MC, Mauri AN, Montero MP. Active nanocomposite films based on soy proteins-montmorillonite- clove essential oil for the preservation of refrigerated bluefin tuna (Thunnus thynnus) fillets. Int J Food Microbiol 2018; 266: 142-9. [http://dx.doi.org/10.1016/j.jifoodmicro.2017.10.003] [PMID: 29216554]
- [57] Mohapatra S, Leelavathi L, Rajeshkumar S, Sakthi DS, Jayashri P. Assessment of Cytotoxicity, Anti-Inflammatory and Antioxidant Activity of Zinc Oxide Nanoparticles Synthesized Using Clove and Cinnamon Formulation--An In-Vitro Study. J Evol Med Dent Sci 2020; 9(25): 1859-65. https://go.gale.com/ps/i.do?p=AONE&sw=w&issn=22784748&v=2.1&it=r&id=GALE%7CA630993 246&sid=googleScholar&linkaccess=fulltext [Internet]. [http://dx.doi.org/10.14260/jemds/2020/405]
- [58] Cui H, Zhang C, Li C, Lin L. Antibacterial mechanism of oregano essential oil. Ind Crops Prod 2019; 139111498

[http://dx.doi.org/10.1016/j.indcrop.2019.111498]

- [59] Soylu S, Yigitbas H, Soylu EM, Kurt S. Antifungal effects of essential oils from oregano and fennel on Sclerotinia sclerotiorum. J Appl Microbiol 2007; 103(4): 1021-30.http://doi.wiley.com/10.1111/j.1365-2672.2007.03310.x [Internet]. [http://dx.doi.org/10.1111/j.1365-2672.2007.03310.x] [PMID: 17897206]
- [60] Veenstra JP, Johnson JJ. Oregano (Origanium Vulgare) Extract for Food Preservation and Improving Gastrointestinal Health. Int J Nutr [Internet]. 2019 Apr 9 [cited 2021 Mar 16];3(4):43–52. Available from: /pmc/articles/PMC6508890/.
- [61] Akinbo DB, Onyeaghala AA, Emomidue JO, Ogbhemhe SO, Okpoli HC. Phytochemical and antiinflammatory activities of aqueous leaf extract of Indian borage (oregano) on rats induced with inflammation. Cancer Biomark 2018; 22(2): 257-65. [http://dx.doi.org/10.3233/CBM-170893] [PMID: 29630520]
- [62] Bajagai YS, Steel JC, Radovanovic A, Stanley D. Prolonged continual consumption of oregano herb interferes with the action of steroid hormones and several drugs, and effects signaling across the braingut axis. Food Funct 2021; 12(2): 726-38.https://pubs.rsc.org/en/content/articlehtml/2021/fo/d0fo02988b [Internet].
 [http://dx.doi.org/10.1039/D0FO02988B] [PMID: 33349823]
- [63] Esmail Al-Snafi A, Author C. The chemical constituents and pharmacological effects of Foeniculum vulgare-A review Medicinal plant with reproductive and endocrine effects View project Medicinal plant with gastrointestinal effects View project The chemical constituents and pharmacological effects of Foeniculum vulgare-A review www.iosrphr.org2018.https://www.researchgate .net/publication/325809622
- [64] Niranjan A, Prakash D. 2008.https://www.researchgate.net/publication/283863862
- [65] Teow SY, Liew K, Ali SA, Khoo ASB, Peh SC. Antibacterial Action of Curcumin against Staphylococcus aureus: A Brief Review. J Trop Med. 2016;2016.
- [66] Lee W, Lee DG. An antifungal mechanism of curcumin lies in membrane-targeted action within *Candida albicans*. IUBMB Life 2014; 66(11): 780-5.http://doi.wiley.com/10.1002/iub.1326 [Internet]. [http://dx.doi.org/10.1002/iub.1326] [PMID: 25380239]
- [67] El-Saber Batiha G, Magdy Beshbishy A. 2020.https://www.mdpi.com/2072-6643/12/3/872
- [68] Shang A, Cao S-Y, Xu X-Y, et al. Bioactive Compounds and Biological Functions of Garlic (Allium sativum L.). Foods 2019; 8(7): 246.https://www.mdpi.com/2304-8158/8/7/246 [Internet]. [http://dx.doi.org/10.3390/foods8070246] [PMID: 31284512]
- [69] Yang C, Li L, Yang L, Lů H, Wang S, Sun G. Anti-obesity and Hypolipidemic effects of garlic oil and onion oil in rats fed a high-fat diet. Nutr Metab (Lond) 2018; 15(1): 43.https://nutritionandmetabolism.biomedcentral.com/articles/10.1186/s12986-018-0275-x [Internet].

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[http://dx.doi.org/10.1186/s12986-018-0275-x] [PMID: 29951108]

- [70] Baek SC, Nam KH, Yi SA, *et al.* Anti-adipogenic Effect of β-Carboline Alkaloids from Garlic (*Allium sativum*). Foods 2019; 8(12): 673.https://www.mdpi.com/2304-8158/8/12/673 [Internet]. [http://dx.doi.org/10.3390/foods8120673] [PMID: 31842405]
- [71] Sripanidkulchai B. Benefits of aged garlic extract on Alzheimer's disease: Possible mechanisms of action. Exp Ther Med 2020; 19(2): 1560-4. [Review].
 [PMID: 32010339]
- [72] Kosuge Y, Koen Y, Ishige K, *et al.* S-allyl-L-cysteine selectively protects cultured rat hippocampal neurons from amyloid β-protein- and tunicamycin-induced neuronal death. Neuroscience 2003; 122(4): 885-95.
 [http://dy.doi.org/10.1016/j.pouroscience.2002.08.0261 [DMID: 14642758]

[http://dx.doi.org/10.1016/j.neuroscience.2003.08.026] [PMID: 14643758]

- [73] Imai T, Kosuge Y, Ishige K, Ito Y. Amyloid β-protein potentiates tunicamycin-induced neuronal death in organotypic hippocampal slice cultures. Neuroscience 2007; 147(3): 639-51. [http://dx.doi.org/10.1016/j.neuroscience.2007.04.057] [PMID: 17560726]
- [74] Führer M, Dejaco C, Kopp B, Hammer J. Gastric administration of garlic powder containing the trpalagonist allicin induces specific epigastric symptoms and gastric relaxation in healthy subjects. Neurogastroenterol Motil 2019; 31(1)e13470https://onlinelibrary.wiley.com/doi/abs/10.1111/nmo.13470 [Internet]. [http://dx.doi.org/10.1111/nmo.13470] [PMID: 30238636]
- [75] Macpherson LJ, Geierstanger BH, Viswanath V, et al. The pungency of garlic: activation of TRPA1 and TRPV1 in response to allicin. Curr Biol 2005; 15(10): 929-34. [http://dx.doi.org/10.1016/j.cub.2005.04.018] [PMID: 15916949]
- [76] Mao QQ, Xu XY, Cao SY, Gan RY, Corke H, Beta T, et al. 2019.
- [77] Abral H, Ariksa J, Mahardika M, Handayani D, Aminah I, Sandrawati N, et al. Transparent and antimicrobial cellulose film from ginger nanofiber. Food Hydrocoll 2020; 98105266 [http://dx.doi.org/10.1016/j.foodhyd.2019.105266]
- [78] Abral H, Ariksa J, Mahardika M, Handayani D, Aminah I, Sandrawati N, et al. Highly transparent and antimicrobial PVA based bionanocomposites reinforced by ginger nanofiber. Polym Test 2020; 81106186 [http://dx.doi.org/10.1016/j.polymertesting.2019.106186]
- [79] Shahrajabian MH, Sun W, Cheng Q. Clinical aspects and health benefits of ginger (Zingiber officinale) in both traditional Chinese medicine and modern industry. Acta Agric Scand Sect B Soil Plant Sci [Internet] 2019; 69(6): 546-56.https://www.tandfonline.com/doi/abs/10.1080/09064710.2019.1606930 [http://dx.doi.org/10.1080/09064710.2019.1606930]
- [80] Ma RH, Ni ZJ, Zhu YY, et al. A recent update on the multifaceted health benefits associated with ginger and its bioactive components. Food Funct 2021; 12(2): 519-42.https://pubs.rsc.org/en/content/articlehtml/2021/fo/d0fo02834g [Internet]. [http://dx.doi.org/10.1039/D0FO02834G] [PMID: 33367423]
- [81] Mahomoodally MF, Aumeeruddy MZ, Rengasamy KRR, et al. Ginger and its active compounds in cancer therapy: From folk uses to nano-therapeutic applications. Semin Cancer Biol 2021; 69: 140-9. [http://dx.doi.org/10.1016/j.semcancer.2019.08.009] [PMID: 31412298]
- [82] Wang J, Wang P, Li D, Hu X, Chen F. Beneficial effects of ginger on prevention of obesity through modulation of gut microbiota in mice. Eur J Nutr 2020; 59(2): 699-718.https://link.springer.com/article/10.1007/s00394-019-01938-1 [Internet]. [http://dx.doi.org/10.1007/s00394-019-01938-1] [PMID: 30859364]
- [83] Ganesh P, Suresh Kumar R, Saranraj P. 2014.http://scholarsresearchlibrary.com/archive.html
- [84] Chen W, Zou L, Chen W, Hu Y, Chen H. Effects of Black Pepper (Piper nigrum L.) Chloroform Extract on the Enzymatic Activity and Metabolism of Escherichia coli and Staphylococcus aureus. J

Food Qual. 2018;2018.

- [85] Amalraj A, Haponiuk JT, Thomas S, Gopi S. Preparation, characterization and antimicrobial activity of polyvinyl alcohol/gum arabic/chitosan composite films incorporated with black pepper essential oil and ginger essential oil. Int J Biol Macromol 2020; 151: 366-75. [http://dx.doi.org/10.1016/j.ijbiomac.2020.02.176] [PMID: 32084477]
- Dheeb BI. Antifungal Activity of Alkaloids and Phenols Compounds extracted from black pepper [86] Piper nigrum against some pathogenic fungi. Magallat Markaz Buhut al-Tiqniyyat al-Ahya'iyyat 2015; 9(2): 46-54.
- [87] Meghwal M, Goswami TK. Piper nigrum and piperine: an update. Phytother Res 2013; 27(8): 1121--. [http://dx.doi.org/10.1002/ptr.4972] [PMID: 23625885]
- [88] Heidari Beni M, Moravejolahkami AR, Gorgian P, Askari G, Tarrahi MJ, Bahreini Esfahani N. 2020.https://onlinelibrary.wiley.com/doi/abs/10.1002/ptr.6671
- [89] Wang D, Zhang L, Huang J, Himabindu K, Tewari D, Horbańczuk JO, et al. Cardiovascular protective effect of black pepper (Piper nigrum L.) and its major bioactive constituent piperine. Trends Food Sci Technol 2020.
- [90] Önder A. Coriander and Its Phytoconstituents for the Beneficial Effects.Potential of Essential Oils. InTech 2018. [http://dx.doi.org/10.5772/intechopen.78656]

- Krishnaveni MFT-IR. GC-MS/MS analysis of essential oil from coriandrum sativum [91] seeds, antibacterial assay. Characterization of reproductive gene in marine fish View project. 2017. http://www.soeagra.com/abr.html
- [92] Porte A, Godoy RLO. Chemical composition of Thymus vulgaris L. (thyme) essential oil from the Rio de Janeiro State (Brazil). J Serb Chem Soc 2008; 73(3): 307-10. [http://dx.doi.org/10.2298/JSC0803307P]
- Boskovic M, Zdravkovic N, Ivanovic J, Janjic J, Djordjevic J, Starcevic M, et al. Antimicrobial [93] Activity of Thyme (Tymus vulgaris) and Oregano (Origanum vulgare) Essential Oils against Some Food-borne Microorganisms. Procedia Food Sci 2015; 5: 18-21. [http://dx.doi.org/10.1016/j.profoo.2015.09.005]
- [94] de Lira Mota KS, de Oliveira Pereira F, de Oliveira WA, Lima IO, de Oliveira Lima E. Antifungal activity of Thymus vulgaris L. essential oil and its constituent phytochemicals against Rhizopus oryzae: with interaction ergosterol. Molecules 2012: 17(12): 14418-33.http://www.mdpi.com/1420-3049/17/12/14418 [Internet]. [http://dx.doi.org/10.3390/molecules171214418] [PMID: 23519243]
- [95] Kulkarni SK, Bhutani MK, Bishnoi M. Antidepressant activity of curcumin: involvement of serotonin Psychopharmacology and dopamine system. 2008; 201(3): (Berl) 435-42.https://link.springer.com/article/10.1007/s00213-008-1300-v [Internet]. [http://dx.doi.org/10.1007/s00213-008-1300-y] [PMID: 18766332]
- Elgayyar M, Draughon FA, Golden DA, Mount JR. Antimicrobial activity of essential oils from plants [96] against selected pathogenic and saprophytic microorganisms. J Food Prot 2001; 64(7): 1019-24.http://meridian.allenpress.com/jfp/article-pdf/64/7/1019/1674454/0362-028x-64 7 1019.pdf [Internet].

[http://dx.doi.org/10.4315/0362-028X-64.7.1019] [PMID: 11456186]

- [97] Al-snafi AE. The pharmacological activities of Cuminum cyminum -A review The pharmacological activities of Cuminum cyminum - A review Prof Dr Ali Esmail Al-Snafi. IOSR J Pharm 2017; 6(2): 46-65.
- DP. [98] IL, Nurjanah S, Putri Sugiarti Antibacterial Activity of Nutmeg Oil 2017.https://knepublishing.com/index.php/KnE-Life/article/view/1074 [http://dx.doi.org/10.18502/kls.v2i6.1076]

- [99] Ibrahim KM, Naem RK, Abd-Sahib AS. Antibacterial Activity of Nutmeg (Myristica fragrans) Seed Extracts Against Some Pathogenic Bacteria. J Al-Nahrain Univ Sci 2013; 16(2): 188-92. [http://dx.doi.org/10.22401/JNUS.16.2.29]
- [100] Nieto G, Ros G, Castillo J. Antioxidant and Antimicrobial Properties of Rosemary (*Rosmarinus officinalis*, L.): A Review. Medicines (Basel) 2018; 5(3): 98.http://www.mdpi.com/2305-6320/5/3/98 [Internet]. [http://dx.doi.org/10.3390/medicines5030098] [PMID: 30181448]
- [101] Clemente I, Aznar M, Silva F, Nerín C. Antimicrobial properties and mode of action of mustard and cinnamon essential oils and their combination against foodborne bacteria. Innov Food Sci Emerg Technol 2016; 36: 26-33. [http://dx.doi.org/10.1016/j.ifset.2016.05.013]
- [102] Delaquis P, Technol GM-F. 1995 undefined. Antimicrobial properties of isothiocyanates in food preservation. Food Technol 1995; 49(11): 73-84. https://www.researchgate.net/profile/Pascal_Delaquis/publication/279891141_Antimicrobial_propertie s_of_isothiocyanates_in_food_preservation/links/55f31ff108ae7a10cf88b23d/Antimicrobialproperties-of-isothiocyanates-in-food-preservation.pdf [Internet].
- [103] Adegbeye MJ, Elghandour MMMY, Faniyi TO, Rivero Perez N, Barbabosa-Pilego A, Zaragoza-Bastida A, et al. Antimicrobial and antihelminthic impacts of black cumin, pawpaw and mustard seeds in livestock production and health. Agrofor Syst 2020; 94(4): 1255-68.https://link.springer.com/article/10.1007/s10457-018-0337-0 [Internet]. [http://dx.doi.org/10.1007/s10457-018-0337-0]
- [104] Bahmid NA, Pepping L, Dekker M, Fogliano V, Heising J. Using particle size and fat content to control the release of Allyl isothiocyanate from ground mustard seeds for its application in antimicrobial packaging. Food Chem 2020; 308125573 [http://dx.doi.org/10.1016/j.foodchem.2019.125573] [PMID: 31639598]
- [105] Mejía-Garibay B, Palou E, López-Malo A. Composition, diffusion, and antifungal activity of black mustard (Brassica nigra) essential oil when applied by direct addition or vapor phase contact. J Food Prot 2015; 78(4): 843-8.http://meridian.allenpress.com/jfp/article-pdf/78/4/843/1688232/0362-028x_jfp-14-485.pdf [Internet].
 [Internet].
 - [http://dx.doi.org/10.4315/0362-028X.JFP-14-485] [PMID: 25836415]
- [106] ENAİBE BU, ADİGUN OO, ADİGUN FM, GBADAMOSİ İT. Dose-dependent effect of black mustard seeds (Brassica nigra) extract on the prefrontal cortex of adult Wistar rats. Anatomy 2017; 11 (3): 107-14.www.anatomy.org.tr [Internet].
 [http://dx.doi.org/10.2399/ana.17.027]
- [107] Jo S-H, Cho C-Y, Ha K-S, Lee J-Y, Choi H-Y, Kwon Y-I, et al. in vitro and in vivo antihyperglycemic effects of green and red mustard leaves (*Brassica juncea* var. integrifolia). J Food Biochem 2018; 42(5)e12583http://doi.wiley.com/10.1111/jfbc.12583 [Internet]. [http://dx.doi.org/10.1111/jfbc.12583]
- [108] Zareen A, Gardezi DA, Naeemullah M, Masood MS, Tahira R. Journal of Medicinal Plants Studies Screening of Antibacterial Potential of Siam Queen, Holy Basil and Italian Basil Essential Oils. J Med Plants Stud Screen Antibact Potential Siam Queen. Holy Basil Ital Basil Essent Oils 2014; 2(2): 63-8.
- [109] Oxenham SK, Svoboda KP, Walters DR. Antifungal Activity of the Essential Oil of Basil (Ocimum basilicum). J Phytopathol 2005; 153(3): 174-80.http://doi.wiley.com/10.1111/j.1439-0434.2005.00952.x [Internet]. [http://dx.doi.org/10.1111/j.1439-0434.2005.00952.x]
- [110] Patil SD, Shinde S, Kandpile P, Jain Shri DDAS. EVALUATION OF ANTIMICROBIAL ACTIVITY OF ASAFOETIDA. Int J Pharm Sci Res 2015; 6(2): 722-7. [Internet].

[http://dx.doi.org/10.13040/IJPSR.0975-8232.6]

- [111] Angelini P, Pagiotti R, Venanzoni R. 2009.https://www.researchgate.net/profile/Paola_ Angelini2/publication/229832851_Antifungal_and_allelopathic_effects_of_Asafoetida_against_Trich oderma_harzianum_and_Pleurotus_spp/links/0912f50114dbb803bc000000/Antifungal-an--allelopathic-effects-of-Asafoetida-against-Trichoderma-harzianum-and-Pleurotus-spp.pdf
- [112] Menon AN. Chemical composition of the volatile oils of *Alpinia galanga* plant parts from Kerala. J Essent Oil-Bearing Plants [Internet] 2006; 9(3): 277-82.https://www.tandfonline.com/doi/abs/10.1080/0972060X.2006.10643504 [http://dx.doi.org/10.1080/0972060X.2006.10643504]
- [113] Oonmetta-aree J, Suzuki T, Gasaluck P, Eumkeb G. Antimicrobial properties and action of galangal (*Alpinia galanga* Linn.) on Staphylococcus aureus. Lebensm Wiss Technol 2006; 39(10): 1214-20. [http://dx.doi.org/10.1016/j.lwt.2005.06.015]
- [114] Haraguchi H, Kuwata Y, Inada K, *et al.* Antifungal activity from *Alpinia galanga* and the competition for incorporation of unsaturated fatty acids in cell growth. Planta Med 1996; 62(4): 308-13.http://www.thieme-connect.de/DOI/DOI?10.1055/s-2006-957890 [Internet]. [http://dx.doi.org/10.1055/s-2006-957890] [PMID: 8792660]
- [115] Souissi M, Azelmat J, Chaieb K, Grenier D. Antibacterial and anti-inflammatory activities of cardamom (Elettaria cardamomum) extracts: Potential therapeutic benefits for periodontal infections. Anaerobe 2020; 61102089 [http://dx.doi.org/10.1016/j.anaerobe.2019.102089] [PMID: 31430531]
- [116] Noumi E, Snoussi M, Alreshidi MM, et al. Chemical and Biological Evaluation of Essential Oils from Cardamom Species. Molecules 2018; 23(11): 2818.http://www.mdpi.com/1420-3049/23/11/2818
 [Internet].
 [http://dx.doi.org/10.3390/molecules23112818] [PMID: 30380739]
- [117] Chahal K, Kaur R, Kumar A, Bhardwaj U. Chemistry and biological activities of Anethum graveolens L. (dill) essential oil: A review. J Pharmacogn Phytochem 2017; 6(2): 295-306.https://www.phytojournal.com/archives/2017/vol6issue2/PartF/6-2-67-817.pdf [Internet].
- [118] Mutlu-Ingok A, Karbancioglu-Guler F. Cardamom, Cumin, and Dill Weed Essential Oils: Chemical Compositions, Antimicrobial Activities, and Mechanisms of Action against Campylobacter spp. Molecules 2017; 22(7)E1191 [http://dx.doi.org/10.3390/molecules22071191] [PMID: 28714890]
- [119] Tian J, Ban X, Zeng H, He J, Chen Y, Wang Y. 2012.https://dx.plos.org/10.1371/journal.pone.0030147
- [120] Amer A, Aly U. 2019.http://www.epj.eg.net/text.asp?2019/18/1/68/254969
- [121] Huang Y, Zhao J, Zhou L, et al. Antifungal activity of the essential oil of Illicium verum fruit and its main component trans-anethole. Molecules 2010; 15(11): 7558-69. [http://dx.doi.org/10.3390/molecules15117558] [PMID: 21030909]
- [122] Liu Q, Meng X, Li Y, Zhao C-N, Tang G-Y, Li H-B. Antibacterial and Antifungal Activities of Spices. Int J Mol Sci 2017; 18(6): 1283.http://www.mdpi.com/1422-0067/18/6/1283 [Internet]. [http://dx.doi.org/10.3390/ijms18061283] [PMID: 28621716]
- [123] Aumeeruddy-Elafi Z, Gurib-Fakim A, Mahomoodally M. Antimicrobial and antibiotic potentiating activity of essential oils from tropical medicinal herbs and spices 2016. http://www.investigacionyposgrado.uadec.mx/site/wp-content/uploads/2020/07/4-Chapter-2016.pdf#p age=294
- [124] Batiha GES, Alkazmi LM, Wasef LG, Beshbishy AM, Nadwa EH, Rashwan EK. Syzygium aromaticum I (myrtaceae): Traditional uses, bioactive chemical constituents, pharmacological and toxicological activities. Biomolecules. MDPI AG 2020; Vol. 10: p. 202.

- [125] Moghrovyan A, Sahakyan N, Babayan A, Chichoyan N, Petrosyan M, Trchounian A. Essential Oil and Ethanol Extract of Oregano (Origanum vulgare L.) from Armenian Flora as a Natural Source of Terpenes, Flavonoids and other Phytochemicals with Antiradical, Antioxidant, Metal Chelating, Tyrosinase Inhibitory and Antibacterial Activity. Curr Pharm Des 2019; 25(16): 1809-16. [http://dx.doi.org/10.2174/1381612825666190702095612] [PMID: 31267860]
- [126] Badgujar SB, Patel VV, Bandivdekar AH. Foeniculum vulgare Mill: A review of its botany, phytochemistry, pharmacology, contemporary application, and toxicology. 2014.
- [127] Prasad S, Aggarwal BB. Turmeric, the golden spice: From traditional medicine to modern medicine. In: Herbal Medicine: Biomolecular and Clinical Aspects: Second Edition. CRC Press 2011; pp. 263-88.
- [128] Mao Q-Q, Xu X-Y, Cao S-Y, et al. Bioactive Compounds and Bioactivities of Ginger (Zingiber officinale Roscoe). Foods 2019; 8(6): 185.https://www.mdpi.com/2304-8158/8/6/185 [Internet]. [http://dx.doi.org/10.3390/foods8060185] [PMID: 31151279]
- [129] Gorgani L, Mohammadi M, Najafpour GD, Nikzad M. Piperine-The bioactive compound of black pepper: from isolation to medicinal formulations. Compr Rev Food Sci Food Saf 2017; 16(1): 124-40. [http://dx.doi.org/10.1111/1541-4337.12246] [PMID: 33371546]
- [130] Önder A. Coriander and its phytoconstituents for the beneficial effects https://books.google.com/books?hl=en&lr=&id=Dm-QDwAAQBAJ&oi=fnd&pg=PA165&dq=%5B67%5D%092018.
- [131] Salehi B, Mishra AP, Shukla I, et al. Thymol, thyme, and other plant sources: Health and potential uses. Phytotherapy Research. John Wiley and Sons Ltd 2018; 32: pp. 1688-704.
- [132] Johri RK. Cuminum cyminum and Carum carvi: An update. Vol. 5, Pharmacognosy Reviews. Wolters Kluwer -- Medknow Publications; 2011. p. 63–72.
- [133] Abourashed EA, El-Alfy AT. Chemical diversity and pharmacological significance of the secondary metabolites of nutmeg (Myristica fragrans Houtt.). Vol. 15, Phytochemistry Reviews. Springer Netherlands; 2016. p. 1035–56.
- [134] De Oliveira JR, Camargo SEA, De Oliveira LD. *Rosmarinus officinalis* L. (rosemary) as therapeutic and prophylactic agent. Vol. 26, Journal of Biomedical Science. BioMed Central Ltd.; 2019. p. 1–22.
- [135] Rahman M, Khatun A, Liu L, Barkla BJ. Brassicaceae mustards: Traditional and agronomic uses in Australia and New Zealand. Molecules. MDPI AG 2018; Vol. 23.
- [136] Joshi RK. Chemical composition and antimicrobial activity of the essential oil of Ocimum basilicum L. (sweet basil) from Western Ghats of North West Karnataka, India. Anc Sci Life 2014; 33(3): 151-6. [http://dx.doi.org/10.4103/0257-7941.144618] [PMID: 25538349]
- [137] Iranshahy M, Iranshahi M. Traditional uses, phytochemistry and pharmacology of asafoetida (Ferula assa-foetida oleo-gum-resin)-a review. J Ethnopharmacol 2011; 134(1): 1-10. [http://dx.doi.org/10.1016/j.jep.2010.11.067] [PMID: 21130854]
- [138] Basri AM, Taha H, Ahmad N. A review on the pharmacological activities and phytochemicals of Alpinia officinarum (Galangal) extracts derived from bioassay-guided fractionation and isolation. Pharmacognosy Reviews. Medknow Publications 2017; 11: pp. 43-56.
- [139] Bajaj YPS, Reghunath BR, Gopalakrishnan PK. Elettaria cardamomum Maton (Cardamom): Aromatic Compounds, In Vitro Culture Studies, and Clonal Propagation. Berlin, Heidelberg: Springer 1993; pp. 132-47.
- [140] Jana S, Shekhawat G. Anethum graveolens: An Indian traditional medicinal herb and spice. Pharmacognosy Reviews. Wolters Kluwer: Medknow Publications 2010; 4: pp. 179-84.
- [141] Shojaii A, Abdollahi Fard M. Review of Pharmacological Properties and Chemical Constituents of *Pimpinella anisum*. ISRN Pharm 2012; 2012510795. [http://dx.doi.org/10.5402/2012/510795] [PMID: 22848853]

Bacterial & Fungal Resistance

[142] Ayush T. Ayurveda's immunity boosting measures for self care during COVID 19 crisis. [Internet]. Tan The Ministry of Ayurvedic, Unani, Siddha and Homeopathy 2020.

CHAPTER 5

Naturally Isolated Compounds from Spices and Herbs and their Medicinal Uses

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Abstract: Spices and herbs have a long history of medicinal uses. They include turmeric, basil, mace, cinnamon, ginger, etc. [1]. Ginseng and Ginkgo biloba are reportedly used to improve stamina and cognitive performance. Spices are used in all the countries for different purposes, such as in cooking and medicines, etc. Spice is a seed, fruit, root, bark, or other plant substance primarily used for coloring, flavoring, and preserving food. Herbs are the leaves, flowers, stems from plants used for flavoring or as garnishing. Medicinal and aromatic plants have also been used therapeutically to improve the health and wellbeing of animals; most were used for prophylactic purposes and to improve the growth rate and feed conversion ratio efficiency [2, 3]. The alternatives to antibiotics as growth stimulators from the group of prebiotics, probiotics, organic acids, essential oils, medicinal plants, or parts of plants, such as thyme, basil, oregano, pepper and plenty of others, are numerous [2]. This chapter includes a wide variety of isolated compounds, such as phenolic compounds and flavanoids present in spices, which are now experimentally documented to possess antioxidant, antiinflammatory, antimutagenic and anticarcinogenic activities. It also includes a list of spices compounds that are experimentally evidenced to control cardiovascular diseases, diabetes, cataract, cancer, etc.

Keywords: Herbs, In-vitro, Isolated compounds, Spices.

INTRODUCTION

Spices and herbs are an important part of the human diet to enhance the flavor, color and aroma of food. They have also been used from ancient times as traditional medicine to improve the health of animals. Spices and herbs can be classified on the basis of flavor, taxonomy or part of the plant from where they came. Presently there has been a trend to use natural substances present in fruits, vegetables, oil seeds, and herbs as antioxidants and rational foods [4 - 6]. According to the World Health Organization (WHO), essentially 20,000 medicinal plants

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reside in 91 countries. The premier steps to make use of the biologically active phytochemicals from plant resources are extraction, pharmacological screening, isolation and characterization of the bioactive compound, toxicological analysis and clinical evaluation [7]. A brief summary of the general approaches in extraction, isolation, and characterization of bioactive compounds from plant extract is presented in Fig. (1).

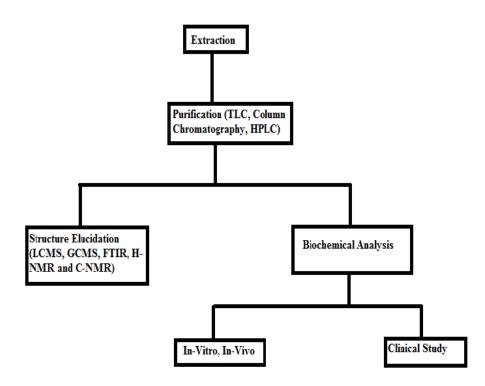


Fig. (1). A brief summary of the general approaches in extraction, isolation, and characterization of a bioactive compound from plant extracts [8].

Spices and herbs are found to be rich sources of phytochemicals [9 - 12]. Phytochemicals are a large group of bioactives derived from plants, which have potential protective effects against diseases. This group consists of flavonoids and other phenolic compounds, carotenoids, plant sterols, glucosinolates and other sulphur-containing compounds. There are more than 6000 known flavonoids (Jaganath & Crozier, 2010). Phenolic chemicals have a variety of activities in plants, including structural and defensive activities, and they serve as pollinators and seed-dispersing animal attractants.

Differences between Spices and Herbs

Spices come from different parts of a plant other than the leaves, while herbs come from the leaves of a plant. They can be classified into various groups based on taste, taxonomy or part of the plant where they came from.

Based on taste, spices and herbs can be classified into the following groups:

a) Hot spices (black and white peppers, Cayenne pepper, mustard, chilies).

b) Mild flavor spices (paprika, coriander), aromatic spices (clove, cumin, dill fennel, nutmeg, mace, cinnamon) and

c) Aromatic herbs and vegetables (thyme, basil, bay leaf, marjoram, shallot, onion, garlic)

Uses and Benefits of Spices

Table 1 enlists the uses and benefits of spices.

Spices	Uses	Benefits
Asafoetida (Hing)	It is used for seasoning food, especially snacks, and has medicinal uses	A good remedy for whooping cough and stomach ache.
Cardamom (Elaichi)	dishes, it used to give a good flavor and	It helps to control bad breath and digestive disorders. Also the whole cardamom chewed is good for coping with Diabetes.
Chilly (Lal Mirch)	It is the main ingredient used for adding hot flavor to the food.	The antioxidants present in chilly help cope up with cholesterol, and also help in burning calories.
Cinnamon (Dalchini)	It is used mainly for seasoning food and preparing masala. It has medicinal value too.	It supports the natural production of insulin and reduces blood cholesterol.
Clove (Laung)		Clove oil is beneficial for coping with toothache, sore gums, chest pains, fever, digestive problems, cough, and cold.
Coriander (Dhaniya)		It can be used externally on aching joints and rheumatism. It is also good for coping with soar throat, allergies, digestion problems, hay fever, <i>etc</i> .

Table 1. List of Important Spices with their Uses and Benefits [13 - 16].

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Spices	Uses	Benefits
Cumin (Zeera)	It is used for cooking and it also possesses medicinal properties.	It is a good source of iron and keeps the immune system healthy. Water boiled with cumin seeds is good for coping with dysentery.
Curry leaves(Curry Patta)	It is used as the main ingredient for seasoning in some countries. It has many medicinal uses.	These leaves are beneficial for reducing blood sugar. Each part of the plant provides some benefit or the other. The dried leaves are extensively used in herbal medicines.
Fenugreek (Methi)		Fenugreek seed tea or sweet fudge is good for increasing breast milk. It is also helpful for treating diabetes and lowering cholesterol.
Garlic (Lassan)	It is used for cooking as well as for medicinal purpose.	It is useful for coping with cough and cold. It also has antibiotic properties.
Ginger (Adrak)	It is used for giving a specific flavor to food and has many medicinal uses.	It helps avoid digestive problems. It is beneficial for coping with cough and cold.
Mustard (Rye)	green leafy vegetables. The use of	Mustard oil is good for body massage and even for getting good hair. It consists of omega-3 fatty acids. It is an excellent source of iron, zinc, manganese, calcium, protein, <i>etc</i> .
Nutmeg (Jaiphal)	garnishing and preparing the masala. It is	It is beneficial for the treatments of asthma, heart disorder, and bad breath problems.
Pepper (Kaali Mirch)	It is extensively used in cooking, especially for garnishing. It has many medicinal uses too.	It helps to deal with cold, cough, infections, muscle pains, and digestive problems.
Saffron (Zaffran/Kesar)	It is used for cooking as well as in beauty products. It is mainly used in sweet dishes. It has good medicinal properties.	It helps cope with skin diseases. It is a good remedy for cough, cold, and asthma.
Star anise (Chakra Phool)	It is used in cooking and for medicinal use.	Star anise oil is beneficial for rheumatism, digestion and avoiding bad breath.
Turmeric (Haldi)	It is used in cooking and skincare products. It has a wide range of medicinal use.	It helps to deal with skin problems, and also it makes coping with diabetes easier. Turmeric powder can be used for healing cuts and wounds.

Chemical Compounds in Herbs and Spices

Anise

Anise is a seed spice derived from a flowering plant belonging to the family *Apiaceae*. Flavor and aroma from anise come from anethole. Anethole is a phenylpropene derivative found in anise (*Pimpinella anisum*) and fennel (*Foeniculum vulgare*). Anethole occurs naturally in high concentrations in volatile oils, such as anise oil (80–90%), star anise oil (over 90%), and fennel oil (80%) [17]. Anethole exists in both a *cis* and a *trans* isomer, with the *trans* isomer being more abundant. It is the main component of the anise essential oil (80–90%), with minor components including para-anisaldehyde, estragole, and pseudoisoeugenyl-2-methylbutyrates, among others [18]. Anethole is also used in medicines as an expectorant, an antitussive, and antispasmodic for treating gastrointestinal tract illnesses. As a result, anise is found in a number of pharmaceutical products.

Basil

Basil (*Ocimum basilicum*) is a culinary herb belonging to the botanical family *Lamiaceae*. It has been used traditionally as a medicinal herb for the treatment of headaches, coughs, diarrhea, constipation, warts, worms, and kidney disorders. It is also a source of aroma compounds and essential oils containing biologically active constituents that possess antimicrobial and antifungal properties [19, 20]. Linalool is the main constituent of the essential oil of *O. basilicum* (28.6–60.6%), followed by estragole, methyl cinnamate, epi- α -cadinol, α -bergamotene, γ -cadinene (3.3–5.4%), germacrene D (1.1–3.3%), and camphor (1.1–3.1%). Other compounds, such as myrcene, pinene, terpineol, 1,8-cineole, eugenol, and methyleugenol, have been identified in basil leaves [21].

Caraway

Carvone is found to be the major compound in caraway. It has two mirror-image isomers, one of which smells like caraway, and the other smells like spearmint. Only S-carvone, the isomer smelling of caraway, is found in caraway seeds.

Cardamom

1,8-cineole is the major compound found in small cardamoms. The cardamom aroma is caused by the combination of 1,8-cineole compound and another compound, alpha-terpinyl acetate.

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Cayenne

Capsaicin is the major compound present in chili peppers, which gives them their spiciness.

Chili Pepper

The chili pepper is a fruit spice derived from plants from the genus *Capsicum*, originated in Mexico and was brought to Asia by Portuguese navigators during the sixteenth century. Chili peppers have a taste that is pungent, hot, and somewhat sweet (depending on the variety and type). Mild or sweet peppers contain similar constituents as *Capsicum* but with little or no pungent components. Chili peppers are used as food colorants, flavoring agents, predator repellants, and as a source of pain relief. The compounds responsible for the "hot" flavor of chili peppers are called capsaicinoids, with capsaicin being the best known. *Capsicum* contains up to 1.5% (by weight) of pungent compounds, commonly composed of capsaicin, dihydrocapsaicin, and others. Other constituents present in chili pepper are carotenoids, vitamins A, C, and small amounts of volatile oils with more than 125 known components. Another class of capsaicin-like compounds found in chili pepper and non-pungent chili pepper are the capsinoids. Many positive health benefits have been ascribed to both capsaicin and capsinoids, including anticancer, anti-inflammatory, and analgesic effects [22].

Chives

Similar to onions and garlic, sulfur-containing organic compounds gives flavor to chives. One of the major contributors to chives is dipropyl disulfide.

Cloves

Eugenol is one of the main compounds found in cloves. More minor constituents also contribute to the characteristic odour, for example, compounds, such as methylamylketone and methylsalicylate. Eugenol is named after the scientific name for cloves, and along with its derivative compounds, it is used in perfumery & flavoring.

Cinnamon

Cinnamon is a bark spice obtained from the inner bark of several tree species from the genus *Cinnamomum*. Cinnamon is native to India, Sri Lanka, Bangladesh, and Myanmar, and it was imported to Egypt 4000 years ago [23]. In addition to its common culinary, also used as condiment and flavoring material, cinnamon is widely known for its anti-diabetic and glucose-lowering effects [24]. The flavor of cinnamon is due to an aromatic essential oil that is largely

composed of cinnamaldehyde (up to 90%); however, there are at least 80 other compounds known to be present in cinnamon oil, including cinnamyl alcohol, cinnamyl acetate, eugenol, and various coumarins that contribute to its overall flavor and aroma [25].

Coriander Leaves

One of the main component of cinnamon leaf oil extract is 2-decenoic acid. The composition also includes many different aldehydes, primarily those of 9-10 carbons in length.

Coriander Seeds

The main compound in coriander seeds is linalool. It has two mirror-image isomers, one of which is known as coriandrol. The other mirror-image isomer is found in lavender and sweet basil.

Cumin

Cuminaldehyde is the main contributor to cumin's warm aroma. Other constituents include a range of other aldehyde compounds.

Dill

Carvone is the main compound found in dill. The spearmint isomer of carvone is used in the manufacture of chewing gum, spearmint chewing gum. It is produced by being soaked in carvone.

Fennel

Fennel is a seed (and bulb) spice, as well as a leaf herb, that is derived from *Foeniculum vulgare*. Fennel is a highly aromatic and flavorful herb/spice and is one of the primary ingredients of absinthe. The distinctive licorice flavor and aroma from fennel comes from anethole. Other compounds known to be in fennel include estragole, fenchone, 1,8-cineole (eucalyptol), and *p*-allylphenol. In addition to its use in culinary applications, fennel has long been used as a medicinal herb to treat gastrointestinal illness and upper respiratory tract infections as well as to increase milk production in breastfeeding mothers through the consumption of fennel tea.

Ginger

Ginger (*Zingiber officinale*) is a root or rhizome-based spice derived from the ginger plant, a member of the turmeric family (both are from *Zingiberaceae*).

Ginger is believed to have originated in India and is widely used as a culinary additive as a hot, fragrant spice as well as a popular medicine. In addition to ginger's well-known use as a treatment for nausea, many components in ginger have been found to have anti-inflammatory, antibacterial, antipyretic, antilipidemic, antitumorigenic, and antiangiogenic effects [26, 27]. A variety of active components have been identified in the oleoresins of ginger, including zingerone, gingerols (6-, 8-, and 10-gingerols), and shogaols (6-, 8-, and 10-shogaols) [28]. Gingerols (especially 6-gingerol) are the major pungent components in the fresh ginger rhizome.

Lemongrass

Citral is a mix of two different isomeric aldehydes, neral and geranial. Citral is also used in perfumery for its citrus odour.

Mace

Mace's chemical composition is similar to that of nutmeg, as they are both obtained from the same plant. The compound of the highest concentration in the essential oil of mace is terpinen-4-ol. It is also found in the essential oil of the tea tree.

Marjoram

Sabinene hydrate is the main component of the extracted oil of marjoram and is responsible, along with other compounds of the terpene family, for the characteristic flavor of the herb.

Mint

The major compound in mint leaves is menthol. This compound is also a popular flavoring for chewing gum and toothpaste and is also used in menthol cigarettes.

Nutmeg

Nutmeg is a fragrant flavoring spice coming from the seed of *Myristica fragrans* (belonging to the *Myristicaceae* family), an evergreen tree indigenous to the Banda Islands in the Moluccas (or Spice Islands) of Indonesia. The nutmeg essential oil is obtained by steam distillation of ground nutmeg, and it is used widely in the perfumery and pharmaceutical industries. This volatile fraction typically contains sabinene (21.38%), 4-terpineol (13.92%), and myristicin (13.5 7%), as well as portions of safrole, elimicin, terpineol, α -pinene d-camphene, limonene, linalool, and isoeugeunol [29].

Paprika

Paprika is a ground spice made from the red, air-dried fruits of larger and sweeter varieties of the plant *Capsicum annuum*, which is also called bell pepper or sweet pepper. Paprika can also be modified with the addition of more pungent chili pepper and cayenne pepper. The red, orange or yellow color of paprika is due to its content of carotenoids. Paprika carotenoids, particularly capsanthin and capsorubin, have been reported to have a strong antioxidant activity [30]. Based on these results, cucurbitaxanthin A, capsanthin, capsanthone, and cryptocapsin could be potential paprika-specific carotenoid biomarkers. However, further analyses using untargeted MS-based approaches should be conducted to evaluate other possible biomarkers of paprika intake.

Parsley

Parsley (*Petroselinum crispum*) is a herb belonging to the *Apiaceae* family. It is native to the central Mediterranean region. Fresh parsley has a clean, green aroma with a versatile, fresh taste that is slightly peppery with an after taste of green apple. Parsley is a source of several flavonoids, especially luteolin and apigenin [31]. Apigenin is associated with anti-inflammatory activities as it appears to downregulate or inhibit cyclo-oxygenoase-2 (COX-2). Apigenin has also been identified as a potential cancer chemopreventive agent [32]. The major essential oil found in parsley leaves is 1, 3, 8-*p*-menthatriene, but other components are also present in lesser amounts, including myristicin and limonene [33, 34].

Pepper

Piperine is the major constituent of the oil that can be extracted from black pepper and is the main compound that gives black pepper its pungency. An isomer of piperine, chavicine, also contributes to the pungency.

Peppermint

Peppermint and spearmint are herbs that belong to the *Laminacea* family. Spearmint (*Mentha spicata*) is believed to be one of the oldest mints. The active constituents of spearmint include spearmint oil, various flavonoids (diosmin, diosmetin), phenolic acids, and lignans. The most abundant compound in spearmint oil is carvone, which gives spearmint its distinctive smell. Peppermint has a high menthol content (40.7%), along with menthone (23.4%), and other essential oils, such as menthyl acetate (4.2%), 1,8-cineole (5.3%), limonene (2. 6%), menthofuran (3.7%), and β -caryophyllene (1.7%) [35]. Peppermint leaves can also be added to herbal tea, ice cream, chewing gum, toothpaste, shampoo, *etc.*, for enhancing their flavor. Peppermint leaves are often used alone or with other

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herbs in herbal teas (tisanes, infusions), ice cream, confectionery, chewing gum, toothpaste, and shampoos. Menthol activates cold-sensitive receptors in the skin and mucosal tissues and is the primary source of the cooling sensation that follows the topical application of peppermint oil. Peppermint also contains terpenoids and flavonoids such as eriocitrin, hesperidin, and kaempferol 7-O-rutinoside.

Saffron

Saffron is among the world's most costly spices. It comes from the dried flower stigma of Crocus sativus. Saffron contains more than 150 volatile and aromavielding compounds. Safranal (2,6,6-trimethyl-1,3-cyclohexadiee-1-carboxaldehyde) is the major compound (70%) in the volatile fraction of saffron [36]. Saffron also has a number of non-volatile active components, many of which are carotenoids, including zeaxanthin, lycopene, and various α - and β carotenes. However, the golden yellow-orange color of saffron is primarily the result of the carotenoid α -crocin, a glycosylester of crocetin. Picrocrocin(4-(β d-glucopyranosyloxy)-2,6,6-trimethyl-1-cyclohexene-1-carboxaldehyde) has also been found in saffron spice from 0.8 to 26.6% on a dry basis. This compound is responsible for saffron's bitter taste. In addition, saffron contains two important vitamins, riboflavin and thiamine. Saffron extracts and tinctures have been used as antispasmodic agents, gingival sedatives, nerve sedatives, expectorants, stimulants, and aphrodisiacs.

Sage

Sage or *Salvia officinalis* is a medicinal plant belonging to the *Lamiaceae* family. It is an aromatic herb native to the Mediterranean region but is now widely distributed throughout the world. Sage has been used in traditional medicine for the treatment of seizures, ulcers, gout, rheumatism, inflammation, dizziness, tremors, paralysis, diarrhea, and hyperglycemia [37]. Sage has a savory, slightly peppery flavor. It is strongly aromatic and is characterized by a medicinal, lemony, or bitter taste. It is used for seasoning and flavoring in many different foods, including sausages and stuffing. The major components present in sage are α -thujone (11.55–19.23%), viridiflorol (9.94–19.46%), 1, 8-cineole (8.85–15.60%), camphor (5.08–15.06%), manool (5.52–13.06%), β -caryophyllene (2.63–9.24%), α -humulene (1.93–8.94%), and β -thujone (5.45–6.17%) [38]. Some of the major phenolic compounds found in sage are rosmarinic acid, caffeic acid, carnosol, and carnosic acid.

Tarragon

Tarragon (Artemisia dracunculus), also known as estragon, is a perennial herb

belonging to the *Asteraceae* (daisy) family. It is widespread across much of Eurasia and North America and is cultivated for culinary and medicinal purposes. *In vitro* pharmacological studies indicate that tarragon has antibacterial, antifungal, and antiplatelet activity [39]. *In vivo* pharmacological studies have shown that tarragon has anti-inflammatory, hepatoprotective, antihyperglycemic, and antioxidant activity [39]. The major components of Russian tarragon are reported to be terpinen-4-ol, sabinene, and elemicin. Methyleugenol and estragole are usually present in tarragon oils at about 10 and 3%, respectively. However, estragole is one of the predominant compounds in the essential oil of French tarragon, constituting up to 82% [39]. *Trans*-anethole (21.1%), α -trans-ocimene (20.6%), limonene (12.4%), α -pinene (5.1%), and allo-ocimene (4.8%) are the other main components of tarragon.

Turmeric

Turmeric is a rhizomatous herbaceous perennial plant (*Curcuma longa*) belonging to the ginger family, Zingiberaceae. It is a key ingredient in many Asian dishes and is used mainly as a coloring agent. The most notable phytochemical components of turmeric root include compounds called curcuminoids, such as curcumin (diferuloylmethane), demethoxycurcumin (DMC), and bisdemethoxycurcumin (BDMC). Curcumin is a polyphenolic molecule that constitutes 3.14% (on average) of powdered turmeric Curcumin is what gives the spice its yellow color [40]. The rhizome oils of turmeric contain more than 40 identifiable compounds, with the major constituents being α -turmerone (30–32%), aromaticturmerone (17–26%), and β -turmerone (15–18%). This interest is likely due to the multiple biological or health activities attributed to it, including antioxidant, antiinflammatory, and anti-tumor activities. Recent clinical studies with curcumin have demonstrated additional health benefits relating to treating immune deficiencies, improving cardiovascular health, treating depression [41], combating Alzheimer's disease, treating diabetes, arthritis, and inflammatory bowel disease [42].

Vanilla

The aroma of vanilla is mainly due to the compound vanillin, which accounts for 74-96% of the flavor & aroma compounds. Over 100 other volatile compounds have been detected, including acids, phenolic compounds, alcohols, and aldehydes.

Oregano, Rosemary and Thyme

Oregano, rosemary and thyme are well known for their beneficial health properties. For example, carnosic acid and some of the diterpenes abundant in rosemary and sage appear to exert anti-obesity effects (including body weight and lipid-lowering effects) [43]. Several compounds found in herbs from the *Laminaceae* family also exhibit antimicrobial activity, such as thymol, carvacrol, carnosol, rosmanol, and caffeic acid [44].

Oregano (*Origanum vulgare*) is a native herb to temperate western and southwestern Eurasia and the Mediterranean region. It has an aromatic, warm, and slightly bitter taste. Among the chemical compounds contributing to the flavor of oregano are carvacrol, thymol, limonene, pinene, ocimene, and caryophyllene. Oregano also contains polyphenols, including caffeic, *p*-coumaric, and rosmarinic acid, which confer antioxidant activity and prevent lipid peroxidation [45].

Rosemary (*Rosmarinus officinalis*) is native to the Mediterranean and Asia. The leaves are used as a flavoring agent in a variety of foods in traditional Mediterranean cuisine. They have a bitter, astringent taste and a very characteristic aroma. Rosemary contains a number of phytochemicals, including rosmarinic acid, camphor, caffeic acid, ursolic acid, betulinic acid, carnosic acid, and carnosol [45]. Major essential oils present in rosemary oil are borneol (26. 5%), α -terpinene (15.6%), and α -pinene (12.7%).

Thyme (*Thymus vulgaris*) is also a member of the *Lamiaceae* family, and it has been used in foods mainly for flavor, aroma, and food preservation. Thyme has also been used in folk medicine since the times of the ancient Egyptians, Greeks, and Romans. The leafy parts of thyme are often added to meat, fish, and food products and are also used as herbal medicinal products. The essential oils of common thyme contain 20–58% thymol and *p*-cymene (15–28%) as the most prevalent compounds, followed by linalool (0.7–6.5%), γ -terpinene (4–10%), carvacrol (1–4%), myrcene (1–3%), 1,8-cineole (0.8%), and borneol (0.7–1.7%) [46]. Thymol is the compound that provides the distinct flavor of thyme. It is also found in oregano and is used as one of many additives in cigarettes.

CONCLUSION

Spices and herbs have been used since the early days of humankind and are still used throughout the world for health promotion and treatment of various diseases. Plants are found to be the source of today's modern medicine and contribute largely to commercial drug preparation.

However, in many developing countries, herbal medicine is used as traditional medicine. Interests in food compounds from spices and herbs will continue to increase as well as research and technology that will develop better ways of growing spices and herbs that contain higher amounts of antioxidants. Over the last many years, several bioactive compounds have been isolated from various spices and herbs, which provide a scientific and medicinal basis to include spices

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and herbs in our diet.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The author declares no conflict of interest, financial or otherwise.

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REFERENCES

- Govindarajan R, Vijayakumar M, Pushpangadan P. Antioxidant approach to disease management and the role of 'Rasayana' herbs of Ayurveda. J Ethnopharmacol 2005; 99(2): 165-78. [http://dx.doi.org/10.1016/j.jep.2005.02.035] [PMID: 15894123]
- Puvaca N, Stanacev V, Glamocic D, *et al.* Beneficial effects of phytoadditives in broiler nutrition. Worlds Poult Sci J 2013; 69: 27-34.
 [http://dx.doi.org/10.1017/S0043933913000032]
- [3] Popovic S, Puvaca N, Kostadinovic LJ, *et al.* Effects of dietary essential oils on productive performance, blood lipid profile, enzyme activity and immunological response of broiler chickens. Arch Geflugelkd 2016; 80: 1-12.
- [4] Farr DR. Functional foods. Cancer Lett 1997; 114(1-2): 59-63.
 [http://dx.doi.org/10.1016/S0304-3835(97)04626-0] [PMID: 9103255]
- [5] Wang H, Cao G, Prior RL. Oxygen radical absorbing capacity of anthocyanins. J Agric Food Chem 1997; 45: 304-9.
 [http://dx.doi.org/10.1021/jf960421t]
- [6] Kitts DD, Wijewickreme AN, Hu C. Antioxidant properties of a North American ginseng extract. Mol Cell Biochem 2000; 203(1-2): 1-10. [http://dx.doi.org/10.1023/A:1007078414639] [PMID: 10724326]
- [7] Srivastava J, Lambert J, Vietmeyer N. Medicinal plants: An expanding role in development. World Bank Technical Paper 1996; 1-36.
- [8] Ganatra SH, Ramteke A. Studies Of Anti-Cancer Activities Of Naturally Occurring Terpene Extracted From The Seeds Of Pithcellobium Dulce, INT INST. Anticancer Res 2014; 34(12): 7479-80.
- Shan B, Cai YZ, Sun M, Corke H. Antioxidant capacity of 26 spice extracts and characterization of their phenolic constituents. J Agric Food Chem 2005; 53(20): 7749-59.
 [http://dx.doi.org/10.1021/jf051513y] [PMID: 16190627]
- Srinivasan K. Antioxidant potential of spices and their active constituents. Crit Rev Food Sci Nutr 2014; 54(3): 352-72.
 [http://dx.doi.org/10.1080/10408398.2011.585525] [PMID: 24188307]
- [11] Surh YJ. Anti-tumor promoting potential of selected spice ingredients with antioxidative and antiinflammatory activities: a short review. Food Chem Toxicol 2002; 40(8): 1091-7. [http://dx.doi.org/10.1016/S0278-6915(02)00037-6] [PMID: 12067569]
- [12] Zheng W, Wang SY. Antioxidant activity and phenolic compounds in selected herbs. J Agric Food Chem 2001; 49(11): 5165-70.

[http://dx.doi.org/10.1021/jf010697n] [PMID: 11714298]

- [13] Sachan AKR, Kumar S, Kumari K, Singh D. Herbs and Spices- Biomarkers of Intake Based on Human Intervention Studies – A Systematic Review. Journal of Medicinal Plants Studies 2018; 6(3): 116-22.
- [14] Dubey S. Indian Spices and their Medicinal Value. Indian Journal of Pharmaceutical Education and Research 2017; 51(3): 330-2.
 [http://dx.doi.org/10.5530/ijper.51.3s.41]
- [15] Chemical Compounds in Herbs and Spices. 2014.
- [16] Vázquez-Fresno R, Rosana ARR, Sajed T, Onookome-Okome T, Wishart NA, Wishart DS. Herbs and Spices- Biomarkers of Intake Based on Human Intervention Studies - A Systematic Review. Genes Nutr 2019; 14: 18.
 [http://dx.doi.org/10.1186/s12263-019-0636-8] [PMID: 31143299]
- [17] Marinov V, Valcheva-Kuzmanova S. Review on the pharmacological activities of anethole. Scr Sci Pharm 2015; 22: 14-9.
 [http://dx.doi.org/10.14748/ssp.v2i2.1141]
- [18] Rodrigues VM, Rosa PT, Marques MO, Petenate AJ, Meireles MA. Supercritical extraction of essential oil from aniseed (Pimpinella anisum L) using CO2: solubility, kinetics, and composition data. J Agric Food Chem 2003; 51(6): 1518-23. [http://dx.doi.org/10.1021/jf0257493] [PMID: 12617576]
- [19] Hussain AI, Anwar F, Hussain Sherazi ST, Przybylski R. Chemical composition, antioxidant and antimicrobial activities of basil (Ocimum basilicum) essential oils depends on seasonal variations. Food Chem 2008; 108(3): 986-95. [http://dx.doi.org/10.1016/j.foodchem.2007.12.010] [PMID: 26065762]
- [20] Wannissorn B, Jarikasem S, Siriwangchai T, Thubthimthed S. Antibacterial properties of essential oils from Thai medicinal plants. Fitoterapia 2005; 76(2): 233-6. [http://dx.doi.org/10.1016/j.fitote.2004.12.009] [PMID: 15752638]
- [21] Politeo O, Jukic M, Milos M. Chemical composition and antioxidant capacity of free volatile aglycones from basil (*Ocimum basilicum* L.) compared with its essential oil. Food Chem 2007; 1011: 379-85.
 [http://dx.doi.org/10.1016/j.foodchem.2006.01.045]
- [22] Chaiyata P, Puttadechakum S, Komindr S. Effect of chili pepper (Capsicum frutescens) ingestion on plasma glucose response and metabolic rate in Thai women. J Med Assoc Thai 2003; 86(9): 854-60. [PMID: 14649970]
- [23] Parry JW. The story of spices. Econ Bot 1955; 92: 190-207. [http://dx.doi.org/10.1007/BF02898800]
- [24] Khan A, Safdar M, Ali Khan MM, Khattak KN, Anderson RA. Cinnamon improves glucose and lipids of people with type 2 diabetes. Diabetes Care 2003; 26(12): 3215-8. [http://dx.doi.org/10.2337/diacare.26.12.3215] [PMID: 14633804]
- [25] Jayaprakasha GK, Rao LJ. Chemistry, biogenesis, and biological activities of *Cinnamomum zeylanicum*. Crit Rev Food Sci Nutr 2011; 51(6): 547-62. [http://dx.doi.org/10.1080/10408391003699550] [PMID: 21929331]
- [26] Surh Y-J. Anti-tumor promoting potential of selected spice ingredients with antioxidative and antiinflammatory activities: a short review. Food Chem Toxicol 2002; 40(8): 1091-7. [http://dx.doi.org/10.1016/S0278-6915(02)00037-6] [PMID: 12067569]
- [27] Park M, Bae J, Lee DS. Antibacterial activity of [10]-gingerol and [12]-gingerol isolated from ginger rhizome against periodontal bacteria. Phytother Res 2008; 22(11): 1446-9. [http://dx.doi.org/10.1002/ptr.2473] [PMID: 18814211]

- [28] Rahmani AH, Shabrmi FM, Aly SM. Active ingredients of ginger as potential candidates in the prevention and treatment of diseases *via* modulation of biological activities. Int J Physiol Pathophysiol Pharmacol 2014; 6(2): 125-36.
 [PMID: 25057339]
- [29] Muchtaridi SA, Subarnas A, Apriyantono A, Mustarichie R. Identification of compounds in the essential oil of nutmeg seeds (*Myristica fragrans* Houtt.) that inhibit locomotor activity in mice. Int J Mol Sci 2010; 11(11): 4771-81. [http://dx.doi.org/10.3390/ijms11114771] [PMID: 21151471]
- [30] Nishino A, Yasui H, Maoka T. Reaction of paprika carotenoids, Capsanthin and Capsorubin, with reactive oxygen species. J Agric Food Chem 2016; 64(23): 4786-92. [http://dx.doi.org/10.1021/acs.jafc.6b01706] [PMID: 27229653]
- [31] Maher HM, Al-Zoman NZ, Al-Shehri MM, *et al.* Determination of Luteolin and Apigenin in herbs by capillary electrophoresis with diode Array detection. Instrum Sci Technol 2015; 436: 611-25. [http://dx.doi.org/10.1080/10739149.2015.1038560]
- [32] Shukla S, Gupta S. Apigenin: a promising molecule for cancer prevention. Pharm Res 2010; 27(6): 962-78.
 [http://dx.doi.org/10.1007/s11095-010-0089-7] [PMID: 20306120]
- [33] Kokkini S, Karousou R, Lanaras T. Essential oils of spearmint (Carvone-rich) plants from the island of Crete (Greece). Biochem Syst Ecol 1995; 234: 425-30. [http://dx.doi.org/10.1016/0305-1978(95)00021-L]
- [34] Zhang H, Chen F, Wang X, Yao H-Y. Evaluation of antioxidant activity of parsley (Petroselinum crispum) essential oil and identification of its antioxidant constituents. Food Res Int 2006; 39: 833-9. [http://dx.doi.org/10.1016/j.foodres.2006.03.007]
- [35] Schmidt E, Bail S, Buchbauer G, et al. Chemical composition, olfactory evaluation and antioxidant effects of essential oil from Mentha x piperita. Nat Prod Commun 2009; 4(8): 1107-12. [http://dx.doi.org/10.1177/1934578X0900400819] [PMID: 19768994]
- [36] Maggi L, Sánchez AM, Carmona M, et al. Rapid determination of safranal in the quality control of saffron spice (Crocus sativus L.). Food Chem 2011; 1271: 369-73. [http://dx.doi.org/10.1016/j.foodchem.2011.01.028]
- [37] Ghorbani A, Esmaeilizadeh M. Pharmacological properties of Salvia officinalis and its components. J Tradit Complement Med 2017; 7(4): 433-40.
 [http://dx.doi.org/10.1016/j.jtcme.2016.12.014] [PMID: 29034191]
- [38] Ben Farhat M, Jordán MJ, Chaouech-Hamada R, Landoulsi A, Sotomayor JA. Variations in essential oil, phenolic compounds, and antioxidant activity of tunisian cultivated Salvia officinalis L. J Agric Food Chem 2009; 57(21): 10349-56. [http://dx.doi.org/10.1021/jf901877x] [PMID: 19886685]
- [39] Obolskiy D, Pischel I, Feistel B, Glotov N, Heinrich M. Artemisia dracunculus L. (tarragon): a critical review of its traditional use, chemical composition, pharmacology, and safety. J Agric Food Chem 2011; 59(21): 11367-84. [http://dx.doi.org/10.1021/jf202277w] [PMID: 21942448]
- [40] Baum L, Cheung SK, Mok VC, et al. Curcumin effects on blood lipid profile in a 6-month human study. Pharmacol Res 2007; 56(6): 509-14. [http://dx.doi.org/10.1016/j.phrs.2007.09.013] [PMID: 17951067]
- [41] Seo H-J, Wang S-M, Han C, *et al.* Curcumin as a putative antidepressant. Expert Rev Neurother 2015; 15(3): 269-80.
 [http://dx.doi.org/10.1586/14737175.2015.1008457] [PMID: 25644944]
- [42] Sreedhar R, Arumugam S, Thandavarayan RA, Karuppagounder V, Watanabe K. Curcumin as a therapeutic agent in the chemoprevention of inflammatory bowel disease. Drug Discov Today 2016;

21(5): 843-9. [http://dx.doi.org/10.1016/j.drudis.2016.03.007] [PMID: 26995272]

- [43] Ibarra A, Cases J, Roller M, Chiralt-Boix A, Coussaert A, Ripoll C. Carnosic acid-rich rosemary (Rosmarinus officinalis L.) leaf extract limits weight gain and improves cholesterol levels and glycaemia in mice on a high-fat diet. Br J Nutr 2011; 106(8): 1182-9. [http://dx.doi.org/10.1017/S0007114511001620] [PMID: 21676274]
- [44] Lai PK, Roy J. Antimicrobial and chemopreventive properties of herbs and spices. Curr Med Chem 2004; 11(11): 1451-60.
 [http://dx.doi.org/10.2174/0929867043365107] [PMID: 15180577]
- [45] Vallverdú-Queralt A, Regueiro J, Martínez-Huélamo M, Rinaldi Alvarenga JF, Leal LN, Lamuela-Raventos RM. A comprehensive study on the phenolic profile of widely used culinary herbs and spices: rosemary, thyme, oregano, cinnamon, cumin and bay. Food Chem 2014; 154: 299-307. [http://dx.doi.org/10.1016/j.foodchem.2013.12.106] [PMID: 24518346]
- [46] Soković M, Glamočlija J, Marin PD, Brkić D, van Griensven LJ. Antibacterial effects of the essential oils of commonly consumed medicinal herbs using an *in vitro* model. Molecules 2010; 15(11): 7532-46.

[http://dx.doi.org/10.3390/molecules15117532] [PMID: 21030907]

Naturally-derived Analgesics and Anti-Inflammatory Agents

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Abstract: Medications for the treatment of pain and inflammatory disorders, although effective, their chronic use and/or misuse can lead to serious side effects. In this regard, naturally-derived antinociceptive and anti-inflammatory agents have emerged as alternatives to synthetically marketed drugs. The current review covers all the nutraceuticals and phytochemicals – derived from medicinal plants– which have been reported to possess analgesic and/or anti-inflammatory effects over the period between 2018 up to June 2020.

Keywords: Pain, Inflammation, Marine organisms, Medicinal plants, Natural products.

INTRODUCTION

Inflammation is one of the responses of innate immunity to infection or tissue injury. Despite being essential for maintaining normal homeostasis, prolonged inflammation could likewise be a sign of a pathological condition resulting from chronic diseases, which could be detected by the presence of inflammatory markers [1]. The molecular mechanisms of inflammation are complicated, and they are initiated by the recognition of the allergen by germline-encoded specific pattern recognition receptors such as Toll-like receptors (TLR), retinoic acid-inducible gene-I (RIG-1)-like receptors, C-type lectin receptors (CLR), and the nucleotide-binding oligomerization domain (NOD) like receptor [1, 2]. These inflammatory responses end up with some typical symptoms, including redness,

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swelling, pain, and heat [1, 3]. Activation of these receptors by bacteria triggers the release of inflammatory cytokines, which in turn mediate the production of Creactive protein (CRP), among others, which promote the secretion of prostaglandins [3]. The latter are responsible for the symptoms of inflammation. Viral infections, on the other hand, trigger the release of type-I interferons, while parasitic infections and other allergens increase the production of histamine and interleukins. The common pathways, which play a major role in inflammation, are the NF-kB, JAK-STAT, and MAPK [3]. Although pain is one of the consequences of inflammation, it could also be a separate sign, even with no existence of an ongoing inflammatory process. Non-steroidal anti-inflammatory drugs (NSAIDs) and opioid analgesics are among the commonly used medications for pain treatment. However, they show many side effects, which lead to patient discomfort, especially in chronic use [4, 5]. Since the last decade, many natural products have proven efficacy for the management of pain and inflammation [2]. Herein, we report on the latest updates in the field of antiinflammatory natural products, which have been reported over the interval between 2018 and June 2020.

Medicinal Plant Extracts with Reported Analgesic and/or Anti-Inflammatory Activities (Table 1)

Spices/plant Name	Mechanism of Action	Refs.
Acanthus ilicifolis Linn.	In vivo analgesic effect for the chloroform and petroleum ether fractions In vitro antioxidant activity	[8]
Ajuga laxmannii (Murray) Benth.	Inhibition of phagocytosis and decreasing the total leukocytes <i>in</i> <i>vitro</i> <i>In vitro</i> antioxidant activity	[9]
Anadenanthera colubrina var. cebil (Griseb.)	<i>In vivo</i> anti-inflammatory potential in paw oedema model Reduction of IL-12 production and TNF-α release accompanied by elevation in IL-10 Inhibition of the production of nitric oxide	[10]
Asphodelus microcarpus Salzm. & Viv.	Strong antioxidant capacity Reduction of the <i>in vivo</i> paw and ear edema induced by carrageenan and xylene	[11]
Athyrium multidentatum (Doll.) Ching	Decreased the expression of iNOS and COX-2 enzymes Downregulation of the <i>in vitro</i> and <i>in vivo</i> mRNA expression of IL-6, IL-1 β , and TNF- α	[12]
Backhousia citriodora (Lemon myrtle)	Antioxidant properties Reduction of IL6, TNF-α, and NO levels	[13]

Table 1. List of previously reported spices with anti-inflammatory activities.

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Spices/plant Name	Mechanism of Action	Refs.
Bistorta officinalis (Bistort)	Inhibition of the release of pro-inflammatory cytokines after LPS stimulation	[14]
Boswellia species oleogum resin	Antioxidant effect Immunomodulatory effects Decreased paw edema, pleural exudates, pain, and fever <i>in vivo</i> .	[16]
Buddleja officinalis Maxim.	Suppression of NO, TNF- α , and IL-1 β in vitro	[17]
Caesalpinia minax Hance	Reduced the proinflammatory cytokines, IL-6, IL-1 β , and TNF- α <i>in vivo</i>	[18]
Caesalpinia sappan L.	Reduced the production of TNF- α	[20]
Calotropis procera	Reduction of myeloperoxidase activity in neutrophils Reduction of the expression of adhesion molecules ICAM-1 and Iba-1	[21]
Centipeda minima (L.) A.Br.	Inhibition of NF- κ B TNF- α , IL-1 β , COX-2, NOX-2, NOX-4, ROS, PGE ₂ , and iNOS <i>in vitro</i> .	[22]
Cissus gongylodes (grapevine)	Inhibition of COX and LOX inflammatory pathways and reducing the concentrations of PGE_2 and LTB_4	[23]
Citrus bergamia	Downregulation of the pro-inflammatory cytokines IL-1 β , IL-6, TNF- α , and the inhibition of the release of NO, PGE ₂ , ROS.	[24]
Cyclamen africanum B. et R.	Strong antioxidant activity	[25]
Elephantopus scaber Linn. herb	Inhibition of the transcription and translation of iNOS and the blockade of NF-κB signaling pathway	[26]
Elsholtzia ciliata (Thunb.) Hyl.	Inhibition of the secretion of the pro-inflammatory cytokines IL-6, TNF- α . and PGE ₂	[27]
Epigynum auritum	Inhibition of the release of TNF- α , IL-6, and NO	[28]
Eugenia stipitate McVaugh. Eupatorium japonicum Thunb.	Reduction of the volume of edema and the migration of leukocytes and neutrophils <i>in vivo</i> Suppressing the expression of IL-1β, the activation of NF-kβ, and	[29] [30]
Forsythia suspensa (Thunb.)	the transcription of MMP-9 Inhibition of NF-kβ pathway <i>via</i> the activation of A20 protein as well as the stimulation of the Nrf2 signaling cascade	[31]
Galinsoga parviflora Cav.	Reduction of IL-6 levels and strong anti-hyaluronidase activity	[32]
Garcinia cambogia Gaertn. and Pothos scandens L.	Suppression for 5-LO in human neutrophils Inhibition for isolated human 5-LO and mPGE ₂ S-1	[33]
Halosarcia indica Willd.	Dose dependent reduction in carrageenan-induced edema and reduction of writhing responses in rats	[34]
Hyusopus cuspidatus Boriss.	Reduction of serum nitric oxide, prostaglandin E_2 , IL-6, IL-1 β , and TNF- α	[35]
Indigofera argentea Burm. F.	Central action on the opioid receptors and the inhibition of COX and LOX metabolites in the peripheral tissues	[36]
Kadsura heteroclita	Reduction of the cytokines level of TNF- α , IL-1 β , and IL-6	[37]

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Spices/plant Name	Mechanism of Action	
Kleinia pendula (Forssk.) herb	Significant analgesic and anti-inflammatory properties in vivo	[38]
Larrea species and Zuccagnia punctate Cav.	Inhibition of lipoxygenase enzyme	[39]
Ligusticum chuanxiong and Radix Paeoniae lactiflora	Reduction of the levels of IL-1 β , IL-6, IL-12, and IFN-c <i>in vivo</i>	[41]
Manilkara zapota	in vivo pain-relieving effect	[42]
Mitrella kentia	Inhibition of the production of PGE ₂ , TXB ₂ , and its antagonism on PAF	[43]
Mycetia cauliflora Reinw.	Reduction of iNOS, NO, IL-1 β , inhibition of the nuclear translocation of the transcription factor p65 and p50 protein	[44]
Niebuhria Apetala Dunn.	Reduction of the thermal-induced pain in rats in a dose dependent manner	[45]
Ocimum forskolei Benth.	Reduction of the carrageenan-induced paw edema	[46]
Opuntia humifusa Raf.	Inhibition of NO production and decreasing the levels of iNOS	[47]
Paeoniae alba and Atractylodis macrocephalae	Reduction of NO production and the proinflammatory cytokines like TNF-α, IL-6, and MCP-1	[48]
Panax ginseng	Decreasing the inflammatory symptoms by reducing IL-6 and TNF-α levels <i>in vivo</i>	[49]
Phyllanthus amarus Schum. & Thonn.	Inhibition of the production of the pro-inflammatory PGE ₂ , IL-1 β , and TNF- α	[50]
Physalis angulata L.	Inhibition of the proinflammatory cytokines IL-1β, TNF-α, IL-6, IL-12, iNOS, COX-2 and increasing the levels of the anti- inflammatory genes of IL-10, arginase-1, TGF-β	[51]
Phytolacca dodecandra	Reduction of pain sensation and edema in the tested mice in vivo	[52]
Picria Fel-Terrae Lour	Downregulation of TNF- α , IL-6, IL-1 β , COX-2, and iNOS	[53]
Pituranthos scoparius Coss. and Dur.	Dose-dependant inhibition of edema was observed in mice	[54]
Pterocephalus hookeri (C.B. Clarke) Höeck	Interfering with NF-кВ pathway	[55]
Reineckia carnea (Andr.) Kunth	Inhibition of the production of NO	[58]
Rhodiola crenulate (golden root) Sambucus australis Cham. & Schltdl	Reduction of the levels of the inflammatory mediators like IL-1β, IL-6, and NO Reduction of interleukins 4 and 5, interferon-γ, nitric oxide, and decreased expression of NF-κB.	[59] [60]
Sarcopoterium spinosum	Increasing the expression of the anti-inflammatory genes IL-10 and Arg-1, the inhibition of the NF- $_{x}$ B inflammatory pathway and Akt phosphorylation	[61]
Scrophularia megalantha Rech. f.	Reducing the levels of IL-17 and interferon-γ while increasing the levels of IL-10	[62]

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Spices/plant Name	Mechanism of Action	Refs.
Scutellaria barbata D. Don	Reducing NO, PGE_2 , IL-6, and IL-1 β production and decreasing the expression of p-JNK and p-ERK	[63]
Sigesbeckiae species	Inhibition of the nuclear factor _k B (NF- _x B) signaling pathway and mitogen-activated protein kinases (MAPKs)	[64]
Sinapis semen	Suppressed the protein and mRNA expression of TNF-α, IL-1β, and IL-6	[66]
Sophora flavescens Ait.	Reduction of IL-6, TNF-α, NO, and MCP-1 levels	[67]
Spatholobus suberectus	Strong free radical scavenging potential and the reduction of iNOS and COX-II expression	[68]
Tephrosia linearis (Willd.) Pers.	Interfering with the release of IL-2, GM-CSF, and TNF- α in vitro	[70]
Thymus zygis subsp. zygis	Inhibition of the release of nitric oxide	[71]

Acanthus Ilicifolis Linn

The "Holy leaved acanthus", *Acanthus ilicifolis* (Acanthaceae), is a perennial herb native to Australia and South Asia. The herb was traditionally used in China and India as a remedy for numerous ailments. Diverse phytochemicals were isolated from their different parts (leaves, bark, fruits, and roots), including terpenoids, flavonoids, steroids, alkaloids, phenolic compounds, and lignins [6, 7]. The chloroform and petroleum ether fractions demonstrated a significant *in vivo* analgesic effect when evaluated using the acetic acid-induced writhing method compared to the standard diclofenac. The plant extract likewise displayed potential *in vitro* antioxidant activity compared to standard ascorbic acid when evaluated using DPPH assay [8].

Ajuga Laxmannii (Murray) Benth

Ajuga laxmannii (Labiatae) is a Romanian herb used as a galactagogue and as an anti-inflammatory remedy. The latter effect is due to its antiradical capacity, which is attributed to the high phenolic content (total content ca. 67.6 ± 1.5 mg eqivalent of gallic agid/ g ethanol extract), including iridoid glycosides (*i.e.*, harpagide, aucubin, catalpol, harpagoside), flavonoids (*i.e.*, luteolin, apigenin, quercetin, rutin), and phytosterols (*i.e.*, stigmasterol, β -sitosterol, ergosterol). The antioxidant effect of Ajuga laxmannii ethanolic extract was evaluated using DPPH (showing IC₅₀ value of 22.6 ± 0.8 µg/ml) and ABTS free radical assay. Moreover, the extract inhibited phagocytosis and decreased the total leukocytes *in vitro*, which was comparable to the positive control (diclofenac) [9].

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Anadenanthera Colubrina Var. Cebil (Griseb.)

The ethanol extract of *Anadenanthera colubrine* (Fabaceaea) leaves exhibited significant *in vivo* anti-inflammatory potential in paw oedema model compared to standard dexamethasone drug. Marked reduction in IL-12 production accompanied by elevation in IL-10 were observed. The extract at 50 μ g/mL inhibited the production of nitric oxide by 80% and reduced the release of TNF- α by more than 50%. Anti-oxidant activity of the extract was confirmed by DPPH assay. The anti-inflammatory and anti-oxidant were attributed to the flavonoid and phenolic content of the extract [10].

Asphodelus Microcarpus Salzm. & Viv

Asphodelus microcarpus (Liliaceae) is an Algerian plant commonly used in the traditional medicine for the treatment of pain and inflammation especially in the cases of rheumatoid arthritis, toothache, asthma, and bronchitis. The crude extract is rich in polyphenols and flavonoids with the highest percentage being in the leaves (equivalent to 755 mg gallic acid for polyphenols and 42 mg rutin for flavonoids). This finding corroborates its strong antioxidant capacity in scavenging the free radicals, which initiate inflammatory cascades. The anti-inflammatory effect of the extract was likewise evaluated by its ability to reduce the *in-vivo* paw and ear edema induced by carrageenan and xylene, respectively, within 6 h after the administration of an oral dose of 500 mg/kg [11].

Athyrium Multidentatum (Doll.) Ching

Athyrium multidentatum (Athyriaceae) is a perennial herb widely distributed in China. This nutritional potherb has a strong antioxidant and pain relieving effect due to its flavonoid (total flavonoid content ca. 36 ± 2.3 mg/g extract) and phenolic acids content mainly myricetin, quercetin, and kaempferol. The herbal extract decreased the expression of iNOS and COX-2 enzymes leading to significant reduction in NO and PGE₂, respectively. Downregulation of LPS-stimulated acute lung injury mice models, mRNA expression of IL-6, IL-1 β , and TNF- α , decreased phosphorylation of ERK (extracellular signal-regulated kinase s), JNK (c-Jun-N-terminal kinases), and other relevant proteins of the TLR-4 pathway [12].

Backhousia Citriodora (Lemon Myrtle)

The lemon myrtle (Myrtaceae) is a shrub native to the rainforests of Australia and

grows mostly 50-800 m above the sea level. Its alcoholic extract is rich in flavonoids (flavonoid content is 14.5 mg/g extract) and polyphenols (total polyphenol content is 118.7 mg/g extract). Its anti-inflammatory activity is due to its antioxidant properties and is mediated through the reduction of IL6, TNF- α , and NO levels in LPS-induced murine macrophages *in vitro* [13].

Bistorta Officinalis (Bistort)

The rhizomes of *Bistorta officinalis* (Polygonaceae) are traditionally used in Asia and Europe for topical treatment of skin conditions. Pawłowska *et al.* reported the influence of the infusion of the subterranean parts of *B. officinalis* and their isolated phytoconstituents on the release of pro-inflammatory cytokines after LPS stimulation. The study demonstrated the *in vitro* activity of the infusion to modulate the inflammatory response of human neutrophils involved in wound healing and other skin problems justifying its ethnopharmacological use [14].

Boswellia species oleogum resin (Frankincense, olibanum)

Many studies revealed the therapeutic potential of the frankincense obtained from the trees of *Boswellia* species (family Burseraceae) in the management of many inflammatory and autoimmune disorders and in cancer therapy. It has been cultivated since ancient ages in China, India, and the Arabic peninsula and it has been used traditionally in the treatment of constipation, flatulence, CNS, and other inflammatory diseases. In the Islamic traditional medicine, the plant was used as a remedy for burns, bruises, infections, and eye sores. At the beginning of the 20^{th} Century, the plant was officially used in the management of inflammation around Europe [15]. Although genus *Boswellia* comprises ca. 30 species or even more, only five of which are mainly used for the production of the frankincense namely B. serrata, B. sacra, B. frerana, B. carterii, and B. papyrifera. The oleogum resin of the frankincense is a complex mixture of more than 200 phytochemicals of different classes including terpenoids (tetra and pentacyclic), polyphenols, tannins, saponins, alkaloids, and sugars. Despite the diversity of its constituents, its activity is mainly pertained to its pentacyclic boswellic acids. The mechanisms pertaining to its anti-inflammatory effects are [16]:

• Antioxidant effect through its free radical scavenging activity including ROS (reactive oxygen species) and RNS (reactive nitrogen species). Interfering with membrane lipid peroxidation. Inhibition of iNOS, COX-2, 5-LOX, prostaglandins, leukotrienes, and proinflammatory cytokines (TNF- α , IL-1, IL-2, IL-4, IL-6, γ -interferon) *in vitro* and *in vivo*.

- Immunomodulatory effects among them decreasing the immune cells infiltration into the inflamed tissue, enhancing the differentiation of the regulatory T-cells, stabilization of mast cells, and decreasing the adhesion between the leukocytes and endothelial cells *in vitro* and *in vivo*.
- Other phenotypic effects including decreased paw edema, pleural exudates, pain, and fever *in vivo*.
- On the molecular level, many inflammatory pathways have been interrupted among them the NF-κB, MAPK, JNK, WNT/β-catenin, and STAT3.

Buddleja Officinalis Maxim

Buddleja officinalis (Loganiaceae) is a Chinese medicinal plant, which has been used in the traditional medicine for the treatment of conjunctivitis, stroke, and headache. Buddlejae Flos (Mi Meng Hua) is a Chinese medicine composed of the dried flower buds of *B. officinalis*. Ateoside and linarin are among the major phytochemicals in Mi Meng Hua. They have a strong antioxidant and antiinflammatory activities which are pertained to the suppression of NO, TNF- α , and IL-1 β in LPS-induced human umbilical vein endothelial cells *in vitro* [17].

Caesalpinia Minax Hance

The seeds of the Chinese plant *Caesalpinia minax* (Fabaceae) are traditionally used for the remedy of common cold, dysentery, and rheumatoid arthritis. Its chloroform fraction is rich in cassane-type diterpenes, which remarkably reduced the proinflammatory cytokines, IL-6, IL-1 β , and TNF- α in the serum of Wistlar rats [18].

Caesalpinia Sappan L

The sappan wood or the Brazilian wood, *Caesalpinia sappan* (Fabaceae), has long been used traditionally for the remedy of inflammatory diseases. Brazilin, the major constituent of the sappan wood, has been reported for its *in vitro* analgesic and anti-inflammatory effects [19]. *C. sappan* was even more potent in lipopolysaccharide-treated bone marrow cells than other related anti-inflammatory herbs as *Machilus thunbergii*, *Agastache rugosa*, and *Saururus chinensis*. The extract of *C. sappan* reduced the production of TNF- α and significantly decreased the metabolic activity as well as the mitochondrial membrane potential of the treated bone marrow cells [20].

Calotropis Procera

Calotropis procera (Apocynaceae) is an Indian medicinal plant that has been used traditionally in wound healing, rheumatism, and in the treatment of different skin diseases. Previous phytochemical investigations revealed that the pharmacological effects of the plant are mainly related to the proteins of the plant latex. A highly homogenous cocktail of laticifer proteins containing peptidases and osmotin was prepared from the latex of *C. procera* and evaluated for its *in vivo* anti-inflammatory activity in treatment of oral mucositis (a severe inflammatory condition of ulceration and severe oral tissue damage). The protein cocktail successfully ameliorated the inflammation in oral mucosa with total reduction of myeloperoxidase activity (an enzyme present in neutrophils usually used as a quantitative marker of neutrophil infiltration in inflammatory conditions). Moreover, the laticifer protein fraction minimized the expression of adhesion molecules ICAM-1 (intercellular adhesion molecule-1) and Iba-1 (ionized calcium binding adapter molecule1), which are important inflammation biomarkers [21].

13. Centipeda Minima (L.) A.Br

The neuroinflammation induced by LPS in BV2 and microglial cells was reduced dramatically by the ethanolic extract of the Chinese herb *Centipeda minima* (Asteraceae). Neuroprotection was mediated through the inhibition of NF- κ B (comparable to dexamethasone), TNF- α , IL-1 β , COX-2, NADPH-oxidases 2 (NOX-2) and 4 (NOX-4), ROS, PGE₂, and iNOS *in vitro*. Phytochemical investigations on the ethanolic extract of *C. minima* using LC-MS/MS revealed the presence of phenolic acids, *viz*. caffeic acid, chlorogenic acid and its isomers, which contribute to the activity. *In vivo* experiment on LPS-induced neuroinflammatory mouse model showed that the extract was able to ameliorate the neuroinflammation after 3 days of treatment [22].

Cissus Gongylodes (Grapevine)

This plant belongs to family Vitaceae and its growth is restricted to the tropical rainforests of South America, Asia, and Australia. It has been used traditionally as an anti-inflammatory remedy for urolithiasis. The decoction of *Cissus gongylodes* was found to decrease the edema induced by croton oil in the ears of the tested mice *in vivo*. It inhibited both COX and LOX inflammatory pathways as it significantly decreased the concentrations of PGE₂ and LTB₄. The decoction likewise decreased TNF- α levels and interfered with the formation of calcium oxalate crystals and decreased their number and area [23].

Citrus Bergamia

Bergamot (*Citrus bergamia* Risso et Poiteau (Bergamot) is a shrub of the Rutaceae family famous for its winter fruits and has a diverse array of pharmacological and nutritional values. Bergamot essential oil fraction deprived of furocoumarins (BEO-FF) displayed potent *in vivo* anti-inflammatory effect as evidenced by the significant inhibition of carrageenan-induced paw edema after an intra-peritoneal injection of carrageenan. The strong anti-inflammatory activity was confirmed by histological and immunohistochemical analysis and was related to the downregulation of the pro-inflammatory cytokines IL-1 β , IL-6, TNF- α , and the inhibition of the release of NO, PGE₂, ROS. BEO-FF also demonstrated prominent antinociceptive activity as demonstrated by the acetic acid-induced writhing test and the hot plate model. The pronounced effects of BEO-FA were attributed to its major monoterpene content including limonene, linalool, and linalyl acetate [24].

Cyclamen Africanum B. et R

Cyclamen africanum B. et R. (Primulaceae) is the only species, among 20 species belonging to this genus, available in Algeria. This perennial plant displayed strong antioxidant and anti-inflammatory activities as revealed from its free radical scavenging potential (polyphenol content: 43.5 ± 1 mg equivalent gallic acid/g extract; flavonoid content: 2.48 ± 0.08 mg equivalent quercetin/g extract) and its ability to stabilize the membrane of red blood cells (especially at low extract concentrations and up to 200 µg/mL). The antioxidant capacity of the crude extract was evaluated by DPPH assay and its metal chelating (reducing) power [25].

Elephantopus Scaber Linn. Herb

Elephantopus scaber (Asteraceae) is an annual herb widely distributed in the old world and is used in the treatment of many inflammatory disorders as fever, sore throat, and dysentery. Its ethanolic extract was found to inhibit the transcription and translation of iNOS, hence suppressing the production of NO. The suppression extends to include some proinflammatory mediators like TNF- α , MCP-1, IL-6, and IL-1 β through the blockade of NF- κ B signaling pathway. These investigations were performed *in vitro* using Western blot, RT-PCR, EMSA (electrophoretic mobile shift assay) and *in vivo* through the analysis of the peritoneal lavage fluid of the LPS-stimulated mice using ELISA [26].

Elsholtzia Ciliata (Thunb.) Hyl

Elsholtzia ciliata (Thunb.) Hyl. (Labiatae) is a medicinal plant widely distributed throughout China, Korea, and Europe. A comparative study of the ethanol extracts of its flowers, leaves, stems, as well as the whole plant was conducted for evaluating their *in vitro* anti-inflammatory and antioxidant activities. All extracts demonstrated significant inhibition of the secretion of the pro-inflammatory cytokines IL-6, TNF- α . and PGE₂ in lipopolysaccharide-induced mouse peritoneal macrophage cells. Meanwhile, the leaf extract was the most potent blocker of PGE₂ secretion. The extracts of the flowers and stems exhibited the most significant suppression of TNF- α and IL-6. On the other hand, the stem extract showed the least antioxidant effect upon testing with different assays [27].

Epigynum Auritum

Epigynum auritum Schneid. (Apocynaceae) is a Chinese herb traditionally used as a remedy for arthritis and other inflammatory disorders. Different fractions of the hydroalcoholic extract of *E. auritum* significantly inhibited the release of TNF- α , IL-6, and NO in LPS-induced RAW264.7 macrophages while stimulating the expression of IL-10. The 100% methanolic fraction down regulated the expression of COX-2 and iNOS enzymes. *In vivo* carrageenan-induced paw edema and xylene-induced ear edema models confirmed the potent anti-inflammatory activity of those fractions through the prominent amelioration of inflammatory cytokines [28].

Eugenia Stipitate McVaugh

Genus *Eugenia* (Myrtaceae) harbors more than 1000 species, many of which show antinociceptive, antioxidant, and anti-inflammatory activities. GC-MS analysis of the essential oil obtained from *E. stipitate* leaves showed the presence of numerous terpenes, the predominant of which are guaiol (13.7%), *trans*caryophyllene (11.3%), γ -eudesmol (6.5%), 10-*epi*- γ -eudesmol (5.9%), and α eudesmol (5.9%). The oil was tested by Costa *et al.* for its acute toxicity, antinociceptive, antipyretic, and anti-inflammatory activities *in-vitro* and *in-vivo*. The results showed that:

- The essential oil of *E. stipitate* was considered safe ($LD_{50} > 2000 \text{ mg/kg}$).
- The antinociceptive activity was based on the acetic acid-induced writhing test,

which showed that the oil significantly inhibited the number of abdominal writhings compared to the control mice *in vivo*.

• The anti-inflammatory activity was assessed by the carrageenan-induced paw edema test, where the oil significantly reduced the volume of edema as well as decreasing the migration of leukocytes and neutrophils in the carrageenan-induced peritonitis model *in vivo*. Moreover, the oil provided protection for the bovine serum albumin (BSA) against denaturation *in vitro*.

The fever induced in mice by the injection of yeast (*Saccharomyces cerevisiae*) was significantly reduced by the oil within the first hour of treatment *in vivo* [29].

Eupatorium Japonicum Thunb

Eupatorium japonicum (Asteraceae) is a Chinese medicinal plant and an effective painkiller. In the rheumatoid arthritis fibroblast-like synoviocytes, *E. japonicum* ethanol extract demonstrated a dose-dependent *in vitro* anti-inflammatory effect. Suppressing the expression of IL-1 β , the activation of NF- \Box B, and the transcription of MMP-9 were the supposed mechanisms underlying its anti-inflammatory effect [30].

Forsythia Suspensa (Thunb.)

The aqueous extract of the fruits of the Asian flowering plant *Forsythia suspensa* (Oleaceae) significantly suppressed inflammation. The molecular mechanism underlying this *in vitro* activity include the inhibition of NF-κB pathway *via* the activation of A20 protein as well as the stimulation of the Nrf2 signaling cascade, which resulted in the activation of NQO1 (NAD(P)H quinone oxidoreductase 1), HO-1 (heme oxygenase-1), and glutamate-cysteine ligase catalytic subunit (GCLC) [31].

Galinsoga Parviflora Cav

Galinsoga parviflora (Asteraceae) is an annual herb used worldwide in the folk treatment of skin inflammation, wound healing, and in the protection against UV radiation. Its essential oil has an antibacterial activity specially against *S. aureus* and *B. cereus*. The extract was found to decrease IL-6 levels induced by IL-1 β (up to 33%), however it has no effect on the constitutive levels of IL-6 in the endothelial cells. It has meanwhile a moderate antioxidant effect but a strong anti-hyaluronidase activity (IC₅₀ = 0.47 mg/ml) [32].

Garcinia Cambogia Gaertn. and Pothos Scandens L.

A comparative study was performed on seven plants traditionally used in Sri Lanka for the treatment of different inflammatory conditions including Argyreia populifolia Choisy (Convolvulaceae), Garcinia cambogia Gaertn. (Clusiaceae), Hibiscus furcatus Willd (Malvaceae), Mollugo cerviana L. (Molluginaceae), Nyctanthes arbor-tristis L. Gaertn. (Oleaceae), Ophiorrhiza mungos L. (Rubiaceae), and Pothos scandens L. (Araceae). The n-hexane, dichloromethane, ethyl acetate, and methanol extracts of the seven herbs were assessed for their in vitro capability to inhibit major pro-inflammatory mediators via cell-free and cellbased assays of 5-lipoxygenase (5-LO), microsomal prostaglandin E_2 synthase-1 (mPGE₂S-1), and nitric oxide (NO) scavenging capacity. The *n*-hexane and dichloromethane extracts of Garcinia cambogia showed the most significant suppression for 5-LO in human neutrophils with IC_{50} values of 0.92 and 1.39 µg/ml, respectively. The same extracts demonstrated significant inhibition for isolated human 5-LO (IC₅₀ = 0.15 and 0.16 μ g/mL) and mPGE₂S-1 (IC₅₀ = 0.29 and 0.49 μ g/ml). The potent anti-inflammatory activities of the lipophilic extracts were attributed to the secondary metabolite garcinol, which had a dual 5-LO/mPGE₂S-1 inhibitory activity. Moreover, the lipophilic extracts of *Pothos* scandens displayed potent inhibition of mPGE₂S-1 [33].

Halosarcia Indica Willd

The aqueous extract of the tropical herb *Halosarcia indica* (Amaranthaceae) demonstrated significant *in vivo* anti-inflammatory activity evidenced by the dose dependent reduction in carrageenan-induced edema with 42% inhibition at the dose of 400 mg/kg compared to the standard indomethacin, which showed a 48% inhibition. The extract likewise reduced the pellets weight in the cotton granuloma assay relative to diclofenac sodium. The analgesic effect was evidenced by the significant reduction of writhing responses in rats at 200 mg/kg and 400 mg/kg compared to aspirin. The prominent activities of *H. indica* extract was most probably related to its phenylpropanoid content [34].

Hyusopus Cuspidatus Boriss

The essential oil of the Chinese herb *Hyusopus cuspidatus* (Labiatae) significantly decreased the inflammatory response of the cotton-ball induced granuloma in mice at a dose of 0.4 mL oil/kg, which is significant to the effect produced by aspirin. The oil reduced serum nitric oxide, prostaglandin E_2 , IL-6, IL-1 β , and TNF- α levels in serum [35].

Indigofera Argentea Burm. F

The true indigo, *Indigofera argentea* (Fabacceae), has long been used in the folk treatment of headache, vertigo, pain, and inflammation. A recent study showed that its hydroalcoholic extract exhibited *in vivo* dose dependent analgesic activity upon evaluation via the capsaicin-induced pain test and the hot plate model. The analgesic activity was also evidenced by the reduction of mice writhes and the inhibition of pain latency at concentrations of 30, 100, and 300 mg/kg. The analgesic effect of the extract was pertained to its central action on the opioid receptors and the inhibition of COX and LOX metabolites in the peripheral tissues. Dose dependent anti-inflammatory activity was exhibited by the extract in carrageenan-induced paw edema test in rats in addition to its in vitro good membrane stability with 49.29% maximum percentage hemolysis inhibition in the human red blood cell membrane stabilization anti-inflammatory method. Moreover, the extract at the doses of 100 and 300 mg/kg demonstrated significant reduction in rectal temperature in yeast-induced pyrexia test in rats. Results validated the traditional use of *I. argentea* in pain management and inflammation and attributed its activities to its content of flavonoids, phenols, alkaloids, saponins and tannins [36].

Kadsura Heteroclita

Kadsura heteroclite (Schisandraceae) has long been used in the traditional Chinese medicine for the treatment of rheumatoid arthritis. *In-vivo* investigations showed that *Kadsura heteroclite* stems decrease paw-edema induced by subcutaneous injection of carrageenan, the ear edema induced by xylene, and the abdominal writhings induced by the intraperitoneal injection of acetic acid. *In-vitro* studies showed that the extract reduce the cytokines level of TNF- α , IL-1 β , and IL-6 [37].

Kleinia Pendula (Forssk.) Herb

This herbal plant is native to the Southwestern mountains of Saudi Arabia and belongs to family Asteraceae. Its chloroform and ethyl acetate fractions, administered at doses 100, 200, and 300 mg/kg of the mice body weight, displayed significant analgesic and anti-inflammatory properties comparable to diclofenac sodium (10 mg/kg, positive control) *in vivo*. Metabolic profiling by UPLC-MS showed the richness of these fractions with phenolic acids (caffeoylquinic acid derivatives, protocatechuic, and chlorogenic acids), tannins, and flavonoidal glycosides [38].

Larrea Species and Zuccagnia Punctate Cav

María *et al.* has investigated the synergistic antifungal activity of the binary mixtures of *Larrea* (*L. divaricate* Cav., *L. cuneifolia* Cav., and *L. nitida* Cav.) (Zygophyllaceae) and *Zuccagnia* (*Z. punctate* Cav.) (Fabaceae) hydroalcoholic extracts. The antifungal activity of the most active combinations was attributed to their anti-inflammatory effect due to the inhibition of lipoxygenase enzyme with IC_{50} values ranging from 0.078 to 0.56 µg/mL, which were more potent than that of the standard quercetin (IC_{50} value at 0.9 µg/mL). The percentage of LOX inhibition ranges from 94-99%, which was comparable to naproxen (95%) [39].

Ligusticum Chuanxiong and Radix Paeoniae Lactiflora

Ligusticum chuanxiong (Apiaceae) rhizomes have long been used in the folk remedy of cardiovascular and inflammatory disorders. Its main active constituents, including ligustrazine, tetramethylpyrazine, butylidenephthalide, and phthalide lactones, inhibited the release of different pro-inflammatory mediators [40]. Paeoniflorin, the major constituent of *Paeonia lactiflora* (Paeoniaceae) roots, demonstrated prominent therapeutic effect in different *in vivo* inflammatory models. Meanwhile, the combination of *L. chuanxiong* and *P. lactiflora* roots demonstrated superior *in vivo* anti-inflammatory potential on focal cerebral ischaemic stroke. It significantly reduced the levels of IL-1β, IL-6, IL-12, and IFN-c in serum and brain tissues of rats. The anti-inflammatory and antiapoptotic potential of the combination was mediated through TLR4/MyD88/MAPK/NF- κ B signalling pathways [41].

Manilkara Zapota (Sapodilla / Chikoo)

Manilkara zapota (Sapotaceae) is a tropical evergreen tree that has long been used as an analgesic for the treatment of common cold, fever, and wounds. The chloroform and methanol extracts of *M. zapota* leaves were investigated for their *in vivo* pain-relieving effect using tail-flick and hot-plate assays. By evaluating three different doses of both extracts (100 mg/kg, 200 mg/kg, and 400 mg/kg), the highest dose (400 mg/kg) displayed the most significant analgesic effect in the two models [42].

Mitrella Kentia Leaf and Stem Extract

Mitrella kentia (Annonaceae), which is commonly distributed in the tropical areas

of Asia-pacific, is traditionally used as remedy for fever. The mechanism of its anti-inflammatory effect is due to the inhibition of the production of PGE_2 , TXB_2 , and its antagonism on PAF (platelet activation factor) receptor *in vitro* [43].

Mycetia Cauliflora Reinw.

Genus *Mycetia* (Rubiaceae) comprises ca. 30-45 species, of which *Mycetia cauliflora* is famous for the healing of pain, ulcers, and inflammation among the tribal people of Bangladesh. *In-vitro* biochemical investigations on the methanolic extract of *M. cauliflora* showed a significant reduction of iNOS, NO, IL-1 β , inhibition of the nuclear translocation of the transcription factor p65 and p50 protein, as well as the suppression of the phosphorylation - hence inactivation – of IkB kinase (IKK), IkBa, and Akt (protein kinase B). These mechanisms are proceeded *via* phosphoinositide-dependent kinase-1 (PDK1), which is thought to be the main target of *M. cauliflora* methanolic extract [44].

Niebuhria Apetala Dunn.

Niebuhria Apetala (Capparis apetala) (Brassicaceae), among other caper plants, is famous for the valuable functional phytoconstituents including vitamins, phytosterol, flavonoids, alkaloids, and tannins. The leaf and stem alcohol extracts of the plant significantly reduced the thermal-induced pain in rats in a dose dependent manner using Eddy's hot plate test. Furthermore, the leaf and stem extracts demonstrated significant increase in the tail flick latency upon testing using the heat conduction model. The maximum analgesia was observed at 500 mg/kg comparable to the standard drug diclofenac. Moreover, the leaf extract of *N. apetala* showed more analgesic activity than the stem [45].

Ocimum Forskolei Benth.

Genus *Ocimum* is one of the largest genera in family Labiatae widely distributed in Africa, Asia, and Central America. *Ocimum forskolei* has been used in the folk medicine of the Arabic peninsula for the treatment of cold symptoms like cough and rhinitis as it is rich in flavonoids and phenolic acids. Phytochemical investigations revealed that the hydroalcoholic extract significantly reduced the carrageenan-induced paw edema (with 40% inhibition) in rats even better than the standard drug indomethacin [46].

Opuntia Humifusa Raf.

The methanolic extract of the Korean plant *Opuntia humifusa* (Cactaceae) displayed promising *in vitro* anti-inflammatory effect by the inhibition of NO production and decreasing the levels of iNOS in the LPS-induced RAW264.7 macrophages [47].

Paeoniae Alba and Atractylodis Macrocephalae

This herbal combination of Radix *Paeoniae alba* (RPA) (Paeoniaceae) and Rhizoma *Atractylodis Macrocephalae* (Asteraceae) have been used in China to enhance the blood circulation, in the treatment of diarrhea and other gastrointestinal disorders. The inflammatory responses are suppressed through reduced NO production (by downregulation of iNOS) and by the inhibition of proinflammatory cytokines like TNF- α , IL-6, and MCP-1. On molecular basis, the herbal combination inhibits NF- κ B pathway by suppressing the translocation of p65 to the nucleus and prohibiting I κ B- α (inhibitor of kappa B) phosphorylation and degradation in murine macrophages *in vitro* [48].

Panax Ginseng

This Asian-Pacific plant belonging to family Araliaceae has long been used in traditional medicine for improving memory and cognitive disorders in Alzheimer's patients as well as a remedy for chronic inflammatory disorders as COPD (chronic obstructive pulmonary disease). Recently, studies showed that *Panax ginseng* could decrease the inflammatory symptoms by reducing IL-6 and TNF- α levels in cancer cachexia mice models *in vivo* due to the active constituent ginsenoside Rb1 [49].

Phyllanthus Amarus Schum. & Thonn.

Phyllanthus amarus (Euphorbiaceae) is a medicinal herb traditionally used for the treatment of various ailments including inflammatory diseases. Its pharmacological effect is due to the diversity of its secondary metabolites among them lignans, tannins, flavonoids, and alkaloids. The ethanol extract of *Phyllanthus amarus* significantly inhibited the production of the pro-inflammatory PGE₂, IL-1β, and TNF-α in LPS-induced human macrophages. Additionally, it suppressed the activation of the inflammatory cascades PI₃K/Akt, MAPKs, and NF-κB [50].

Physalis Angulata L.

The calyces of the Colombian Caribbean annual herb *Physalis angulate* (Solanaceae) displayed significant anti-inflammatory effect in DSS (dextran sulfate sodium)-induced colitis mice. The intestinal inflammation was reduced due to the inhibition of the pro-inflammatory cytokines IL-1 β , TNF- α , IL-6, IL--, iNOS, COX-2 and increasing the levels of the anti-inflammatory genes of IL-10, arginase-1, TGF- β (transforming growth factor beta), and MRC-1 (mannose receptor c) *in vitro*. Moreover, the extract improved the histological score and reduced the severity of the inflammatory symptoms in mice with DSS-induced colitis [51].

Phytolacca Dodecandra

This Ugandan herbal plant has traditionally been used for the management of pain and inflammatory disorders. The crude extract of *Phytolacca dodecandra* (Phytolaccaceae) was evaluated for its analgesic and anti-inflammatory activity using the acetic acid-induced writhing test and histamine-induced pw edema test, respectively. The results showed that the extract significantly reduced pain sensation and edema in the tested mice *in vivo* [52].

Picria Fel-Terrae Lour

This Indonesian herb belongs to genus *Picria* (Linderniaceae). Although the plant has antidiabetic, hepato- and cardioprotective effect, recent studies showed that it exerts an analgesic and anti-inflammatory activity through its immunomodulatory effect by the downregulation of TNF- α , IL-6, IL-1 β , COX-2, and iNOS in LPS-treated macrophages without inducing cytotoxicity [53].

Pituranthos Scoparius Coss. and Dur

Pituranthos scoparius (Apiaceae) is a medicinal plant well known in Algeria and North Africa for the treatment of measles, rheumatism, spasms, pain, diabetes, hepatitis, and urinary tract infections. The hydroalcoholic extract of the plant was evaluated for its *in vivo* anti-inflammatory activity using carrageenan-induced paw edema, xylene, and croton oil-induced ear edema. A dose dependant inhibition of edema was observed in mice (stimulated with xylene and croton oil) at concentrations of 100, 300, and 600 mg/kg, however in those stimulated with carrageenan the anti-edematous effect was seen at doses of 100, 250, and 500 mg/kg. Moreover, the analgesic activity of *P. scoparius* extract was investigated

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using the acetic-acid-induced writhing model. The extract exhibited potent effect with 69.92% writhing inhibition at 500 mg/kg [54].

Pterocephalus Hookeri (C.B. Clarke) Höeck

Pterocephalus hookeri (Caprifoliaceae) is a herbal plant distributed in the area of Tibet and is known among the native people as "pang-ziduo-wo". It is traditionally being used for the treatment of cold and flu symptoms as well as rheumatoid arthritis. Recent investigations showed that the plant extract is rich in bis-iridoids, which display potent analgesic and anti-inflammatory effects. The extract significantly reduced the writhing effect induced by acetic acid in mice (measure of acute pain) and decreases the ear and paw edema induced by xylene and carrageenan, respectively (measure of inflammation). These compounds primarily work by interfering with NF-κB pathway [55, 56].

Qing Re Zao Shi Liang Xue Decoction (QRZSLXF)

The Chinese decoction is composed of 11 different herbs, namely gypsum fibrosum, Rhizoma Anemarrhenae, Flos Lonicerae, Fructus Forsythiae Suspensae, Radix Scutellariae Baicalensis, Radix Salviae Miltiorrhizae, Folium Phyllostachydis Henonis, Cortex Moutan Radicis, Radix Rehmanniae, Fructus Gardeniae, and Cornu Saigae Tataricae. The formulation showed strong efficacy in the treatment of inflammatory bowel disease by decreasing the expression of interleukin-6, the transcription factor STAT3, and RAR-related orphan receptor gamma (RORyt) *in vitro* as evidenced by Western blotting and quantitative real time PCR [57].

Reineckia Carnea (Andr.) Kunth

This Chinese herb, *Reineckia carnea* (Asparagaceae) is rich in saponin glycosides, which displayed significant *in vitro* anti-inflammatory activity by inhibiting the production of NO in LPS-stimulated RAW 264.7 cells [58].

Rhodiola Crenulate (Golden Root)

Rhodiola crenulata (Crassulaceae) is a medicinal plant traditionally used in Asia and East Europe for the treatment of various epidemic diseases. The water extract of the plant demonstrated *in vitro* anti-inflammatory activity on LPS-stimulated macrophage cells. On extract treatment, the levels of the inflammatory mediators like IL-1 β , IL-6, and NO were diminished. The activity was related to the Naturally-derived Analgesics

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inhibiting capacity of the extract for the expression of NF- $_{\kappa}$ B, iNOS, and MAPK proteins in addition to its induction for autophagy related molecules [59].

Sambucus Australis Cham. & Schltdl

Elderberry (*Sambucus australis*) (Adoxaceae) is commonly used in Brazil for the treatment of various inflammatory disorders due to its richness with flavonoids, triterpenoids, volatile oil, and phenolic acids. *In-vivo* and *in-vitro* investigations on its methanolic extract revealed significant reduction of interleukins 4 and 5, interferon- γ , nitric oxide, and decreased expression of NF- κ B. On the other hand, the extract increased the production of the anti-inflammatory cytokine IL-10 [60].

Sarcopoterium Spinosum

Sarcopoterium spinosum (Rosaceae) is a Mediterranean plant, which is used in the Beduin traditional medicine for the treatment of diabetes and inflammation. Its anti-inflammatory effect is mediated by increasing the expression of the antiinflammatory genes IL-10 and Arg-1, the inhibition of the NF- κ B inflammatory pathway and Akt phosphorylation in RAW264.7 cells (as evidenced by real time PCR and Western blot analysis), and by the inhibition of NO secretion (using Griess reagent) *in vitro*. The plant was also found to decrease the expression of several adipocytokines involved in metabolic inflammation like fetuin-1, resistin, lipocalin-2, PAI-1 (plasminogen activator inhibitor-1), RBP-4 (retinol binding protein-4), and ICAM-1 (intercellular adhesion molecule-1). Reduction of inflammation by *S. spinosum* aqueous root extract is not restricted to the adipose tissue but is also observed in the liver of obese diabetic mice [61].

Scrophularia Megalantha Rech. f

Scrophularia megalantha (Scrophulariaceae) is a medicinal herb growing mainly in the city of Kelardasht in the North of Iran. It has traditionally been used for the treatment of eczema, goiter, stomach ulcers, and gall bladder infections. The 80% ethanol extract of the aerial parts reduced the infiltration of inflammatory cells to the CNS and decreased disease progression in the multiple-sclerosis mice model. *In-vitro* studies revealed that the extract reduced the levels of IL-17 and interferon- γ while increasing the levels of IL-10 [62].

Scutellaria Barbata D. Don

Scutellaria barbata (barbed skullcap) is a flowering plant commonly grown in

Asia and belongs to the mint family (Labiatae) with reported anti-cancer and antiinflammatory activities. Recent phytochemical investigations showed that the ethanol and ethyl acetate extracts of *S. barbata* are rich sources of phenolics/flavonoids and carotenoids, respectively. Biological studies *in-vitro* revealed that *S. barbata* extracts displayed dose-dependent anti-inflammatory activity by reducing NO, PGE₂, IL-6, and IL-1 β production and decreasing the expression of p-JNK and p-ERK [63].

Sigesbeckiae Species

Sigesbeckiae Herba is a Chinese herbal medicine with known anti-inflammatory activity. Linghu et al. [64] performed a comparative study on three different species viz. S. pubescens Makino (SP), S. orientalis L. (SO), and S. glabrescens Makino (SG) to investigate the underlying molecular mechanisms. Immunofluorescence staining accompanied with ELISA test showed that the three species *in-vitro* inhibited the nuclear factor κB (NF- κB) signaling pathway and mitogen-activated protein kinases (MAPKs). They likewise suppressed the inflammatory enzymes cyclooxygenase-2 (COX-2) and the inducible nitric oxide synthase (iNOS) - hence the release of NO - in the order of SP > SO > SG. They also decreased the secretion of post-inflammatory cytokines like tumor necrosis factor- α (TNF- α), interleukin-6 (IL-6), and monocyte chemoattractant protein-1 (MCP-1). In another study conducted by Zhong et al., the anti-inflammatory activity of S. glabrescens was related to in vitro reduction of NO, MCP-1, IL-6 secretion in the addition to inhibition of NF-KB activation. In-silico investigations showed that toll-like receptor-4 (TLR-4) as well as transcription factors 65 and 105 (p65 and p105) could likewise be potential anti-inflammatory targets for S. glabrescens [65].

Sinapis Semen

Sinapis semen is the dried ripe seeds of Sinapis alba L. (white mustard) or Brassica juncea (L.) Czern. (yellow mustard) (Brassicaceae). The study of the *in vivo* anti-inflammatory potential of sinapis semen in acute and chronic mouse models revealed that it suppressed the protein and mRNA expression of TNF- α , IL-1 β , and IL-6 as well as inhibiting the myeloperoxidase activity in the ears of the treated mice [66].

Sophora Flavescens Ait

Sophora flavescens (Fabaceae) is one of 52 species belonging to genus Sophora,

which is distributed in Asia and the pacific. It is used in the traditional Chinese medicine as a strong antipyretic agent due to its flavonoid and alkaloid content. The extract reduced xylene-induced ear edema (with inhibition of 41% and 29% for extract doses of 100 and 200 mg/kg, respectively, compared to 47% inhibition with indomethacin) and carrageenan-induced paw edema in mice. *In-vitro* studies on the LPS-induced RAW 264.7 cells showed that the extract significantly decreased IL-6, TNF- α , NO, and MCP-1 levels [67].

Spatholobus Suberectus

Spatholobus suberectus (Fabaceae) is a Chinese medicinal herb, which has traditional uses for the treatment of rheumatism, menstrual abnormalities, anemia as well as being used as a dietary supplement in tea and soup. DPPH (2,2-diphenyl-1-picrylhydrazyl) and nitric oxide *in-vitro* assays showed that the hot aqueous extract of *S. suberectus* has a strong free radical scavenging potential (due to its high gallic acid content which is ca. 108.4 mg/g extract), nitric oxide reducing ability (in a dose-dependent manner with no significant difference at 200 μ g/mL), and reduction of iNOS and COX-II expression in murine RAW 264.7 cells treated with LPS [68].

Swertia punicea Hemsl

Swertia punicea (Gentianaceae) is a medicinal herb traditionally used in China for the alleviation of fever, jaundice, and hepatitis. Enzyme linked immunosorbent assay (ELISA) method was adopted for investigating the *in vitro* antiinflammatory activities of its isolated compounds. All metabolites demonstrated promising anti-inflammatory activities with IC_{50} values ranging from 1.237 to 3.319 mM. Insights into the structure-activity relationships of the isolated xanthone glycosides revealed better anti-inflammatory activity of compounds with increased methoxy (OCH₃) substitution and a sugar residue [*O*-glc-(6-1)-glc] at C-1. However when the sugar part was replaced by [*O*-glc-(6-1)-xyl] at the same carbon atom, the activity was enhanced only in compounds with less number of methoxy (OCH₃) groups [69].

Tephrosia linearis (Willd.) Pers

The flavonoidal content of the tropical plant *Tephrosia linearis* (Fabaceae) interferes with the release of IL-2, GM-CSF (granulocyte-macrophage colony stimulating factor), and TNF- α *in vitro*. Some metabolites can meanwhile further inhibit the production of IL-1 β and IL-6 with superior activity than the standard

anti-inflammatory drug ibuprofen. The percentage of the cytokines release were lowered to 80% (for IL-2), 46% (for IL-6), 47% (for GM-CSF), and 12% (for TNF- α) compared to the control (LPS) [70].

Thymus zygis subsp. zygis

Thymus zygis Loefl. (Labiatae) is a famous Portuguese spice and a medicinal herb that grows in ountries around the Mediterranean Sea. *Thymus zygis subsp. zygis* is considered one of the thyme species of highest commercial value. Both the aqueous decoction and the hydroalcoholic extract of the herb demonstrated promising *in vitro* anti-inflammatory activities at their non-cytotoxic concentrations (50 µg/ml). The aqueous decoction and the hydroalcoholic extract inhibited the release of nitric oxide in lipopolysaccharide (LPS)-stimulated RAW264.7 cells by 48% and 89% respectively [71].

Isolated Natural Products with Reported Analgesic and/or Anti-Inflammatory Activities

Flavonoids and Phenolic Acids

The hydro-alcoholic extract of the leaves of the Brazilian plant *Licania rigida* (Chrysobalanaceae) is rich in flavonoids and phenolic acids like **kaempferol** (Table **2**) and **chlorogenic acid** (Table **3**). The latter is known to reduce CRP (C-reactive protein), iNOS, and COX-2. The anti-inflammatory effect is observed *via* the reduction of paw edema (induced by arachidonic acid) in mice, which is due to the inhibition of leukocytes migration, the reduction of vascular permeability, and preventing the action of vasoactive amines *in vivo* [72].

Compound (Structure / Name)		Isolated From	Mechanism	Refs.
HO CH OH OH	Kaempferol	0	 CRP (C-reactive protein), iNOS, and COX-2 reduction Inhibition of leukocytes migration Reduction of vascular permeability Preventing the action of vasoactive amines 	[72]

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Compound (structure / na	ame)	Isolated from	Mechanism	Ref.
HO OH OH O	Dihydrokaempferol	Hosta plantaginea (Liliaceae)	- COX-1 and COX-2 inhibition - Antioxidant	[73, 74]
HO CH ₃ O OH OH OH	Plantanone D			
H0H0 H0H0	Hostaflavanol A			
	Naringnin			
	Chrysoeriol	<i>Lonicera japonica</i> (Caprifoliaceae)	 NO production inhibition Inhibited production of IL, IL-1β, TNF-α, PGE₂ Reduction of iNOS, COX-2 	[75]
	Baicalein	Scutellaria baicalensis (Lamiaceae)	- interference with NO, IL-6, and TNF-α - Inhibition of COX-2 and NF- κB/p65 expression	[77]

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Compound (structure / na	ame)	Isolated from	Mechanism	Ref.
$HO_{C} = OH_{O} + O$	Dihydrofisetin	Cotinus coggygria (Anacardiaceae) Gleditsia sinensis (Fabaceae) Capsella bursa- pastoris (Brassicaceae)	 Decreased release of NO and PGE₂ Inhibition of iNOS and COX-2 expression. Inhibition of TNF-α, IL-1β, IL- 6, and MCP-1 expression Inhibition of IκB-α phosphorylation and decreased the concentration of p65. Antioxidant activity 	
	Sulphuretin	Gueldenstaedtia verna (Fabaceae)	 NO production inhibition Downregulation of IL-6, 1β, and PGE₂ 	[79]

Table 3. List of phenolic acids previously reported for their anti-inflammatory effects.

Compound (structure / Name)		Isolated from	Mechanism	Refs.
	Rosmarinic acid	Thymus atlanticus (Lamiaceae) Origanum majorana (Lamiaceae)	Inhibition of TNF- α and IL 6 and 1 β in THP-1	[80, 81]

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(Table 3) cont Compound (structure / Name)		Isolated from	Mechanism	Refs.
	Rosmarinic acid methyl ester	Salvia miltiorrhiza Bunge (Labiatae)	- COX-2 and iNOS inhibition - induction of HO-1expression	[82]
	Shimobashiric acid B	Salvia miltiorrhiza Bunge (Labiatae)		
	Chlorogenic acid	Licania rigida (Chrysobalanaceae)	- CRP (C- reactive protei n), iNOS, and COX-2 reduction - inhibition of leukocytes migration - reduction of vascular permeability - preventing the action of vasoactive amines	[72]
	Lithospermic acid	Salvia miltiorrhiza (Lamiaceae)	Inhibition of IL-1 β , IL-6, TNF- α , TLR-4, phosphorylated p65, and I κ B- α expression	[83]

The flowers of *Hosta plantaginea* (Liliaceae) have long been used in the traditional Chinese medicine for the treatment of various inflammatory disorders. **Plantanone D** is a methylated flavonoid isolated by Yang *et al.* from *H. plantaginea* flowers along with three other flavonoids, **hostaflavanol A**, **dihydrokaempferol, and naringnin** (Table 2). These compounds are responsible for the *in vitro* anti-inflammatory activity of the plant, which was due to the inhibition of COX-1 (IC₅₀ at 37.2 ± 3.2 µM) and to less extent COX-2 (IC₅₀ at 50.2 ± 3.5 µM) enzymes compared to celecoxib (IC₅₀ of 9.0 ± 0.6 µM for COX-1 and 1.0 ± 0.1 µM for COX-2). Moreover, they displayed antioxidant activities

with IC₅₀ values at $35.2 \pm 0.8 \mu$ M and $9.12 \pm 0.3 \mu$ M as measured by the DPPH and ABTS free radical scavenging assays, respectively [73, 74].

The anti-inflammatory activity of *Lonicera japonica* Thunb. (Caprifoliaceae), an ornamental plant native to East Asia – is due to its content of the flavone **chrysoeriol** (Table 2). The latter decreased ear edema in the mouse model *in vivo*, decreased NO production, inhibited IL-6, IL-1 β , TNF- α , PGE₂, lowered the levels of iNOS, COX-2, and suppressed the phosphorylation of IkB and p65, hence interfering with JAK2/STAT3 and NF-kB pathways [75].

The traditional Chinese herb, *Scutellaria baicalensis* Georgi belonging to family Labiatae has been used in the treatment of several neurodegenerative diseases [76]. One important constituent, **baicalein** (Table 2), a flavone with promising antioxidant and anti-inflammatory activities. The Antioxidant effect originates from its ability to scavenge the free radicals (ROS) while the anti-inflammatory effect is due to the interference with the production of NO, IL-6, and TNF- α as well as the inhibition of COX-2 and NF- κ B/p65 expression in the LPS-treated microglial cells *in vitro* [77].

Dihydrofisetin (Table 2) is a naturally-occurring flavanonol in many vegetables among them *Cotinus coggygria* (smoke tree) (Anacardiaceae), *Gleditsia sinensis* (Chinese honey locust) (Fabaceae), and *Capsella bursa-pastoris* (shepherd's purse) (Brassicaceae). The mechanisms of its anti-inflammatory effects are [78]:

- Decreased release of NO and PGE₂ by inhibiting the expression of iNOS and COX-2 in EAW 264.7 macrophages *in vitro*.
- Decreased *in-vitro* expression of the proinflammatory cytokines TNF- α , IL-1 β , IL-6, and MCP-1.
- Inhibition of $I\kappa B-\alpha$ phosphorylation and decreased the concentration of p65.
- Inhibition of the phosphorylation of ERK and p-38 mitogen-activated protein kinase.
- Increased the expression of the antioxidant protein, HO-1 (heme-oxidase).
- Inhibition of the carrageenan-induced paw edema in vivo.

Gueldenstaedtia verna (Fabaceae) is a perennial herb commonly distributed in Asia. Although it is traditionally used for the treatment of inflammatory ailments, recently it was found that the flavonoidal compound, **sulphuretin** (Table 2) significantly inhibited NO production in LPS-induced RAW 264.7 cells and caused downregulation of interleukins-6, 1 β , and prostaglangin E₂*in vitro* [79].

Rosmarinic acid (Table 3) is one of the major metabolites in the crude extract of the Moroccan plant *Thymus atlanticus* (Labiatae). The aqueous extract and the

polyphenols-rich fraction of *T. atlanticus* significantly reduced the levels of the chemokine MCP-1 *in vitro*. They also reduced the edema induced by carrageenan in Wistar rats *in vivo* [80].

The anti-inflammatory effect of the culinary herb *Origanum majorana* (Labiatae) is attributed to the high phenolic and flavonoid content especially **rosmarinic** acid (Table 3). Studies showed that the latter significantly inhibited the production of TNF- α and interleukins 6 and 1 β in THP-1 human macrophages *in vitro* [81].

The roots of *Salvia miltiorrhiza* Bune (Labiatae) have been traditionally used in Chinese medicine for the treatment of chronic inflammatory ailments. Hyun *et al.* evaluated the *in vitro* anti-inflammatory activities of ten isolated caffeic acid derivatives using inducible nitric oxide synthase (iNOS), nitric oxide (NO), cyclooxygenase (COX)-2 inhibition assays and by investigating the expression of heme oxygenase (HO)-1. **Rosmarinic acid methyl ester** and **shimobashiric acid B** (Table **3**) inhibited NO production at IC_{50} values of 0.6 and 1.4 μ M, respectively. Rosmarinic acid methyl ester exhibited more pronounced concentration-dependent inhibition of COX-2 and iNOS generation than that of shimobashiric acid B. Besides, it demonstrated dose-dependent induction of HO-lexpression [82].

Salvia miltiorrhiza (Labiatae) is a Chinese herb, which is used since decades in the folk medicine for the treatment of thromboangiitis obliterans (an inflammatory thrombotic disorder of the small/medium arteries and veins of the hands and feet). The plant is rich in phenolic acids among them **lithospermic acid** (Table **3**), which showed a strong immunomodulatory potential by decreasing the expression of the inflammatory proteins IL-1 β (with 45%, 56%, and 67% inhibition at 1, 5, and 25 μ M, respectively), IL-6 (with 40%, 55%, and 75% inhibition at 1, 5, and 25 μ M, respectively), TNF- α (with 19%, 35%, and 54% inhibition at 1, 5, and phosphorylated IkB- α in LPS-induced THP-1 macrophages (using qRT-PCR and Western blot) [83].

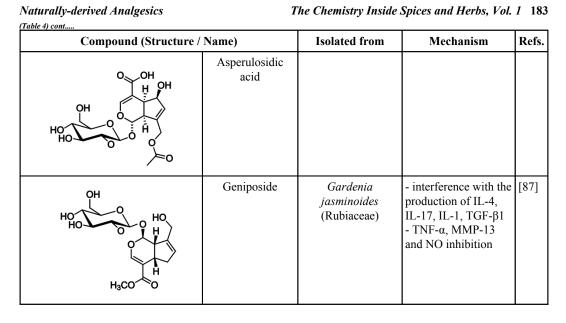
Iridoids

Yuan *et al.* attributed the anti-inflammatory activity of the Chinese medicinal plant *Cornus officinalis* (Cornaceae) to its iridodial glycosides content. Studies done on **morroniside** and **loganin** (Table 4) showed that they were effective in the treatment of ulcerative colitis *in vivo* due to their antioxidant, anti-inflammatory, and antiapoptotic activities. They maintained the integrity of the intestinal mucosa by suppressing the proinflammatory cytokines, IL-1 β , TNF- α ,

IFN- γ , and IL-6 in the mouse model. On a molecular basis, they inhibited the NF- κ B/STAT3 signaling pathway [84].

Compound (Structure /	Name)	Isolated from	Mechanism	Refs.
	Morroniside	Cornus officinalis (Cornaceae)	Suppression of IL-1β, TNF-α, IFN-γ, and IL-6 release	[84]
	Loganin			
он о , , , , он , , , , он , , , , он , , , он , , , он , , , он	Camptoside	Camptosorus sibiricus (Aspleniaceae)	Inhibition of NO production	[85]
	Asperuloside	Hedyotis diffusa (Rubiaceae)	Reduction of NO, PGE ₂ , IL-6, and TNF-α	[86]

Table 4. List of previously reported iridoids with anti-inflammatory activities.



Camptosorus sibiricus Rupr. (Aspleniaceae) is indigenous to North China and is rich in flavonoids and iridoid glycosides like **camptoside** (Table 4). The latter was found to exert significant anti-inflammatory activity, similar to curcumin, through the inhibition of NO production in RAW 264.7 cells *in vitro* [85].

Hedyotis diffusa (Rubiaceae) herb is widely distributed in many Asian countries and is famous in the folk remedy of several inflammatory diseases like nephritis, bronchitis, and arthritis. Recent studies showed that it is rich in iridoids, among them, **asperuloside** and **asperulosidic acid** (Table 4), which were found to reduce the levels of NO (*via* decreased expression of iNOS enzyme), PGE₂ (*via* decreased expression of COX-2 enzyme), IL-6, and TNF- α in the LPS-stimulated RAW 264.7 cells *in vitro*. On the molecular level, the compounds affected the NF- κ B/ MAPK signaling pathway by suppressing the phosphorylation of I κ B- α , p38, ERK, and JNK kinases [86].

Geniposide (Table 4) is an iridoid glycoside isolated from the fruits of the medicinal herb *Gardenia jasminoides* (Rubiaceae), which is traditionally used for the treatment of rheumatism and neurodegenerative disorders due to its antiinflammatory potential. In-depth biochemical studies showed that it is a potent p38-MAPK inhibitor since it interferes with the production of IL-4, IL-17, IL-1, TGF- β 1 (transforming growth factor β 1), TNF- α , MMP-13 (matrix metalloproteinase-13), and NO in the chondrocytes isolated from osteoarthritic rabbit models *in vitro* [87].

Alkaloids

The neuroprotective activity of the Chinese herb, *Corydalis tomentella* (Papaveraceae) is attributed to its anti-inflammatory isoquinoline alkaloids viz. (13*R*,14*R*)-13-hydroxy-13-methyl-8-oxosinactine and (13*S*,14*S*)-tomentelline E (Table 5). The neuroprotective effects exerted by 10 and 11 against the LPS-induced BV2 microglial cells were mediated by decreasing the levels of TNF- α , IL-6, and IL1 β in cultured cells *in vitro* [88].

Table 5. List of previously reported alkaloids with anti-inflammatory activities.

Compound (Str	ucture / Name)	Isolated from	Mechanism	Refs.
	(13 <i>R</i> ,14 <i>R</i>)-13-Hydroxy-13-met- yl-8-oxosinactine	Corydalis tomentella (Papaveraceae)	Reduction of TNF-α, IL-6, and IL1β levels	[88]
	(13 <i>S</i> ,14 <i>S</i>)-Tomentelline E			
	Dauricine	<i>Menispermum</i> species (Menispermaceae)	- Suppression of TNF-α, IL- 1β, IL-6 secretion - Interference with the activity of COX-2 and iNOS enzymes	[89]
	Berberine	Berberis vulgaris (Berberidaceae)	- Downregulation of IL-1β, IL-6, and TGF-β - Reduction of IL-17 and BAFF production	[90]
он о , , , , , , , , , , , , , , , , , , ,	Narciclasine	<i>Lycoris radiata</i> (Amaryllidaceae)	Inhibition of TNF-α, IL-1β, IL-6, COX-2, iNOS, and NF- κB	[91]

Dauricine (Table **5**) is an isoquinoline alkaloid isolated from *Menispermum* species (Menispermaceae) distributed in Asia and North America. Although the compound is famous for its strong cytotoxicity by inducing cell cycle arrest and apoptosis, recently group of scientists in the city of Wuhan discovered its significant anti-inflammatory effect in acute lung injury. The alkaloid works by inhibiting the inflammatory NF- κ B cascade as well as suppressing the secretion of the proinflammatory cytokines TNF- α , IL-1 β , IL-6, interfering with the activity of COX-2 and iNOS enzymes in a dose-dependent manner *in vitro* [89].

Berberine (Table 5) is a benzylisoquinoline alkaloid present in many medicinal plants among them *Berberis vulgaris* (barberry), *Berberis aristata* (tree turmeric), *Rhizoma coptidis* (huang lian), *Mahonia aquifolium* (Oregon grape), *Cortex phellodendri* (huang bai), *Hydrastis canadensis* (goldenseal), and *Phellodendron amurense* (Amur cork tree). Although it is known for its antimicrobial, anti-diabetic, and antitumor activity, recent *in-vitro* studies showed that it displays good anti-inflammatory effect. It is used in the treatment of chronic gastritis induced by *Helicobacter pylori* through the up-regulation of IL-10 and down-regulation of IL-1 β , IL-6, and TGF- β in the gastric mucosa. Berberine decreases the production and the expression of IL-17 (up to 50%) and BAFF (B-cell activating factor cytokine which belongs to the TNF family), therefore attenuating Th-17 (T-helper 17 cell) response [90].

Lycoris radiata (Amaryllidaceae) is a Chinese herb commonly used in the folk medicine for the treatment of Alzheimer's disease. Recent studies by Shen *et al* [91] revealed that the plant has an anti-inflammatory activity which is attributed to its alkaloid **Narciclasine** (Table 5). It inhibits the inflammatory cytokines TNF- α , IL-1 β , IL-6, enzymes as COX-2, iNOS, and signaling pathways as NF- κ B in human RAW 264.7 macrophages *in vitro*.

Saponins

The rhizome of *Anemarrhena asphodeloides* (Asparagaceae) is native to China, Japan, and Korea. It has traditionally been used in China for the treatment of fever, allergies, and Alzheimer's disease. Recent studies performed on LPS-induced RAW 264.7 cells revealed significant *in vitro* anti-inflammatory activity of the aqueous extract of *A. asphodeloides*, which is attributed to its steroidal saponins **timosaponin B** and **timosaponin B-II** (Table 6). They inhibited the expression of the proinflammatory cytokines genes (as revealed by real-time PCR) like IL-6, IL-1 β , TNF- α , COX-2, interfered with the secretions of NO and ROS, and inhibited the phosphorylation of p38 (class of MAPKs), JNK (c-Jun-N-terminal kinases), IKB α , and the p65 subunit of NF- κ B, with timosaponin B-II being more potent than timosaponin B [92].

Compound (Structure / Name)		Isolated from	Mechanism	Refs.
HO OH HO OH HO OH	Timosaponin B	Anemarrhena asphodeloides (Asparagaceae)	- Inhibition of IL-6, IL- 1β, TNF-α, COX-2 production - Interference with NO and ROS formation	[92]
HO OH HO OH HO OH	Timosaponin B-II			

Table 6. List of previously reported saponins with anti-inflammatory activities.

5. TERPENOIDS

Citral (Table 7) is a common ingredient in *Cymbopogon citratus* (lemongrass) (Poaceae) and it is a tautomeric mixture of the *cis* and *trans* isomers. Phytochemical investigations by Gonçalves *et al* revealed the molecular mechanisms of the analgesic and anti-inflammatory effects of citral. Beside its anti-oxidant activity and its NF-κB and COX-2 inhibition, the compound blocks the transient receptor potential channels of the vanilloid type 1-3 (TRPV1-3) and of the melastatin type-8 (TRPM8) in a sustained form *in vitro*. It reduces paw edema and thermal allodynia induced by carrageenan, LPS, and zymosan upon oral treatment of mice with a dose of 50-300 mg/kg. The analgesic and immunomodulatory activity of citral is dependent on toll-like receptor-4 (TLR-4) (in LPS-induced inflammation) and TLR2/dectin-2 (in zymosan-induced inflammation) modulation [93].

Naturally-derived Analgesics

Compound (Stru	cture / Name)	Isolated from	Mechanism	Refs.
Contraction of the second seco	Citral	Cymbopogon citratus (Poaceae)	- Antioxidant activity - NF-κB and COX-2 inhibition - Blockade of TRPV1-3 and TRPM8	[93]
	Malloconspur B	<i>Mallotus</i> <i>conspurcatus</i> (Euphorbiaceae)	- Inhibition of prostaglandin E ₂ , NO - COX-2, TNF-α, and NF-κB/p65	[94]
OH H H H H OH OH	Oridonin	<i>Rabdosia rubescens</i> (Labiatae)	- Suppression of IL-6 and TNF-α - Inhibit the assembly of NLRP3	[95]
C C C C C C C C C C C C C C C C C C C	9-hydroxyisoegomaketone	Perilla frutescens (Lamiaceae)	Suppression of nitric oxide, IL-6 and TNF-α production	[96]
HO" HO" HO	3-dehydroandrographolide	Andrographis paniculata (Acanthaceae)	- Attenuation of TNF-α and IL-6 release - Inhibition of NF-κB and protein kinase B	[97]
	Lecocarpinolide	Sigesbeckiae Herba (Asteraceae)	- Inhibition of NO, TNF- α , PGE ₂ , IL-6, MCP-1, and ROS release - Suppression of the expression of iNOS and COX-2	[98]

Table 7. List of previously reported terpenoids with anti-inflammatory activities.

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Compound (Strue	cture / Name)	Isolated from	Mechanism	Refs.
	Salviplenoid A	Salvia plebeia (Lamiaceae)	- Inhibition of proinflammatory mediators induced by the MAPK pathway - Inhibition of NF, iNOS and COX-2	[99]
HO HO HO HO HO HO HO	leojaponin E	<i>Leonurus</i> <i>japonicus</i> (Lamiaceae)	Inhibition of PGE ₂	[100]

Terpenoids like **malloconspur B** (Table 7) obtained from the Chinese plant *Mallotus conspurcatus* (Euphorbiaceae) were found to exert an anti-inflammatory action through the *in vitro* inhibition of prostaglandin E₂, NO (through interference with iNOS; $IC_{50} = 10.4 \pm 0.3 \mu M$), COX-2, TNF- α , and NF- κ B/p65 in RAW 264.7 macrophages. The cell viability was maintained up to 90% along its different concentrations (from 0 to 50 μ M) [94].

Rabdosia rubescens (Labiatae) is an OTC herbal medicine commonly used in China for the treatment of several inflammatory disorders. One of its major constituents is the diterpene compound, **oridonin** (Table 7). The latter inhibits the *in-vitro* activation of NF- κ B and MAPK so the release of IL-6 and TNF- α are suppressed. It was also found to inhibit the assembly of NLRP3 (NOD-, LRR- and pyrin domain-containing protein 3) inflammasome by covalently binding NLRP3 at cys-279, therefore preventing the activation of caspase-1 and decreasing the release of IL-1 β [95].

Perilla frutescens (Labiatae) commonly known as the Korean perilla is an annual edible herb distributed in Southeast Asia including South China, Japan, Korea, and India. Hundreds of constituents belonging to different classes of phytochemicals (*i.e.* terpenes, flavonoids, phytosterols, fatty acids) have been isolated from its seeds, stems, and leaves. They contribute to its nutritional and medicinal uses as anti-inflammatory, antioxidant, and as an antidote for fish and crab allergy. Detailed phytochemical study of the *in vitro* anti-inflammatory effect

of its metabolites revealed the monoterpene, **9-hydroxyisoegomaketone** (Table 7) to exhibit a remarkable suppressing effect on the production of nitric oxide (IC₅₀ value of 6 μ M) and the inflammatory cytokines IL-6 (IC₅₀ value of 5.7 μ M) and TNF- α (IC₅₀ value of 20.5 μ M) [96].

Andrographis paniculata (Acanthaceae) is a bitter herb commonly cultivated in South Asia especially in China, India, and Srilanka. The whole herb (in particular its leaves and roots) has long been used in the traditional Chinese medicine as a remedy for the constipation, sore throat, fever, and the respiratory disorders associated with common cold. One potent anti-inflammatory agent is 3dehydroandrographolide (Table 7), which attenuates the *in vitro* release of proinflammatory cytokines like TNF- α and IL-6. This is also supported by the inhibition of NF- κ B (by preventing the nuclear translocation of its p65 subunit) and protein kinase B (through prevention of its phosphorylation at the Ser-473 position) in LPS-induced RAW 264.7 macrophages, which is closely related to the activation of the expression of α 7 nicotinic acetylcholine receptors (α 7nAchR) that showed a potential role in reducing the inflammatory signs in patients with stroke, Alzheimer, and myocardial infarction [97].

The Chinese medicine *Sigesbeckiae Herba* has long been used for the treatment of rheumatoid arthritis. Its anti-inflammatory activity is attributed to the sesquiterpene lactone, **lecocarpinolide** (Table 7), which significantly inhibits the production of NO, TNF- α , PGE₂, IL-6, MCP-1, and ROS and suppresses the expression of the inflammatory enzymes iNOS and COX-2 in LPS-induced RAW264.7 *in vitro*. On the molecular basis, it interfered with NF- κ B signaling pathway through blocking the activation of p65. Its antioxidant effect is mediated through increasing the expression of nuclear factor erythroid 2-related factor-2 (Nrf2) protein and heme oxygenase-1 (HO-1) enzyme. The *in vitro* investigations were carried out using ELISA, flow cytometric analysis, immunofluorescence microscopy, and Western blotting [98].

Salvia plebeia (Labiatae) is an annual herb growing in wide areas of Asia. It is used as a folk remedy for hepatitis, cough, and diarrhea in China. The antiinflammatory effects of the ethanolic extract of the aerial parts of *S. plebeian* are pertained to a group of eudesmane-type sesquiterpenes, among them **salviplenoid A** (Table 7), which significantly reduced the release of the proinflammatory cytokine TNF- α at an IC₅₀ value of 8.0 μ M. Moreover, it decreased the expressions of iNOS and COX-2 proteins, suppressed the activity of NF- κ B (IC₅₀ = 5.0 μ M) through preventing its p50/p65 nuclear translocation and I κ B phosphorylation, and inhibiting the proinflammatory mediators induced by the MAPK (mitogen-activated protein kinase) pathway by preventing the phosphorylation of Erk 1/2 (extracellular signal-regulated kinases 1/2) [99].

The Chinese motherwort *Leonurus japonicus* (Labiatae) is distributed in many Asian countries and it has traditionally been used for improving blood circulation and treatment of edema. The latter activity was attributed to the existence of diterpenoids of the labdane type viz. **leojaponin E** (Table 7), which inhibit the *in vitro* production of prostaglandin E_2 in LPS-induced RAW 264.7 cells [100].

Polysaccharides

Wang *et al.* reported in 2018 the isolation of the anti-diabetic polysaccharide macromolecule **GPP** (molecular weight of 4.070×10^4 Da) from *Gynostemma pentaphyllum* (Cucurbitaceae) herb. It consists of rhamnose, arabinose, galactose, glucose, xylose, mannose, galacturonic acid, and glucuronic acid subunits [101]. One year later, they reported on the anti-inflammatory activity of GPP, which was due to its antioxidant and its radical scavenging activity. *In-vitro* tests showed that the compound reduced the levels of TNF- α and IL-6 while increasing the levels of IL-4 and IL-10 [102].

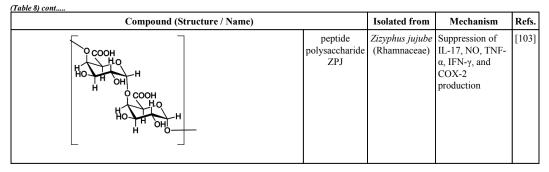
Zizyphus jujube (Rhamnaceae) is commonly known as the Chinese date or the jujube. Its fruits have numerous culinary and medicinal applications in the traditional Chinese medicine. The plant is rich in polysaccharides among them **ZPJ** (Table **8**), which is a peptide polysaccharide composed of glucuronic acid units linked at 1,4-position. ZPJ suppresses the production of IL-17 (at dose of 50 μ g/ml), NO (in a dose-dependent manner up to 300 μ g/ml), TNF- α (at dose of 50 μ g/ml), IFN- γ (at dose of 100 μ g/ml), and COX-2. On the molecular level, it inhibits MAPK signaling pathway through the preventing the phosphorylation of p38 and JNK [103].

Table 8. Reported polysaccharides with anti-inflammatory activities.

Compound (Structure / Name)		Isolated from	Mechanism	Refs.
Rhamnose-Arabinose-Galactose-Glucose-Xylose-Mannose-Galacturonic acid-Glucuronic acid (in the ratio of 4.11: 7.34: 13.31: 20.99: 1.07: 0.91: 4.75: 0.36)	GPP	pentaphyllum (Cucurbitaceae)	 Antioxidant Improve the anti-inflammatory cytokines IL-4 and IL-10 Reducing the levels of TNF-α and IL-6 	[102]



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7. MISCELLANEOUS COMPOUNDS BELONGING TO OTHER PHYTOCHEMICAL CLASSES

Atractylodes chinensis Koidzumi (Asteraceae) is one of two Atractylodes species (the other is A. lancea De Candolle) present in the Japanese crude drug, Sojutsu. The rhizomes of Atractylodes species have been reported to possess antiinflammatory effects due to the inhibition of iNOS activity, NO, and prostaglandin E_2 production. Atractylodin (Table 9) isolated from the ethyl acetate fraction of the rhizomes of A. chinensis was the most potent constituent, which significantly decreased the expression of iNOS in the IL-1 β -treated rat hepatocytes *in vivo* [104].

Compound (Structure / Name)		Isolated from	Mechanism	Refs.
Comments and the second	Atractylodin	Atractylodes chinensis (Asteraceae)	 Inhibition of NO and PGE₂ production reduction reduction of iNOS 	[104]
$HO_{HO} \rightarrow HO_{HO} 3f 3a:6f 2a-[(3 <i>R</i> , 4 <i>R</i>)- 4',6-dihydroxy-3', 5,7-trimethox- -4,10-cycloligna-1-en-2a,3adicarbonyl]-6g-(<i>p</i> -courmaroyl) sucrose	Corispermum mongolicum (Amaranthaceae)	Suppression of NO, L-6 and TNF-α production	[105]	

Table 9. List of miscellaneous natural products with anti-inflammatory activities.

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(Table 9) cont Compour	nd (Structure / Name)	Isolated from	Mechanism	Refs.
	(S)-Asperpyrone A	Chaetomium nigricolor (Chaetomiaceae)	- Inhibition of NO production - Suppression of NF-κB and JNK activation	[106]
он он	1-(4"-Methoxyphenyl)-7-(2',4'-di-hydroxyphenyl)-(<i>E</i>)- hept-2-ene	Pleuranthodium racemigerum (Zingiberaceae)	Suppression of NO and TNF-α production	[107]
	Fraxinellone	Dictamnus dasycarpus (Rutaceae)	- Inhibition of NO production - Reduction of IL-6, TNF-α, iNOS, COX-2, and NF-κB expression	[108]
HO X*	Columbianetin	Angelicae pubescentis (Apiaceae)	Reduction of TNF-α, IL-6, IL-1β, MCP-1	[109]
v v v v v v v v v v v v v v v v v v v	Dihydronortanshinone	Salvia miltiorrhiza (Lamiaceae)	Suppression of NO, TNF-α, IL- 6, ROS	[110]
	Shikonin	Lithospermum erythrorhizon (Boraginaceae)	- Reduction of TNF-α, IL-1β and IL-6 - Interference with NF-κB signaling cascade	[111]
	Physcion 8- <i>O-β</i> -glucopyranoside	Polygonum cuspidatum (Polygonaceae)	Suppression of TNF-α, IL-1β, IL-6, IL-8, IL-12, and IL-17A expression	[112]
	3',5',6'-Trimethoxy[1,1'-biphenyl]-2,2',3,4'-tetraol	Crataegus dahurica (Rosaceae)	Inhibition of NO release	[113]

The Aryldihydronaphthalene-type lignan, **3f 3a:6f 2a-[(3***R*, 4*R*)- 4',6-dihydrox--3',5,7-trimethoxy-4,10-cycloligna-1-en-2a,3a dicarbonyl]-6g-(*p*-courmaroyl) sucrose (Table 9) isolated from the ethanol extract of the Chinese plant *Corispermum mongolicum* (Amaranthaceae) displayed significant *in vitro* antiinflammatory activity by the suppression of nitric oxide production and other proinflammatory mediators as IL-6 and TNF- α in LPS-stimulated RAW 264.7 cells without affecting the cell viability even at concentration of 100 μ M. Significant inhibition of NO and TNF- α was observed at concentrations of 10, 30, and 100 μ M in a dose-dependent manner [105].

From the potato dextrose broth of *Chaetomium nigricolor* (Chaetomiaceae) fungal extract, a quinoid metabolite, **(S)-asperpyrone** A (Table 9) was isolated. The compound inhibited NO production in LPS-induced RAW 264.7 macrophages *in vitro* by suppressing the activation of NF-κB and JNK (c-Jun-N-terminal kinases) [106]

Family Zingiberaceae is famous for its anti-inflammatory perennial herbs like *Curcuma longa* and *Ginger officinalis*. Raju *et al.* investigated the anti-inflammatory potential of the rhizomes of *Pleuranthodium racemigerum* growing in Australia. The results showed that the extract is rich in diarylheptanoids, as in **1-(4"-Methoxyphenyl)-7-(3',4'-di-hydroxyphenyl)-(E)-hept-2-ene** (Table 9), which significantly decreased NO (IC₅₀ = $25 \pm 2 \mu$ M) and TNF- α (IC₅₀ = $16 \pm 9 \mu$ M) production in RAW 264.7 macrophages and N-11 microglial cells *in vitro* in comparison to curcumin having IC₅₀ values of $12.3 \pm 1.1 \mu$ M (for NO) and $11.2 \pm 2.2 \mu$ M (for TNF- α) [107].

Cortex dictamni (Dictamnus dasycarpu) is a perennial herb which belongs to family Rutaceae. The latter - together with family Meliaceae - are rich sources of oxygenated tetranorterpenoids known as limonoids. The root bark of *C. dictamni* has been used to treat rheumatic pain and other inflammatory diseases. **Fraxinellone** (Table 9) is one member of the limonoid family present in *C. dictamni* showing anti-inflammatory effects. The compound, among others, inhibited NO (nitrile relative concentration = $56.5 \pm 3.5\%$ expressed as a percentage of the control group with LPS) production in LPS-treated RAW 264.7 cells and reduced the expression levels of IL-6, TNF- α , iNOS, COX-2, and NF- κ B *in vitro* [108].

Columbianetin (Table 9) is a furanocoumarin and one of the major constituents of the Chinese medicinal plant *Angelicae pubescentis* (Apiaceae). It has been used since long as a prescription remedy for arthritis and asthma in China. *In-vitro* studies showed that the compound reduced the levels of TNF- α , IL-6, IL-1 β ,

MCP-1 in a dose-dependent manner and downregulated the NOD-1 (nucleotidebinding and oligomerization domain-1) / NF-κB pathway [109].

Dihydronortanshinone (Table 9) is a diketone natural product isolated from *Salvia miltiorrhiza* Bunge (Labiatae), which is commonly used in China and Japan for the treatment of cardiovascular and circulatory problems. The compound has a strong potential to suppress the inflammatory responses by reducing the levels of iNOS (hence NO), TNF- α , IL-6, ROS (reactive oxygen species) in murine macrophage model *in vitro*. The compound suppressed NF- κ B/p65 translocation and inhibited MAPK/p38 and JNK-1/2 activation induced by LPS [110].

The gromwell (Boraginaceae) is a Chinese herb used to alleviate several inflammatory disorders. In a recent study by Wang *et al.*, they described the main bioactive metabolite of *Lithospermum erythrorhizon* as **shikonin** (Table 9), which is a naphthoquinone with significant anti-inflammatory potential as demonstrated from its ability to reduce the proinflammatory cytokines TNF- α and IL-1 β and 6 *in vitro*. It decreases the migration of neutrophils to the mammary glands in the LPS-induced mastitis mice *in vivo* and mechanistically, it interferes with the NF- κ B signaling cascade [111].

Physcion 8-*O*-**β**-glucopyranoside (POGD) (Table 9) is an anti-proliferative anthraquinone glycoside isolated from the roots of *Polygonum cuspidatum* (Polygonaceae). Its anti-inflammatory activity was evaluated using *in vivo* type II collagen-induced arthritis rat model. POGD reduced the levels of the pro-inflammatory cytokines TNF-α, IL-1β, IL-6, IL-8, IL-12, and IL-17A causing marked reduction in the mRNA expression levels of VEGF (vascular endothelial growth factor), MMP-9 (matrix metalloproteinase-9), MMP-3, MMP-2, and COX-2 through the inhibition of NF-κB, MAPK, and TGF-β (transforming growth factor-β) signaling pathways [112].

Crataegus is a large genus belonging to family Rosaceae. It is a rich source of phytochemicals including mono, sesqui, and triterpenoids as well as flavonoids and phenylpropanoids. Recently, it has been reported that the petroleum ether fraction of *C. dahurica* fruits showed anti-inflammatory activity, which was attributed to its biphenyl constituents. The most active of them is **3',5',6'-trimethoxy[1,1'-biphenyl]-2,2',3,4'-tetraol** (Table **9**), which inhibited nitric oxide production in RAW264.7 cells *in vitro* without showing cytotoxicity. It was isolated from the petroleum ether fraction having an IC₅₀ value of 46.1 μ M, which was even stronger than indomethacin (IC₅₀ 53.2 μ M), which was the positive control in this study [113].

CONCLUDING REMARKS

Inflammation is a pathological condition involved in several disorders like cancer, Alzheimer's, autoimmune diseases (rheumatoid and multiple sclerosis), ulcers, pneumonia, gall bladder infections, diabetes, cardiovascular diseases, and eczema. Natural products play a key role in combating inflammation due to the presence of a diverse pool of anti-inflammatory secondary metabolites, including flavonoids, iridoids, glycosides, phenolic acids, polysaccharides, quinones, and alkaloids. These compounds act by decreasing the formation of pro-inflammatory cytokines like interleukins (1 β , 2, 4, 5, 6, 12, 17), TNF- α , CRP, IFN- γ , prostaglandins (PGE₂)), thromboxanes (TXB₂), leukotrienes (LTB₄), NO, GM-CSF and by increasing the release of anti-inflammatory cytokines as IL-10, Arg-1, TGF-β, MRC-1. Some of these secondary metabolites suppress the inflammatory proteins like fetuin-1, resistin, lipocalin-2, RBP-4, ICAM-1 and reduce the levels of inflammatory enzymes as COX-2, LOX, iNOS, HO-1, MMP, NOX-2/4. The majority of these compounds exhibit antioxidant activity by scavenging and reducing the concentration of free radicals as peroxides and superoxides. Moreover, they significantly decrease edema, vascular permeability, leukocytes migration, neutrophils infiltration, and vasoactive amines. On the molecular level, several inflammatory signalling pathways were the common target of these natural products, including NF-κB, MAPKs, JAK/STAT, WNT/β-catenin, and PI₂K/AkT.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The author declares no conflict of interest, financial or otherwise.

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GLOSSARY

Akt	Protein kinase B
PAF	Platelet activation factor
Arg-1	Arginase
COX	Cyclooxygenase
NF-кВ	Nuclear factor kappa light chain enhancer of activated B cells
DSS	Dextran sulphate sodium
Iba-1	Ionized calcium-binding adapter molecule

NO	Nitric oxide
p65	Transcription factor p65
ICAM-1	Intercellular adhesion molecule-1
PAI-1	Plasminogen activator inhibitor-1
IFN-γ	γ-interferon
PGE2	prostaglandin E2
IL	Interleukin
PI3K	Phosphatidylinositol 3-kinase
ІкВ	Inhibitor of kB
RBP-4	Retinol binding protein-4
iNOS	Inducible nitric oxide synthase
ROS	Reactive oxygen species
JAK-STAT	Janus kinases-signal transducer and activator of transcription
TGF-β	Transforming growth factor beta
TNF	tumor necrosis factor
LOX	Lipoxygenase
TXB2	Thromboxane B2
LPS	Lipopolysaccharide
UPLC-MS	Ultraperformance liquid chromatography-mass
VEGF	Vascular endothelial growth factor
LTB4	Leukotriene B4
MAPK	Mitogen activated protein kinase
MCP-1	Monocyte chemoattractant protein -1
MMP	Matrix metallopeptidases
mRNA	messanger Ribonucleic acid

REFERENCES

- Ahmed AU. An overview of inflammation: mechanism and consequences. Frontiers in Biology 2011; 6(4): 274.
 [http://dx.doi.org/10.1007/s11515-011-1123-9]
- Yuan G, Wahlqvist ML, He G, Yang M, Li D. Natural products and anti-inflammatory activity. Asia Pac J Clin Nutr 2006; 15(2): 143-52.
 [PMID: 16672197]
- [3] Chen L, Deng H, Cui H, *et al.* Inflammatory responses and inflammation-associated diseases in organs. Oncotarget 2017; 9(6): 7204-18.
 [http://dx.doi.org/10.18632/oncotarget.23208] [PMID: 29467962]
- [4] Porreca F, Ossipov MH. Nausea and vomiting side effects with opioid analgesics during treatment of chronic pain: mechanisms, implications, and management options. Pain Med 2009; 10(4): 654-62. [http://dx.doi.org/10.1111/j.1526-4637.2009.00583.x] [PMID: 19302436]

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- [5] Won A, Lapane KL, Vallow S, Schein J, Morris JN, Lipsitz LA. Long-term effects of analgesics in a population of elderly nursing home residents with persistent nonmalignant pain. J Gerontol A Biol Sci Med Sci 2006; 61(2): 165-9. [http://dx.doi.org/10.1093/gerona/61.2.165] [PMID: 16510860]
- [6] Tan D, Jiang C, Tao Y. Chemical constituents of *Acanthus ilicifolius*. Chem Nat Compd 2016; 52(5): 951-2.
 [http://dx.doi.org/10.1007/s10600-016-1830-1]
- [7] Huang M-Y, Zhong L-J, Wang F, Liu Q-Y, Zhang Y-H. Chemical constituents from the roots of
- *Acanthus ilicifolius* var. xiamenensis. Biochem Syst Ecol 2014; 55: 145-7. [http://dx.doi.org/10.1016/j.bse.2014.03.006]
- [8] Faysal MAR, Barek MA, Jahan S, Shahid-Ud-Daula A, Basher MA, Hasanuzzaman M. Petroleum Ether and Chloroform Soluble Fractions of Whole Plant Extract of *Acanthus ilicifolis* Linn. Possesses Potential Analgesic and Antioxidant Activities. Discovery Phytomedicine 2020; 7(1): 12-8. [http://dx.doi.org/10.15562/phytomedicine.2020.113]
- [9] Toiu A, Mocan A, Vlase L, et al. Phytochemical composition, antioxidant, antimicrobial and in vivo anti-inflammatory activity of traditionally used Romanian Ajuga laxmannii (Murray) Benth. ("Nobleman's Beard" - Barba Împăratului). Front Pharmacol 2018; 9: 7. [http://dx.doi.org/10.3389/fphar.2018.00007] [PMID: 29551972]
- [10] Junior OC, Lima NM, Silva MG, Aguiar VB, Carli GP, Scherrer EC, et al. In vitro and in vivo evaluation of anti-inflammatory activity and free radical scavenging potential of leaves extract from Anadenanthera colubrina. Nat Prod Res 2020; •••: 1-5. [PMID: 32064928]
- [11] Mayouf N, Charef N, Saoudi S, Baghiani A, Khennouf S, Arrar L. Antioxidant and anti-inflammatory effect of *Asphodelus microcarpus* methanolic extracts. J Ethnopharmacol 2019; 239: 111914. [http://dx.doi.org/10.1016/j.jep.2019.111914] [PMID: 31029760]
- [12] Han XZ, Ma R, Chen Q, et al. Anti-inflammatory action of Athyrium multidentatum extract suppresses the LPS-induced TLR4 signaling pathway. J Ethnopharmacol 2018; 217: 220-7. [http://dx.doi.org/10.1016/j.jep.2018.02.031] [PMID: 29476961]
- [13] Shim S-Y, Kim J-H, Kho K-H, Lee M. Anti-inflammatory and anti-oxidative activities of lemon myrtle (*Backhousia citriodora*) leaf extract. Toxicol Rep 2020; 7: 277-81. [http://dx.doi.org/10.1016/j.toxrep.2020.01.018] [PMID: 32071880]
- Pawłowska KA, Hałasa R, Dudek MK, Majdan M, Jankowska K, Granica S. Antibacterial and antiinflammatory activity of bistort (*Bistorta officinalis*) aqueous extract and its major components. Justification of the usage of the medicinal plant material as a traditional topical agent. J Ethnopharmacol 2020; 260: 113077. [http://dx.doi.org/10.1016/j.jep.2020.113077] [PMID: 32531411]
- [15] Moussaieff A, Mechoulam R. Boswellia resin: from religious ceremonies to medical uses; a review of *in-vitro*, *in-vivo* and clinical trials. J Pharm Pharmacol 2009; 61(10): 1281-93.
 [http://dx.doi.org/10.1211/jpp/61.10.0003] [PMID: 19814859]
- [16] Efferth T, Oesch F. Anti-inflammatory and anti-cancer activities of frankincense: Targets, treatments and toxicities. Semin Cancer Biol 2020; S1044-579X(20)30034-1. [http://dx.doi.org/10.1016/j.semcancer.2020.01.015] [PMID: 32027979]
- Xie G, Yang J, Wei X, Xu Q, Qin M. Separation of acteoside and linarin from *Buddlejae* Flos by high-speed countercurrent chromatography and their anti-inflammatory activities. J Sep Sci 2020; 43(8): 1450-7.
 [http://dx.doi.org/10.1002/jssc.201901062] [PMID: 32031325]
- [18] Tong Z, Cheng L, Song J, *et al.* Therapeutic effects of *Caesalpinia minax* Hance on complete Freund's adjuvant (CFA)-induced arthritis and the anti-inflammatory activity of cassane diterpenes as main

active components. J Ethnopharmacol 2018; 226: 90-6. [http://dx.doi.org/10.1016/j.jep.2018.08.011] [PMID: 30114517]

- [19] Nirmal NP, Rajput MS, Prasad RG, Ahmad M. Brazilin from *Caesalpinia sappan* heartwood and its pharmacological activities: A review. Asian Pac J Trop Med 2015; 8(6): 421-30. [http://dx.doi.org/10.1016/j.apjtm.2015.05.014] [PMID: 26194825]
- [20] Ahn S-Y, Joo H-G. Anti-inflammatory activity of *Caesalpinia sappan* extract on lipopolysaccharidetreated bone marrow cells. Journal of Preventive Veterinary Medicine 2018; 42(3): 107-11. [http://dx.doi.org/10.13041/jpvm.2018.42.3.107]
- [21] Ramos MV, Freitas APF, Leitão RF, Costa DV, Cerqueira GS, Martins DS, *et al.* Anti-inflammatory latex proteins of the medicinal plant Calotropis procera: a promising alternative for oral mucositis treatment Inflammation Research. Official Journal of the European Histamine Research Society 2020. [*et al*]
- [22] Li SY, Zhou YL, He DH, Liu W, Fan XZ, Wang Q, et al. Centipeda minima extract exerts antineuroinflammatory effects via the inhibition of NF-κB signaling pathway. Phytomedicine: international journal of phytotherapy and phytopharmacology 2020; 67: 153164.
- [23] Salem PPO, Vieira NB, Garcia DA, et al. Anti-urolithiatic and anti-inflammatory activities through a different mechanism of actions of Cissus gongylodes corroborated its ethnopharmacological historic. J Ethnopharmacol 2020; 253: 112655. [http://dx.doi.org/10.1016/j.jep.2020.112655] [PMID: 32045681]
- [24] Lombardo GE, Cirmi S, Musumeci L, *et al.* Mechanisms underlying the anti-inflammatory activity of bergamot essential oil and its antinociceptive Effects. Plants 2020; 9(6): 704. [http://dx.doi.org/10.3390/plants9060704] [PMID: 32492797]
- [25] Sofiane G, Wafa N. Antioxidant, antimicrobial and anti-inflammatory activities development of methanol extract of *Cyclamen africanum* B. et R., growth in Jijel-Algeria. J Drug Deliv Ther 2020; 10 (1-s): 130-4.
 [http://dx.doi.org/10.22270/jddt.v10i1-s.3883]
- [26] Qi R, Li X, Zhang X, et al. Ethanol extract of Elephantopus scaber Linn. Attenuates inflammatory response via the inhibition of NF-κB signaling by dampening p65-DNA binding activity in lipopolysaccharide-activated macrophages. J Ethnopharmacol 2020; 250: 112499. [http://dx.doi.org/10.1016/j.jep.2019.112499] [PMID: 31877363]
- [27] Pudžiuvelytė L, Liaudanskas M, Jekabsone A, Sadauskienė I, Bernatonienė J. Elsholtzia ciliata (Thun b.) Hyl. Extracts from different plant parts: phenolic composition, antioxidant, and anti-inflammatory activities. Molecules Basel. MDPI 2020; 25(5): 2020. [http://dx.doi.org/10.3390/molecules25051153]
- [28] Yang M, Wang Y, Patel G, et al. In vitro and in vivo anti-inflammatory effects of different extracts from Epigynum auritum through down-regulation of NF-κB and MAPK signaling pathways. J Ethnopharmacol 2020; 261: 113105. [http://dx.doi.org/10.1016/j.jep.2020.113105] [PMID: 32590114]
- [29] Costa WK. Essential oil from Eugenia *stipitata McVaugh* leaves has antinociceptive, antiinflammatory and antipyretic activities without showing toxicity in mice. Ind Crops Prod 2020; 144: 112059. [http://dx.doi.org/10.1016/j.indcrop.2019.112059]
- [30] Shin J-I, Jeon Y-J, Lee S, Lee YG, Kim JB, Kwon HC, et al. Apoptotic and anti-inflammatory effects of Eupatorium japonicum thunb. in rheumatoid arthritis fibroblast-like synoviocytes. BioMed research international 2018.
- [31] Lee JJ, Kim KH, Kim EJ, et al. Anti-inflammatory activity of the decoction of Forsythia suspensa (Thunb.) Vahl is related to Nrf2 and A20. J Ethnopharmacol 2018; 227: 97-104. [http://dx.doi.org/10.1016/j.jep.2018.08.027] [PMID: 30145174]

- [32] Studzińska-Sroka E, Dudek-Makuch M, Chanaj-Kaczmarek J, et al. Anti-inflammatory activity and phytochemical profile of *Galinsoga parviflora* Cav. Molecules 2018; 23(9): 2133. [http://dx.doi.org/10.3390/molecules23092133] [PMID: 30149540]
- [33] Napagoda M, Gerstmeier J, Butschek H, et al. The anti-inflammatory and antimicrobial potential of selected ethnomedicinal plants from Sri Lanka. Molecules 2020; 25(8): 1894. [http://dx.doi.org/10.3390/molecules25081894] [PMID: 32326068]
- [34] Bhanuvalli R S, Lotha R, Sivasubramanian A. Phenyl propanoid rich extract of edible plant Halosarcia indica exert diuretic, analgesic, and anti-inflammatory activity on Wistar albino rats. Nat Prod Res 2020; 34(11): 1616-20. [http://dx.doi.org/10.1080/14786419.2018.1521404] [PMID: 30394103]
- [35] Hongping Z, Yashen H, Tuyghun E, Min J, Fengsen L. A comparative analysis of the antiinflammatory effects of *Hyssopus cuspidatus* Boriss. Essential oil and aspirin on chronic inflammation models in mice. Int J Clin Exp Med 2019; 12(7): 8261-70.
- [36] Javed F, Jabeen Q, Aslam N, Awan AM. Pharmacological evaluation of analgesic, anti-inflammatory and antipyretic activities of ethanolic extract of Indigofera argentea Burm. f. J Ethnopharmacol 2020; 259: 112966.
 [http://dx.doi.org/10.1016/j.jep.2020.112966] [PMID: 32418900]
- [37] Yu HH, Lin Y, Zeng R, *et al.* Analgesic and anti-inflammatory effects and molecular mechanisms of *Kadsura heteroclita* stems, an anti-arthritic Chinese Tujia ethnomedicinal herb. J Ethnopharmacol 2019; 238: 111902.
 [http://dx.doi.org/10.1016/j.jep.2019.111902] [PMID: 31018145]
- [38] Alfaifi M, Alsayari A, Gurusamy N, et al. Analgesic, Anti-Inflammatory, Cytotoxic Activity Screening and UPLC-PDA-ESI-MS Metabolites Determination of Bioactive Fractions of *Kleinia* pendula. Molecules 2020; 25(2): E418. [http://dx.doi.org/10.3390/molecules25020418] [PMID: 31968561]
- [39] Moreno MA, Zampini IC, Isla MI. Antifungal, anti-inflammatory and antioxidant activity of bi-herbal mixtures with medicinal plants from Argentinean highlands. J Ethnopharmacol 2020; 253: 112642. [http://dx.doi.org/10.1016/j.jep.2020.112642] [PMID: 32035220]
- [40] Chen Z, Zhang C, Gao F, et al. A systematic review on the rhizome of Ligusticum chuanxiong Hort. (Chuanxiong). Food Chem Toxicol 2018; 119: 309-25.
 [http://dx.doi.org/10.1016/j.fct.2018.02.050] [PMID: 29486278]
- [41] Gu J, Su S, Guo J, Zhu Y, Zhao M, Duan JA. Anti-inflammatory and anti-apoptotic effects of the combination of *Ligusticum chuanxiong* and *Radix Paeoniae* against focal cerebral ischaemia *via* TLR4/MyD88/MAPK/NF-κB signalling pathway in MCAO rats. J Pharm Pharmacol 2018; 70(2): 268-77.
 [http://dx.doi.org/10.1111/jphp.12841] [PMID: 29193143]
- [42] Yong KY, Shukkoor MSA, Chin JH. Analgesic activity of chloroform and methanolic leaf extracts of Manilkara zapota. Mater Today Proc 2020. [http://dx.doi.org/10.1016/j.matpr.2020.05.687]
- [43] Jasamai M, Jalil J, Saadawi S, Jantan I. Inhibitory effects of *Mitrella kentii* extracts on inflammatory mediators' biosynthesis and binding. J Herbs Spices Med Plants 2020; 26(1): 30-9. [http://dx.doi.org/10.1080/10496475.2019.1663771]
- [44] Jeong SG, Kim S, Kim HG, *et al.* Mycetia cauliflora methanol extract exerts anti-inflammatory activity by directly targeting PDK1 in the NF-κB pathway. J Ethnopharmacol 2019; 231: 1-9. [http://dx.doi.org/10.1016/j.jep.2018.11.013] [PMID: 30415059]
- [45] Beaulah GGP, Paulpriya K, Doss A. Elucidation of Analgesic Activity of *Niebuhria Apetala* Dunn. Studies in Indian Place Names 2020; 40(70): 3386-92.
- [46] Zahran EM. The anti-inflammatory activity and LD50 of Ocimum forskolei Benth., family Lamiaceae.

Journal of advanced Biomedical and Pharmaceutical Sciences 2019; 2(3): 116-20.

[47] Jo MS, Lee S, Yu JS, Baek SC, Cho Y-C, Kim KH. Megastigmane derivatives from the cladodes of *Opuntia humifusa* and their nitric oxide inhibitory activities in macrophages. J Nat Prod 2020; 83(3): 684-92.
 [http://dx.doi.org/10.1021/accineture.d0h011201[DMID: 22118424].

[http://dx.doi.org/10.1021/acs.jnatprod.9b01120] [PMID: 32118424]

- [48] Zhou Y, Tao H, Wang A, *et al.* Chinese herb pair Paeoniae Radix Alba and Atractylodis Macrocephalae Rhizoma suppresses LPS-induced inflammatory response through inhibiting MAPK and NF-kB pathway. Chin Med 2019; 14: 2. [http://dx.doi.org/10.1186/s13020-019-0224-2] [PMID: 30728853]
- [49] Lu S, Zhang Y, Li H, Zhang J, Ci Y, Han M. Ginsenoside Rb1 can ameliorate the key inflammatory cytokines TNF-α and IL-6 in a cancer cachexia mouse model. BMC complementary medicine and therapies 2020; 20(1): 11.
- [50] Harikrishnan H, Jantan I, Haque MA, Kumolosasi E. Anti-inflammatory effects of Phyllanthus amarus Schum. & Thonn. through inhibition of NF-κB, MAPK, and PI3K-Akt signaling pathways in LPSinduced human macrophages. BMC Complement Altern Med 2018; 18(1): 224. [http://dx.doi.org/10.1186/s12906-018-2289-3] [PMID: 30045725]
- [51] Rivera D, Ocampo Y, Franco LA. *Physalis angulata* calyces modulate macrophage polarization and alleviate chemically induced intestinal inflammation in mice. Biomedicines 2020; 8(2): E24. [http://dx.doi.org/10.3390/biomedicines8020024] [PMID: 32033338]
- [52] Nakalembe L, Kasolo J, Nyatia E, Lubega A, Bbosa G. Analgesic and anti-inflammatory activity of total crude leaf extract of *Phytolacca dodecandra* in wistar albino rats. Neurosci Med 2019; 10: 259-71.
 [http://dx.doi.org/10.4236/nm.2019.103020]
- [53] Auliafendri N, Rosidah , Yuandani , Suryani S, Satria D. Rosidah, Yuandani, Suryani S, Satria D. The immunomodulatory activities of Picria fel-terrae lour herbs towards RAW 264.7 Cells. Open Access Maced J Med Sci 2019; 7(1): 24-8. [http://dx.doi.org/10.3889/oamjms.2019.017] [PMID: 30740154]
- [54] Karbab A, Mokhnache K, Ouhida S, *et al.* Anti-inflammatory, analgesic activity, and toxicity of Pituranthos scoparius stem extract: An ethnopharmacological study in rat and mouse models. J Ethnopharmacol 2020; 258: 112936.
 [http://dx.doi.org/10.1016/j.jep.2020.112936] [PMID: 32376367]
- [55] Chen Y, Yu H, Guo F, Wu Y, Li Y. Antinociceptive and anti-inflammatory activities of a standardizedextract of bis-iridoids from *Pterocephalus hookeri*. J Ethnopharmacol 2018; 216: 233-8. [http://dx.doi.org/10.1016/j.jep.2018.01.035] [PMID: 29410154]
- [56] Wu Y-C, Guo C-X, Zhu Y-Z, Li Y-M, Guo F-J, Zhu G-F. Four new bis-iridoids isolated from the traditional Tibetan herb *Pterocephalus hookeri*. Fitoterapia 2014; 98: 104-9. [http://dx.doi.org/10.1016/j.fitote.2014.07.015] [PMID: 25065705]
- [57] Zhang M, Fan H, Tan S, *et al.* The Chinese medicinal herb decoction QRZSLXF enhances antiinflammatory effect in TNBS-induced colitis *via* balancing Th17/Tregs differentiation. J Ethnopharmacol 2020; 251: 112549. [http://dx.doi.org/10.1016/j.jep.2020.112549] [PMID: 31918016]
- [58] Xu X, Tan T, Zhang J, et al. Isolation of chemical constituents with anti-inflammatory activity from *Reineckia carnea* herbs. J Asian Nat Prod Res 2020; 22(4): 303-15. [http://dx.doi.org/10.1080/10286020.2019.1575818] [PMID: 30843729]
- [59] Lee J-W, Kim Y-S, Dong X, Park J-S, Shin W-B, Kim S-J, et al. Anti-inflammatory effect of *Rhodiola crenulata* extracts through the down-regulation of MyD88 dependent pathway and induction of autophagy. J Funct Foods 2020; 64: 103703. [http://dx.doi.org/10.1016/j.jff.2019.103703]

112391.

- [60] Carneiro NVQ, Silva HBFD, Silva RRD, *et al.* Sambucus australis Modulates Inflammatory Response *via* Inhibition of Nuclear Factor Kappa B (NF-kB) in vitro. An Acad Bras Cienc 2019; 91(1): e20170831.
 [http://dx.doi.org/10.1590/0001-3765201920170831] [PMID: 30916148]
- [61] Rozenberg K, Wollman A, Ben-Shachar M, Argaev-Frenkel L, Rosenzweig T. Anti- Sambucus australis inflammatory effects of Sarcopoterium spinosum extract. J Ethnopharmacol 2020; 249:

[http://dx.doi.org/10.1016/j.jep.2019.112391] [PMID: 31730890]

- [62] A Azadmehr*, M Goudarzvand, P Saadat , H Ebrahimi, R Hajiaghaee, Miri N Sadat. Immunomodulatory and anti-inflammatory effects of Scrophularia megalantha ethanol extract on an experimental model of multiple sclerosis. Research Journal of Pharmacognosy 2019; 6(1): 43-50.
- [63] Liu H-L, Kao T-H, Shiau C-Y, Chen B-H. Functional components in *Scutellaria barbata* D. Don with anti-inflammatory activity on RAW 264.7 cells. J Food Drug Anal 2018; 26(1): 31-40. [http://dx.doi.org/10.1016/j.jfda.2016.11.022] [PMID: 29389569]
- [64] Linghu K-G, Zhao GD, Xiong W, *et al.* Comprehensive comparison on the anti-inflammatory effects of three species of Sigesbeckia plants based on NF-κB and MAPKs signal pathways in vitro. J Ethnopharmacol 2020; 250: 112530. [http://dx.doi.org/10.1016/j.jep.2019.112530] [PMID: 31883476]
- [65] Zhong Z, Zhang Q, Tao H, et al. Anti-inflammatory activities of Sigesbeckia glabrescens Makino: combined in vitro and in silico investigations. Chin Med 2019; 14: 35. [http://dx.doi.org/10.1186/s13020-019-0260-y] [PMID: 31572487]
- [66] Xian Y-F, Hu Z, Ip S-P, et al. Comparison of the anti-inflammatory effects of Sinapis alba and Brassica juncea in mouse models of inflammation. Phytomedicine 2018; 50: 196-204. [http://dx.doi.org/10.1016/j.phymed.2018.05.010] [PMID: 30466979]
- [67] Ma H, Huang Q, Qu W, et al. In vivo and in vitro anti-inflammatory effects of Sophora flavescens residues. J Ethnopharmacol 2018; 224: 497-503. [http://dx.doi.org/10.1016/j.jep.2018.06.019] [PMID: 29913301]
- [68] Mohibbullah M, Lee YJ, Park H-J, Kim SK, Kang J-S, Kim A, et al. The medicinal herb Spatholobus suberectus with promising in vitro antioxidant and anti-inflammatory potentials and its phytochemical characterization by RP-HPLC analysis. J Food Biochem 2018; 42(2): e12480. [http://dx.doi.org/10.1111/jfbc.12480]
- [69] Mou L-Y, Wu H-Y, Ma E-G, et al. Two new xanthone glycosides from Swertia punicea Hemsl. and their anti-inflammatory activity. Nat Prod Res 2020; 34(10): 1423-9. [http://dx.doi.org/10.1080/14786419.2018.1509325] [PMID: 30453776]
- [70] Owor RO, Bedane KG, Zühlke S, et al. Anti-inflammatory flavanones and flavones from *Tephrosia linearis*. J Nat Prod 2020; 83(4): 996-1004.
 [http://dx.doi.org/10.1021/acs.jnatprod.9b00922] [PMID: 32155073]
- Silva AM, Martins-Gomes C, Souto EB, *et al.* Thymus zygis subsp. zygis an endemic portuguese plant: phytochemical profiling, antioxidant, anti-proliferative and anti-inflammatory activities. Antioxidants 2020; 9(6): 482.
 [http://dx.doi.org/10.3390/antiox9060482] [PMID: 32503184]
- [72] Santos ES, de Morais Oliveira CD, Alencar Menezes IR, et al. Anti-inflammatory activity of herb products from *Licania rigida* Benth. Complement Ther Med 2019; 45: 254-61. [http://dx.doi.org/10.1016/j.ctim.2019.06.001] [PMID: 31331571]
- [73] Yang L, Zhu Y, He Z, Zhang T, Xiao Z, Xu R, *et al.* Plantanone D, a new rare methyl-flavonoid from the flowers of *Hosta plantaginea* with anti-inflammatory and antioxidant activities. Nat Prod Res 2020;
 •••: 1-7.
 [PMID: 32067484]

- Yang L, Lin Y-m, He Z-w, Zhang T-f, Li Y, Xie X-t, *et al.* Hostaflavanol A, a new anti-inflammatory and antioxidant activities flavanol from the flowers of *Hosta plantaginea*. Med Chem Res 2020; 29(3): 426-30.
 [http://dx.doi.org/10.1007/s00044-019-02491-6]
- [75] Wu JY, Chen YJ, Bai L, Liu YX, Fu XQ, Zhu PL, et al. Hostaflavanol A, a new anti-inflammatory and antioxidant activities flavanol from the flowers of Hosta plantaginea. Medicinal Chemistry Research 2020; 29(3): 426-30.
- [76] Lin AM, Ping YH, Chang GF, et al. Neuroprotective effect of oral S/B remedy (Scutellaria baicalensis Georgi and Bupleurum scorzonerifolfium Willd) on iron-induced neurodegeneration in the nigrostriatal dopaminergic system of rat brain. J Ethnopharmacol 2011; 134(3): 884-91. [http://dx.doi.org/10.1016/j.jep.2011.01.056] [PMID: 21296142]
- [77] Yan JJ, Du GH, Qin XM, Gao L. Baicalein attenuates the neuroinflammation in LPS-activated BV-2 microglial cells through suppression of pro-inflammatory cytokines, COX2/NF-κB expressions and regulation of metabolic abnormality. Int Immunopharmacol 2020; 79: 106092. [http://dx.doi.org/10.1016/j.intimp.2019.106092] [PMID: 31863920]
- [78] Li KK, Shen SS, Deng X, et al. Dihydrofisetin exerts its anti-inflammatory effects associated with suppressing ERK/p38 MAPK and Heme Oxygenase-1 activation in lipopolysaccharide-stimulated RAW 264.7 macrophages and carrageenan-induced mice paw edema. Int Immunopharmacol 2018; 54: 366-74. [http://dx.doi.org/10.1016/j.intimp.2017.11.034] [PMID: 29202300]

- [79] Liu X, Yin C, Cao Y, Zhou J, Wu T, Cheng Z. Chemical constituents from *Gueldenstaedtia verna* and their anti-inflammatory activity. Nat Prod Res 2018; 32(10): 1145-9. [http://dx.doi.org/10.1080/14786419.2017.1320795] [PMID: 28441883]
- [80] Khouya T, Ramchoun M, Amrani S, et al. Anti-inflammatory and anticoagulant effects of polyphenolrich extracts from *Thymus atlanticus*: An *in vitro* and *in vivo* study. J Ethnopharmacol 2020; 252: 112475.

[http://dx.doi.org/10.1016/j.jep.2019.112475] [PMID: 31843575]

- [81] Villalva M, Jaime L, Aguado E, Nieto JA, Reglero G, Santoyo S. Anti-Inflammatory and Antioxidant activities from the basolateral fraction of Caco-2 cells exposed to a rosmarinic acid enriched extract. J Agric Food Chem 2018; 66(5): 1167-74. [http://dx.doi.org/10.1021/acs.jafc.7b06008] [PMID: 29345918]
- [82] Choi HG, Tran PT, Lee J-H, Min BS, Kim JA. Anti-inflammatory activity of caffeic acid derivatives isolated from the roots of *Salvia miltiorrhiza* Bunge. Arch Pharm Res 2018; 41(1): 64-70. [http://dx.doi.org/10.1007/s12272-017-0983-1] [PMID: 29124660]
- [83] Liu H, Ma S, Xia H, Lou H, Zhu F, Sun L. Anti-inflammatory activities and potential mechanisms of phenolic acids isolated from *Salvia miltiorrhiza* f. *alba* roots in THP-1 macrophages. J Ethnopharmacol 2018; 222: 201-7. [http://dx.doi.org/10.1016/j.jep.2018.05.008] [PMID: 29751125]
- [84] Yuan J, Cheng W, Zhang G, *et al.* Protective effects of iridoid glycosides on acute colitis *via* inhibition of the inflammatory response mediated by the STAT3/NF-κB pathway. Int Immunopharmacol 2020; 81: 106240.
 [http://dx.doi.org/10.1016/j.intimp.2020.106240] [PMID: 32044657]
- [85] Wang F, Jia Q-W, Yuan Z-H, et al. An anti-inflammatory C-stiryl iridoid from Camptosorus sibiricus Rupr. Fitoterapia 2019; 134: 378-81.
 [http://dx.doi.org/10.1016/j.fitote.2019.03.009] [PMID: 30880242]
- [86] He J, Lu X, Wei T, Dong Y, Cai Z, Tang L, et al. Asperuloside and asperulosidic acid exert an antiinflammatory effect via suppression of the NF-κB and MAPK signaling pathways in LPS-induced RAW 264.7 macrophages. Int J Mol Sci 2018; 19(7): 2027. [http://dx.doi.org/10.3390/ijms19072027]

- [87] Chen Y, Shou K, Gong C, Yang H, Yang Y, Bao T. Anti-Inflammatory effect of geniposide on osteoarthritis by suppressing the activation of p38 MAPK Signaling Pathway. BioMed Res Int 2018; 2018: 8384576.
 [http://dx.doi.org/10.1155/2018/8384576] [PMID: 29682561]
- [88] Wang Y-M, Ming W-Z, Liang H, Wang Y-J, Zhang Y-H, Meng D-L. Isoquinolines from national herb *Corydalis tomentella* and neuroprotective effect against lipopolysaccharide-induced BV2 microglia cells. Bioorg Chem 2020; 95: 103489. [http://dx.doi.org/10.1016/j.bioorg.2019.103489] [PMID: 31862456]
- [89] Qiao B, Wang H, Wang C, Liang M, Huang K, Li Y. Dauricine negatively regulates lipopolysaccharide- or cecal ligation and puncture-induced inflammatory response *via* NF-κB inactivation. Arch Biochem Biophys 2019; 666: 99-106. [http://dx.doi.org/10.1016/j.abb.2019.03.018] [PMID: 30946805]
- [90] Wu X, Li X, Dang Z, Jia Y. Berberine demonstrates anti-inflammatory properties in Helicobacter pylori-infected mice with chronic gastritis by attenuating the Th17 response triggered by the B cellactivating factor. J Cell Biochem 2018; 119(7): 5373-81. [http://dx.doi.org/10.1002/jcb.26681] [PMID: 29345340]
- [91] Shen CY, Xu XL, Yang LJ, Jiang JG. Identification of narciclasine from Lycoris radiata (L'Her.) Herb. and its inhibitory effect on LPS-induced inflammatory responses in macrophages. Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association 2019; 125: 605-13.
- [92] Ji KY, Kim KM, Kim YH, et al. The enhancing immune response and anti-inflammatory effects of Anemarrhena asphodeloides extract in RAW 264.7 cells. Phytomedicine 2019; 59: 152789. [http://dx.doi.org/10.1016/j.phymed.2018.12.012] [PMID: 31009851]
- [93] Gonçalves ECD, Assis PM, Junqueira LA, et al. Citral Inhibits the Inflammatory Response and Hyperalgesia in Mice: The Role of TLR4, TLR2/Dectin-1, and CB2 Cannabinoid Receptor/ATP-Sensitive K⁺ Channel Pathways. J Nat Prod 2020; 83(4): 1190-200. [http://dx.doi.org/10.1021/acs.jnatprod.9b01134] [PMID: 32150408]
- [94] Zhang Y, Huang X, Chen H, et al. Discovery of anti-inflammatory terpenoids from Mallotus conspurcatus croizat. J Ethnopharmacol 2019; 231: 170-8.
 [http://dx.doi.org/10.1016/j.jep.2018.11.002] [PMID: 30445108]
- [95] He H, Jiang H, Chen Y, et al. Oridonin is a covalent NLRP3 inhibitor with strong anti-inflammasome activity. Nat Commun 2018; 9(1): 2550. [http://dx.doi.org/10.1038/s41467-018-04947-6] [PMID: 29959312]
- [96] Wang X-F, Li H, Jiang K, et al. Anti-inflammatory constituents from Perilla frutescens on lipopolysaccharide-stimulated RAW264.7 cells. Fitoterapia 2018; 130: 61-5. [http://dx.doi.org/10.1016/j.fitote.2018.08.006] [PMID: 30121232]
- [97] Lu Z, Xie P, Zhang D, et al. 3-Dehydroandrographolide protects against lipopolysaccharide-induced inflammation through the cholinergic anti-inflammatory pathway. Biochem Pharmacol 2018; 158: 305-17.
 [http://dx.doi.org/10.1016/j.bcp.2018.10.034] [PMID: 30391477]
 - 3] Linghu K-G, Ma QS, Zhao GD, *et al.* Leocarpinolide B attenuates LPS-induced inflammation on
- [98] Linghu K-G, Ma QS, Zhao GD, *et al.* Leocarpinolide B attenuates LPS-induced inflammation on RAW264.7 macrophages by mediating NF-κB and Nrf2 pathways. Eur J Pharmacol 2020; 868: 172854. [http://dx.doi.org/10.1016/j.ejphar.2019.172854] [PMID: 31837308]
- [99] Zou YH, Zhao L, Xu YK, *et al.* Anti-inflammatory sesquiterpenoids from the Traditional Chinese Medicine Salvia plebeia: Regulates pro-inflammatory mediators through inhibition of NF-κB and Erk1/2 signaling pathways in LPS-induced Raw264.7 cells. J Ethnopharmacol 2018; 210: 95-106. [http://dx.doi.org/10.1016/j.jep.2017.08.034] [PMID: 28847754]

- [100] Hu Y-M, Liu W-J, Li M-X, et al. Two new labdane diterpenoids from aerial parts of Leonurus japonicus and their anti-inflammatory activity. Nat Prod Res 2019; 33(17): 2490-7. [http://dx.doi.org/10.1080/14786419.2018.1455040] [PMID: 29631433]
- [101] Wang Z, Zhao X, Liu X, et al. Anti-diabetic activity evaluation of a polysaccharide extracted from Gynostemma pentaphyllum. Int J Biol Macromol 2019; 126: 209-14. [http://dx.doi.org/10.1016/j.ijbiomac.2018.12.231] [PMID: 30590141]
- [102] Wang Z, Wang Z, Huang W, *et al.* Antioxidant and anti-inflammatory activities of an anti-diabetic polysaccharide extracted from *Gynostemma pentaphyllum* herb. Int J Biol Macromol 2020; 145: 484-91.

[http://dx.doi.org/10.1016/j.ijbiomac.2019.12.213] [PMID: 31883891]

- [103] Zhan R, Xia L, Shao J, Wang C, Chen D. Polysaccharide isolated from Chinese jujube fruit (*Zizyphus jujuba* cv. Junzao) exerts anti-inflammatory effects through MAPK signaling. J Funct Foods 2018; 40: 461-70.
 [http://dx.doi.org/10.1016/j.jff.2017.11.026]
- [104] Ishii T, Okuyama T, Noguchi N, *et al.* Antiinflammatory constituents of *Atractylodes chinensis* rhizome improve glomerular lesions in immunoglobulin A nephropathy model mice. J Nat Med 2020; 74(1): 51-64.

[http://dx.doi.org/10.1007/s11418-019-01342-3] [PMID: 31270736]

- [105] Zhou Y, Jin M, Jin C, et al. A new aryldihydronaphthalene-type lignan and other metabolites with potential anti-inflammatory activities from *Corispermum mongolicum* Iljin. Nat Prod Res 2020; 34(2): 225-32. [http://dx.doi.org/10.1080/14786419.2018.1527835] [PMID: 30580619]
- [106] Kim MJ, Kim D-C, Kwon J, et al. Anti-inflammatory Metabolites from Chaetomium nigricolor. J Nat Prod 2020; 83(4): 881-7. [http://dx.doi.org/10.1021/acs.jnatprod.9b00560] [PMID: 32163284]
- [107] Raju R, Singh A, Gunawardena D, Reddell P, Münch G. Diarylheptanoids with anti-inflammatory activity from the rhizomes of *Pleuranthodium racemigerum* (Zingiberaceae). Phytochem Lett 2019; 30: 10-3.
 [http://dx.doi.org/10.1016/j.phytol.2019.01.004]
- [108] Chen Y, Ruan J, Sun F, et al. Anti-inflammatory Limonoids From Cortex Dictamni. Front Chem 2020; 8: 73.
 [http://dx.doi.org/10.3389/fchem.2020.00073] [PMID: 32185157]
- [109] Lu J, Fang K, Wang S, et al. Anti-Inflammatory effect of columbianetin on lipopolysaccharidestimulated human peripheral blood mononuclear cells. Mediators Inflamm 2018; 2018: 9191743. [http://dx.doi.org/10.1155/2018/9191743] [PMID: 29849500]
- [110] Wu X, Gao H, Hou Y, *et al.* Dihydronortanshinone, a natural product, alleviates LPS-induced inflammatory response through NF-κB, mitochondrial ROS, and MAPK pathways. Toxicol Appl Pharmacol 2018; 355: 1-8. [http://dx.doi.org/10.1016/j.taap.2018.06.007] [PMID: 29906494]
- [111] Yang C, Liu P, Wang S, et al. Shikonin exerts anti-inflammatory effects in LPS-induced mastitis by inhibiting NF-κB signaling pathway. Biochem Biophys Res Commun 2018; 505(1): 1-6. [http://dx.doi.org/10.1016/j.bbrc.2018.08.198] [PMID: 30224056]
- [112] Geng Q, Wei Q, Wang S, et al. Physcion 8 □ O □β □ glucopyranoside extracted from Polygonum cuspidatum exhibits anti □ proliferative and anti □ inflammatory effects on MH7A rheumatoid arthritis □ derived fibroblast □ like synoviocytes through the TGF □ β/MAPK pathway. Int J Mol Med 2018; 42(2): 745-54. [PMID: 29717774]
- [113] Wang B-B, Gao Y, Chen L-Y, Zhang C-L, Zhang X-Q, Zhang H-L. New constituents from the low

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polar fraction of the fruits of *Crataegus dahurica* and their anti-inflammatory activity in RAW264.7 cells. Chem Biodivers 2020; 17(2): e1900609. [http://dx.doi.org/10.1002/cbdv.201900609] [PMID: 31916412]

Phenolic Compounds and their Biological and Pharmaceutical Activities

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Abstract: Phenolic compounds play an essential role in plants and foods. These compounds are well known for their biological and pharmaceutical activities. These compounds act as colorants and antioxidants. Research on phenolic compounds is mainly focused on their antioxidant properties. These compounds showed significant effects on chronic degenerative diseases, such as central neurodegenerative disorders, cataracts, macular degeneration (age-related), diabetes mellitus, cardiovascular complication, and cancer. These compounds also showed implications on human health since increased exposure to free radicals might lead to an increased risk of degenerative diseases. Fruits and vegetables are rich in phenolic compounds. The phenolic compound consists of one (phenolic acids) or more polyphenols aromatic structures attached to a hydroxyl group. The phenolic compound is found in combination with mono or polysaccharides, and they can occur in the group as an ester or methyl ester. Their biological and pharmaceutical activities are based on their phenolic ring and a hydroxyl group. Apart from antioxidant activity, they have many other therapeutic effects on human health. Among the several classes of phenolic compounds, flavonoids, tannins, and phenolic acids are considered as main dietary phenolic compounds. In this chapter, we have summarized the biological and pharmaceutical activities related to different classes of phenolic compounds.

Keywords: Antioxidant activity, Biological activity, Cardio-protective, Flavonoids, Oxidative stress, Pharmaceutical activity, Polyphenols.

INTRODUCTION

Phenolic compounds are very common and widespread groups of plant secondary metabolites. The "phenolic" or "polyphenol" substances possess one aromatic ring, and one hydroxyl (phenol), or multiple hydroxyls (polyphenol), which

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includes functional groups (esters, glycosides, methyl ethers) that are biogenetically derived from shikimate phenylpropanoids- flavonoids pathways [1, 2]. Various classes of phenols have been classified on the presence of their basic skeleton: C6 (simple phenol, benzoquinones), C6-C1 (phenolic acids like gallic ellagic acid, protocatechuic acid, syringic acid, vanillic acid, acid. hydroxybenzoates and p-hydroxybenzoic acid), C6-C2 (phenylacetic acid and acetophenone), C6-C3 (aesculatin, caffeic acid, coumaric acid, ferulic acid, hydroxycinnamic acid derivatives, chlorogenic acid, curcumin, phenylpropanes, chromones, umbelliferone), C6-C4 (napthoquinones), C6-C1-C6 (xanthones), C6-(stilbenes, resveratrol, anthroquinones), C6-C3-C6 (flavonoids, C2-C6 isoflavonoids, isoflavones, flavanones, anthocyanidins, apigenin, (+)-catechin, (---epicatechin, cyanidin, daidzein, delphinidin, erodictoyl, Isorhamnetin, genistin, glycitien, hesperetin, kaempferol, luteolin, malvidin, myricetin, narengenin, pelargonidin, petunidin, peounidin, quercetin), (C6-C3), (lignans, neolignans, lariciresinol, matairesinol, medioresinol, pinoresinol, secoisolariciresinol), (C6- $(C3-C6)_2$ (bioflavonoids), $(C6)_n$ (catechol, melanins), $(C6-C3-C6)_n$ (condensed tannins, catechin polymers, epicatechin polemers, hydrolysable tannins, casuarictin, punicalagin, gallotannins) [2 - 4]. Polyphenols are further divided into two main groups; one is a flavonoid, and the other is a non-flavonoid [5]. Flavonoids are the most studied and largest group amongst plant phenols. Anthocyanidins, flavones, isoflavones, flavanones, flavonols, dihydroflavonols, flavan-3-ols, proanthocyanidins, and chalcones are all members of the flavonoid group, which includes compounds with the C6-C3-C6 structure. The nonflavonoid group includes simple phenols, acetophenones, hydrolyzable tannins, benzophenones, benzoic aldehydes, coumarins, stilbenes, lignans, xanthones, secoiridids, and phenolic acids, such as gallic acid. Because of their potential free radical scavenging activity [6], anti-inflammatory activity [7], and capacity to lower oxidative stress, polyphenols can be an important part of the human diet, preventing neurodegenerative disorders [8], cancer [9], and other diseases. In plants, they are responsible for the development of resistance to pathogens, growth, pigmentation, and reproduction. Polyphenols also show a protectant nature against bacterial and viral pathogens. They form main classes of secondary metabolites, and almost 8150 flavonoids have been identified, having different structures, such as monomeric, dimeric, and polymeric [10, 11]. They are abundantly present in leaves or bark together with other metabolites. As far as the activity is concerned, almost all the flavonoids possess strong antioxidant activity, and these are commonly present in vegetables and fruits. Therefore, plant containing polyphenols are rich source of compounds having beneficial effect on health and are indicated in chronic diseases related to oxidative stress [2]. After ingestion, these are either absorbed through intestine (small percentage up-to 5-10 %) or found unchanged in the colon. After absorption, polyphenols exert

biological action or works as a prodrug. These compounds are metabolized through phase I (oxidation, reduction, or hydrolysis) or phase II reaction (conjugated reaction) to form the water-soluble metabolite and excreted through the urine [12].

Food Sources of Polyphenols

Phenolic compounds are abundantly present in plants (whole grains, legumes, fruits, and vegetables, coffee and tea beverages). In fruits, they are mainly found in apples, berries, cherries, citrus fruits, grapes, peaches, *etc*. The most common polyphenols are phenolic acids (benzoic acids, gallic acid, cinnamic acid, sinapic acid, vanillic acid, and ferulic acid, *etc*.), flavonoids, coumarins, tannins, lignans, stilbenes, and proanthocyanidins [13].

Biological and Pharmaceutical Activities of Polyphenols

Plant polyphenols received scientific attention due to their beneficial effect on human health. As per the literature reviewed, plant phenolic compounds bear strong antioxidant activity. These compounds are used in the treatment of several diseases such as cancer and other oxidative stress related human diseases. Multiple studies supported that oxidative stress plays a vital role in the occurrence of neurodegeneration, cancer, cardiovascular complication, muscular degeneration, antibacterial effect, immune system promoting effect, anti-inflammatory effect, UV radiation protective effect, etc. The reactive oxygen species include superoxide radical, nitric oxide radical, peroxynitrite anion, hydroxyl-peroxyl, peroxyl, alkoxyl, and hydroxyl free radicals. These oxidative species cause damage to vital biomolecules such as lipids, DNA, and proteins. Polyphenols are divided into different classes such as flavonoids, stilbenes, coumarins, lignans, tannins, curcuminoids, phenolic acid, etc. Flavonoid is the largest class and is further subdivided into flavones, isoflavones, flavonols, flavanones, flavanonols, flavanols, anthocyanidins, and anthocyanins. Table 1 enlisted the types of polyphenols, and Table 2 enlisted the detailed biological and pharmaceutical activities of individual polyphenolic compounds. Flavonoids have a 15-carbon skeleton structure with two phenyl rings and a heterocyclic ring. Brief details of flavonoids are as follows:

• Anthocyanidins: They are commonly present in fruits, flowers, leaves, and tubers. The main sources are red, purple, and blue berries, pomegranates, plums, red wine, red and purple grapes. These are plant pigments. Change in pH may lead to a change in its colour, such as red, purple, black, blue, *etc*. These compounds have flavylium and oxonium ions. These compounds showed benefits to heart health, antioxidant, anti-obesity, and anti-diabetic effects. Example-Cyanidin, Delphinidin, Pelargonidin, *etc*.

• Flavones: They are commonly present in spices, fruits, and vegetables, *e.g.*, celery, various herbs, hot peppers, and parsley. Studies showed protective effects in cardiovascular complications and neurological disorders. *In vitro* and *in vivo* studies confirmed the neuroprotective effect and antioxidant benefits. These compounds delay drug metabolism. Example- Apigenin, Luteolin, Tangeritin, Chrysin, *etc*.

• **Flavonols:** They are commonly present in fruits and vegetables, *e.g.*, onions, leeks, brussels sprouts, broccoli, tea, berries, beans, apples, *etc.* These compounds have a 3-hydroxy flavone backbone. These compounds have antioxidant, antihistamine property and also help in the treatment of inflammatory bowel disease. Example- 3-Hydroxyflavone, Azaleatin, Fisetin, Galangin, Gossypetin, Kaempferide, Kaempferol, Myricetin, Quercetin *etc.*

• Flavan-3-ols (or Flavanols): They are commonly present in teas, cocoa, grapes, apples, berries, fava beans, and red wine. These compounds are derived from flavans. Structurally, these compounds have a 2-phenyl-3,4-dihydro-2H-ch-omen-3-ol skeleton. These compounds have anti-mutagenic, anti-diabetic, hypolipidemic, and anti-thrombogenic properties. Example- proanthocyanidins, thearubigins, catechin, theaflavins, epicatechin gallate, epigallocatechin gallate, epigallocatechin, *etc*.

• Flavanones: They are commonly present in citrus fruits. These are derived from flavone. These compounds are aromatic and colourless ketones. These compounds have a positive effect on cardiovascular problems, antioxidant and anti-inflammatory activity. Example- Naringenin, Hesperetin, Homoeriodictyol, Eriodictyol, *etc.*

• **Flavanonols:** They are commonly present in plants. These compounds have 3-hydroxy-2,3-dihydro-2-phenylchromen-4-one backbone. Example- Aromadedrin (Dihydrokaempferol), Taxifolin (Dihydroquercetin) *etc*.

• **Isoflavones:** They are commonly present in soybeans and legumes (Fabaceae or Leguminosae family plants). These are derived from isoflavone and act as phytoestrogens. These are bioactive nonsteroidal polyphenolic metabolites with antioxidant properties and showed an estrogenic effect. It showed oxidants as well as antioxidants effects. Thus, the effect on cancer (breast, endometrial, prostate cancer) is still doubtful and a subject of further research work. Example-Genistein, Daidzein, Glycitein, *etc*.

Table 1. Categories of	phenolic compounds a	and representative chemi	ical structures.

Chemical Class	Categories	Chemical Sub- Type	Compounds	Chemical Structure
Phenolic Compounds	Phenolic acids	Hydroxybenzoic acid	4-hydroxybenzoic acid, gallic acid, gentisic acid, salicylic acid, syringic acid, vanillic acid, β- resorcylic acid, protocatechuic acid	НО
				4-hydroxybenzoic acid CAS Number: 99-96-7
		Hydroxycinnamic acid	 α-Cyano-4-hydroxycinnamic acid, chlorogenic acid, caffeic acid, cichoric acid, ferulic acid, isoferulic acid, sinapic acid, trans-cinnamic acid, p-coumaric acid 	HO-OH α-Cyano-4-hydroxycinnamic acid
				CAS Number: 28166-41-8
	Flavonoids	Flavan-3-ols (flavanols)	 (+)-catechin, (-)-gallocatechin, (-)-gallocatechin gallate, (-)-epigallocatechin, (-)-epigallocatechin gallate, (-)-epicatechin gallate, (-)-epicatechin 	HO OH Catechin
				CAS Number: 7295-85-4
		Flavonols	Fisetin, flavonol, galangin, datiscetin, hyperoside, Kaempferol, morin, myricetin, quercitin, isorhamnetin, robinetin, rutin, quercetagetin	HO OH OH
				Fisetin CAS Number: 528-48-3
		Flavones	Apigenin, aposide, baicalein, baicalin, chrysin, chrysoeriol, diosmetin, luteolin, vitexin, sinensetin	HO OH OH OH
				Apigenin CAS Number: 520-36-5

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(<u>Table 1) cont</u> Chemical class	Categories	Chemical sub- type	Compounds	Chemical structure
Phenolic Compounds	Flavonoids	Isoflavones	Biochanin A, daidzein, genistein, genistin, glycetin, glycitein, puerarin	HO OH OCH ₃ Biochanin A CAS Number: 491-80-5
		Flavanones	Dihydromyricetin, eriodictyol, hesperetin, hesperidin, liquiritin, liquiritigenin, naringin, naringenin, narirutin, (+)- taxifolin	HO HO OH OH OH OH OH OH OH OH OH OH OH O
		Anthocyanidins	Peonidin, malvidin, delphinidin, petunidin, cyanidin, pelargonidin, propelargonidin	HO HO HO Peonidin CAS Number: 134-01-0
	Tannins	Hydrolyzable tannins	Gallotannins [Aglycones: Digallic acid, Gallic acid; Galloyl glucoses: Glucogallin, 1,6-Digalloyl glucose, 1,2,3-Trigalloyl glucose, 1,2,3,6-tetragalloylglucose, 1,2,3,4,6-Pentagalloyl-glucose, Hexagalloyl glucose; Galloyl quinic acids: theogallin, 1,4-Di-O-galloylquinic acid; 1,3,4-Tri-O-galloylquinic acid; Galloyl shikimic acids: 4-O-Galloyl shikimic acid; Others: 1,2,6-trigalloyl alloside], ellagitannins	$\begin{array}{c} \begin{array}{c} & & \\ & & \\ & & \\ & \\ & \\ & \\ & \\ & \\ $
		Condensed tannins	Proanthocyanidin, leucoanthocyanidin	HO OH OH Catechin CAS Number: 7295-85-4
	Coumarins	Simple coumarins	7-hydroxycoumarin (Umbelliferone), 6,7- dihydroxy coumarin (Aesculetin), 7- methoxycoumarin (Herniarin)	HO HO Aesculetin CAS Number: 305-01-1

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(Table 1) cont Chemical class	Categories	Chemical sub-type	Compounds	Chemical structure
Phenolic Compounds	Coumarins	Furanocoumarins	Psoralen, bergapten, imperatorin	
				Psoralen CAS Number: 66-97-7
		Pyranocoumarins	Khellactones, calanolides	CAS Number: 91-64-5
		Isocoumarins	Capillarin, artemidin	
				Capillarin CAS Number: 3570-28-3
		Neoflavones	4-Phenylcoumarin	°
				4-Phenylcoumarin CHEBI:71972
	Lignans	Lignanolides	Arctigenin kaerophyllin	о с с с с с с с с с с с с с с с с с с с
				Arctigenin CAS Number: 7770-78-7
		Cyclolignanolides	Chinensin	Chinensin ChemSpider ID: 4475036
		Bisepoxylignans	Eudesmin, isoeudesmin	
				,
				Eudesmin CAS Number: 29106-36-3
		Neolignans	Magnolol, Burseneolignan	K → HO → HO → HO → HO → HO
				Magnolol CAS Number: 528-43-8

Table 1) cont	1		The Chemistry Inside Spices and Herbs, Vol. 1 2			
Chemical class	Categories	Chemical sub-type	Compounds	Chemical structure		
Phenolic Compounds	Quinones	Anthraquinones	Rhein (cassic acid), dantron, emodin, aloe emodin, alizarin, munjistin, parietin (physcion)	OH O OH O OH O OH		
				Rhein CAS Number: 478-43-3		
		Phenanthraquinones	Tanshinone: dihydrotanshinone, tanshinone I, or tanshinone IIA; denbinobin			
				Dihydrotanshinone CAS Number: 87205-99-0		
		Napthoquinones	Alkannin, shikonin (enantiomer of alkannin), shikalkin (racemic mixture)	OH O CH ₃ CH ₃ CH ₃		
				Alkannin CAS Number: 517-88-4		
		Benzoquinones	Embelin, embelinol	но, но, но, но, но, но, но, но, но, но,		
				Embelin CAS Number: 550-24-3		
	Cu	ircuminoids	Curcumin, demethoxycurcumin, bis-demethoxycurcumin, ginerol	CH ₃ OH O CH ₃ O OH O OH		
			Emeror	Curcumin (Enol form) HO CUrcumin (Keto form) CAS Number: 458-37-7		

214 The Chemistry Inside Spices and Herbs, Vol. 1 (Table 1) cont.....

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(Table 1) cont Chemical class	Categories	Chemical sub- type	Compounds	Chemical structure
Phenolic Compounds	Stilbenes		Oxyresveratrol, Isorhapontigenin, piceid, piceatannol, pinostilbene, pterostilbene, resveratrol	HO HO OH OXyresveratrol
	Others	Chalcone derivatives	Isoliquiritigenin (phenolic compound of licorice), butein, phloretin	CAS Number: 29700-22-9
		Phenolic alkaloids	Demethylsalsoline, magnoflorine (benzylisoquinoline alkaloid), oleracein A, oleracein B, oleracein E	Ho Ho Ho Ho Ho Ho Ho Ho Ho Ho Ho Ho Ho H
		Phenolic terpenoids	Carnosic acid (a benzenediol abietane diterpene found in rosemary & common sage), carnosol, thymol	CAS Number: 2141-09-5
				Carnosic acid CAS Number: 3650-09-7
		m-benzo-triphenol derivatives	Agrimol A, Agrimol B, filicic acids	
				Agrimol B CAS Number: 55576-66-4

Biological & Pharmaceutical activities Human Target Phenolic compound Food source Target class Virus Target Bacteria Target PDB Entries & 3D Protein complex crystal structure Gallic acid, Ellagic blueberry, blackberry, Example- Gallic Example- Gallic acid: Example- Gallic Example- Gallic acid: Example- Gallic acid: 4Z5X, 3WRB, 3WR9, 3WR4, 3WR3 Anti-inflammatory acid, p□ Ellagic acid acid, Protocatechuic cashew nut, grapes [16], Cardioacid: Enzyme lpha-(1,3)-fucosyltransfer acid: Human Anthrax lethal factor hazelnut, mango, plums strawberry, tea, walnut, tective effect [1] (Lyase, Fucosyltransferase 4, UDP immunodeficiency (Bacillus anthracis) 3WPM, 3WKU, 4ICO, 4JOH 4'-phosphopantetheinyl ransferase ffp (Bacillus subtilis) acid. - 191. Oxidoreductas glucuronosyltransferase 1-1. irus type 1 prote glucuronosyltransferase 1-1, Carbonic anhydrase (I, II, III, V, VI, VII, IX, XII, XIV, VA VB), LDL-associated phospholipase A2, Aldose reductase, Angiotensin-Hydroxybenzoates Syringic acids, Vanillic acid Hepatoprotective [20], Anti-neoplasti activity [21], Metabolic disease Hydrolase, Protease, Transferase, Kinase Cytochrome P450, Human immunodeficiency virus type 1 reverse wine [14, 15] transcriptase, Influenza A virus [22], urogenital lisease [23], derma disease [24], hosphodiesterase Transcription factor converting enzyme, ceraldehyde-3-phosphate Neuraminidase unc ssified Glva respiratory disease [25], oral health [26] dehydrogenase liver, ADAM17, Cytochrome P450 19A1, Dipeptidyl peptidase IV, Hepatocyte growth factor oroteins, epigenetic egulator, adhesion transporter, nembrane receptor Tannin acyl hydrolase in plex with gallic acid (PDB 4J0H) other cytosolic ceptor. Cyclooxygenase-2 receptor, Cyclooxygenase-2, DNA polymerase kappa, Thrombin, Endoplasmic reticulum-associated amyloid beta-peptide-binding protein, Glutaminase kidney isoform, mitochondrial, MAP kinase ERK2, 15proteins, other nuclear proteins EKK2, 15-hydroxyprostaglandin dehydrogenase [NAD+], Isocitrate dehydrogenase [NADP] cytoplasmic, DNAapurinic or apyrimidinic site lyase, Aldehyde dehvdrogenase 1A1. Tvrosvl DNA phosphodiesterase 1 Caffeic acid, Chlorogenic acid, p Coumaric acid, Curcumin, Ferulic Apples, apricots, Blueberry, carrots, cereals citrus fruit, cherry, coffee Example- Caffeic acid: Enzyme, epigenetic regulator Example- Caffeic acid: Cerebroside-sulfatase, Carbonic anhydrase (I, II, III, V, VI, VII, IX, XII, XIV, VA, Example- Caffeic acid: Integrase (Human Example- Caffeic acid: 4'-phosphopantetheinyl transferase ffp (Bacillus Example- Caffeic acid: 1KOU, 207D, 3HOF, 3S2Z, 4EYQ, 4FB4, 4N0S, 4YU7, 5VFJ, Caffeic acid (Anticarcinogenic tivity [30], Anti beans, grape, kiwis, oil inflammatory [31], inclassified protein immunodeficiency subtilis), Endonuclease 6AWU, seeds, orange, peaches, plum, potato, spinach, acids. Anti-oxidant anscription factor VB). DNA polymerase beta. virus 1). (Escherichia coli K-12) 6172 Anti-oxidant activity [32], cosmetic use [33]); Chlorogenic acid (Antidiabetic and antiobesity activity ranscription factor other cytosolic protein, ion channel nembrane receptor transporter, other nuclear protein Hydroxycinammate: Hydroxycinnamic derivatives DNA polymerase kappa Neuraminidase eptyl-4(1H)-quinolon Influenza A virus) Large T antigen (Simian virus 40) synthase PqsD (Pseudomonas aeruginosa) rotein veet pear, spices, toma wheat bran [27 - 29] Protein-tyrosine phosphatase LC-PTP, Matrix metalloproteinase-1, Egl nine homolog 1, Protein-tyrosine homotog 1, Protein-tytosme phosphatase 2C, Tyrosyl-DNA phosphodiesterase 1, MAP kinase ERK2, Epidermal growth factor receptor erbB1, Acetylcholinesterase, DNA-(apurine or apyrimidinic site) [34], [34], antihypertensive [35], antioxidant and anti-inflammatory effect [36], antimicrobial effect Structure of Fragaria anana: [37], neuroprotective effect [38]; p-Coumarie acid [UV protective, hypopigmentation and anti-melanogenic effect [39], immuno-modulatory and anti-inflammatory activity [40], anti-platelet activity [41], Antidiabetic and antihyperlipidemic [37], O-methyltransferase in complex with S-adenosylhomocysteine and caffeic acid (PDB: 6172) lyase, Tyrosinase, Protein-Iyase, Fyrosinase, Protein-tyrosine phosphatase IB, Endoplasmic reticulum-associated amyloid beta-peptide-binding protein, Matrix metalloproteinase 9, Lysosomal alpha-glucosidas Aldose reductase, Matrix metalloproteinase-2, antihyperlipidemic activity [42]) Cyclooxygenase-1, Cyclooxygenase-2, Dipeptidyl peptidase IV, Aldo-keto reductase family 1 member (C1, C2, C3, C4, B10), Curcumin (Anti-HIV activity [43], anti-mutagenic and anti- carcinogenic [44], Antiaflatoxin (CI, C2, C3, C4, B10), Hyaluronidase-1, Xanthine dehydrogenase, DNA---methyladenine glycosylase, Alpha-galactosidase A, Aldehyde dehydrogenase 1A1 15 bedrowenare for dia and antifungal acti-vity [45] Anti-athero-sclerotic [46], Anti-angiogenic [47], Antioxidant 15-hydroxyprostaglandin dehydrogenase [NAD+], LDL-associated [48]. Anti-ischemi [49], Anti-ischerine [49], fibrinolytic [50], hepatoprotective [51], Ornithine phospholipase A2, Dual specificity protein phosphatase 3, Cytochrome P450 3A4, Arachidonate 5decarboxylase inhibitor activity lipoxygenase, Thrombin, Alanine aminotransferase 1 [52]. Protease hibitor action [53] Glutaminase kidney isoform Protein kinase hibitor action [54] mitochondrial,

Table 2. Food sources, biological and pharmaceutical activities of some phenolic compounds.

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Phenolic compound	Food source	Biological & Pharmaceutical activities	Target class	Human Target	Virus Target	Bacteria Target	PDB Entries & 3D Protein complex crystal structure
Flavonols: <u>Kaempferol</u> Isorhamnetin, Myricetin, Quercetin, Araleatin, Fischer Jalangin, Gossypetin, Pachypodol, Rhammazin, Rhammetin Hillwee dendive, fennel lettuce, olive, pears, pepper, plums, red wine, strawberries, tea, tomatoes and turnip.	Isorhannetin (Antimicrobial activity 155, 56], anti-diabetic effects [57], hepatoprotective activity [58], anti- inflammatory activity [59], anti- ocagulant activity [60], cardiovascular protection [61], anticancer activity [62], neurological effects [63]); Knempferol (Cardioprotective activity [66], anti-diabetic activity [66],	Example- Kaempferol: Enzyme, Transcription factor, unclassified protein, transporter, other cytosolic protein, epigenetic regulator, ion channel, membrane receptor, other nuclear protein, structural protein	Example- Kaempferol: Carbonic anhydrase (I, II, IV, VII, XII), DNA polymerase (eta, iota), Receptor-type tyrosine-protein phosphatase S, Cytochrome P450 (2D6, 2C9, 2C19, JA1, IA2, IB1, 3A4), Cyclin-dependent kinase 1/cyclin-dependent endopertidase, Tyrosyl-IDNA phosphodiesterase I, Beta- secretase I, Epidermal growth factor receptor erbB1, Serinet/Hroonine-protein kinase PIMI, Sialidase 2, Acetylcholinesterase, DNA- (apurinic or apyrimidinic sile) lyase, Myeloperoxidase, Estradiol 17-bet- -dehydrogenase (1, 2),	Example- Kaempferol: Avian myoblastosis virus polyprotein II, Reverse transcriptase, integrase (Human immunodeficiency virus 1), Neuraminidase (Influenza A virus- strain A/USSR/90/1977 H1N1; A/Puerto Rico/8/1934(H1N1))	Example- Kaempferol: Anthrax lethal factor (Bacillus anthracis), Sialdase (Clostridium perfringens), Oleandomycin glycosyltransferase (Streptomyces antibioticus)	Example- Kaempferol: <u>5AUX</u> SAV2, 4REL, 6MBB, 1H1M 2C1Z, 5AV3, 4DET, 3QWH Crystal structure of DAPK1 i complex with kaempferol (PDB: 5AUX)	
		neuroprotective activity [67]); Myricetin (antoxidant activity [68], antiphetoaging activity [69], anti- photoaging anticancer activity [70], anti-platelet antihypertensive activity [71], anti- inflammatory activity [72], anti- inflammatory activity [71], anti- andlergie activity [72], anti- activity [71], anti- activity [71], anti- cativity [71], anti- cativity [71], anti- cativities [77], anti- cativities [77], anti- cativities [77], anti- cativities [77], anti- cativities [77], anti- cativities [72], anti- cativities [73], anti- cativities [73], anti- cativities [73], anti- cativities [73], anti- cativities [73], anti- cativities [73], anti- cativity [68]). Quercifin (anti- obesity activities [74], antimerobial activity [85], cencer and apotasis [83], ulcer and gastritis [84], antimerobial activity [85], cencer and apotasis [83], ulcer and gastritis [84], antimerobial activity [85], cencer and apotasis [83], ulcer and gastritis [84], antimerobial activity [85], cencer and apotasis [83], ulcer and gastritis [84], antimerobial activity [85], cencer and apotasis [83], ulcer and gastritis [84], antimerobial activity [85], cencer and apotasis [83], ulcer and gastritis [84], antimerobial activity [85], cencer and apotasis [83], ulcer and gastritis [84], antimerobial activity [85], cencer and apotasis [83], ulcer and gastritis [84], antimerobial activity [85], cencer and apotasis [84], ulcer and gastritis [84], antimerobial activity [85], cencer and apotasis [85], ulcer and gastritis [85], cencer and gastritis [85], cencer and gastritis [85], cencer and gastritis [85], cencer and gastritis [85], cencer and		Salivary alpha-amylase, Superoxide dismutase, Tyrosinase, Death-associated protein kinase 1, Arachidonate 13-lipoxygenase, type II, Carboxy-terminal domain RNA polypeptide A small phosphatase 1, Endoplasmic reticulum-associated amyloid beta-peptide-inding protein, Ubiquitin carboxyl-terminal hydrolase 1, Aldose reductase, Glycogen synthase kinase 3, DNA topoisomerase II alpha, Casein kinase II, Cyclim-dependent kinase S/CDKS activator 1, CDK6, Dipeptidyl peptidase IV, Tyrosine-protein kinase receptor FL73, Xanhine dehydrogenase, Flap endonuclease 1, DNA -methyladenine glycosylase, NADPH oxidase 4, Beta- glucocerebroxidase, Aldehyle dehydrogenase, IS- hydroxyprostaglandin dehydrogenase [NADP], Isociritate dehydrogenase [NADP] (voplasnic, Butyrylcholinesterase, Serum paraoxonase/rylesterase 1, Giyoxalase I, Catalase			

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Phenolic compound	Food source	Biological & Pharmaceutical activities	Target class	Human Target	Virus Target	Bacteria Target	PDB Entries & 3D Protein complex crystal structure
Flavones: <u>Apigenin</u> , Luteolin, tangerin, chrysin, 6- hydroxyflavone)	Allium cepa, Allium sativa, artichoke, artemisia, achillea, citrus fruits, celery, chamomile tea, matricaria, olive, oregano, parsley, rosemary, spinach, sweet peppers, Tanacetum, thyme	Apigenia (As nutraceuticals [87], as anticance [88], as anticance [88], as anticance [88], as anticance [88], as anticance [88], as antioxidant and treatment of Alzheimer's disease (90), insomnia [91], knee otecarthritis edepression [93], anti-diabetic activity [94]); Luteolin (in the treatment of ageing, atherosclerosis, anti-inflammatory cardiovascular disease, diabetes mellitus, hypertension, ischemia/ reperfusion injury, neurodegenerative disordera- activity [98], antimirrobial activity [99], antioxidant activity [100])	Example-Apigenin: Enzyme, transcription factor, unclassified protein, transporter, other cytosolic protein, epigenetic regulator, ion channel, membrane receptor, secreted protein, others	Example- Apigenin: Cerebroside-sulfatase, Casein kinase I, Ubiquitin carboxyl- terminal hydrolase 2, DNA polymerase eda/ota, Serine/threonine-protein kinase (RKI, DNA polymerase kappa, CaM kinase I gamma, CaM kinase II gamma, Poly (ADP-ribose] polymerase-I. Cytochrome P450 (2D6, 2C9, 2C19, IA1), RAF2- and NCK-interacting kinase, Pyruvate kinase isozymes M1/M2, CaM kinase II delta, Death- associated protein kinase 3, Serine/threonine-protein kinase PMST, Cyclin- dependent kinase I/AgCI and kinase IPAKT, Tyrost-IDNA phosphodiestersse 1, Breast cancer type 1 susceptibility protein, MAP kinase ERK2, Serine/threonine-protein kinase PMKA, MP-activated protein kinase-IAK 2, Cyclin- dependent kinase-Ibke 1, Phosphodiestersse 5, MAP kinase PBA 2, Suba, Beta- secretase 1, Epidermal growth factor receptor erbb1, Serine/threonine-protein kinase PIM1, and other signalling proteins	Example- Apigenin: Integrase, Reverse transcriptase (HIV-1), Neuraminidase polyprotein Iab (SARS coronavirus)	Example- Apigenin 4'- phosphopantchieniy1 transferase ffb (Bacillus subilis), Sialdase (Clostridium perfrigens), D-alanylalanine synthetase, Endonuclease 4, DNA gyrase, (Escherichia colo K-12), Urease subunit alpaUrease subunit beta, D-alanine-D-alanine ligase (Helicobacter pylori strain ATCC 700392 / 26695/ strain HPAG1)	Example-Apigenin: 4WO0, <u>5AUV</u> , 512H, 4DGM, 4DER 4HKK, 3AMY, 3CP, 5UQT Crystal structure of DAPK1 i complex with apigenin (PDB 5AUV)
Flavands frue Flavanols: <u>Catechin</u> . Epicatechin gallate, Epigallocatechin gallate, Proanthocyanidins, Theaflavins, Theaflavins, Thearubigins	apples, apricots, barley, cereal, beer, chocolate, cocca, grapes and blackberrie, apples, peaches, nectarines, nuts, plums, red wine, sour cherries, and tea	For antimutagenic activity [101], antioxidant activity [102], antihypertensive activity [103], anti- protestive activity [104], cardio- protestive activity [105], anti- thrombogenic activity [106], hypolipidemic and antidiabetic activity [107], anti-allergic antidiabetic activity [107], anti-allergic [108], antibasterial and antiviral effect [109], anti-tumor effect [110]	Example-Catechin: Enzyme, unclassified protein, epigenetic regulator, ion channel, olicy protein, transcription factor	Example- Catechin: Carbonic anhydrase (I, III, III, IV, VI, VII, IX, XII, XIII, XIV, VA, VB), Lymphocyte differentiation antigen CD38, ATP-dependent DNA helicase (O, Ribounclease pancratic, DNA polymerase kappa, DNA polymerase kappa, DNA polymerase kappa, DNA polymerase kappa, DNA polymerase kappa, DNA polymerase kappa, DNA polymerase kappa, Manase ERK2, DNA-(appurine or apyrimidinic site) lyase, Salivary alpha-amylase, Valiquitin carboxyl-terminal hydrolase 1, Intestinal alkaline phosphatase, Aldehydrogenase IA1, Arachidonate 15–, 15-hydroxyprostaglandini dehydrogenase [NAD+], LDP-associated phospholipase A2, Serum paraoxonase/arylesterase 1, Alkaline phosphatase placental-like, Glutaminase kidney isoform, mitochondrial,	Example- Catechin Neuramindase (Influenza A virus), Large T antigen (Simian virus 40)	Example- Catechin 4 ¹ phosphopantetheinyl transferase fin (Bacillus subtilis), Urease subunit alphu/Ureas exubunit beta (Helicobacter pylori strain ATCC 700392 / 26695)	Example- Catechin: 3152, 4C94, 4C91

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Phenolic compound	Food source	Biological & Pharmaceutical activities	Target class	Human Target	Virus Target	Bacteria Target	PDB Entries & 3D Protein complex crystal structure
Flavanones: Hesperein, Eriodictyol, Naringenin, Sakurametin, Blumeatin, Butin, Hesperidin, Isosakuranetin, Isosakuranetin, Naringin, Pinocembrin, Pinocembrin, Pinocembrin, Pinotrobin	Citrus fruits: grapes, lemons, orange, and tomatoes (Naringenin)	Naringenin (anticancer and antiatherogenic properties [111], prevention of 7] cardioprotective activity [113], Antimutagenic activity [114], anti- inflammatory activity [114], anti- fibrotic activity [117], antimicrobial activity [118], anti- diabetic activity [117], antimicrobial activity [118], anti- diabetic activity [117], antimicrobial activity [118], anti- diabetic activity [117], antimicrobial activity [118], anti- diabetic activity [117], antimicrobial activity [120], hypolipidemic activity [121]); Eriodictoyl (for the treatment of cancer [122], anti- inflammatory properties [123], antioxidant activity [124], cardioprotective properties [127]); Hesperetin (antioxidant activity [128], antioxidant activity [130], antibacterial activity [134]); sakuranetin (antiacancer properties [137], anti-mutagenic properties [137], anti-mutagenic properties [137], anti-mutagenic properties [137], anti-mutagenic properties [137], anti-mitagenic properties xample- Hesperein: Enzyme, unclassified protein, epigenetic regulator, transcription factor, membrane receptor, transporter	Example-Hesperein: Carhonic anhydrase (I, II, VV VII, XII), Ras-related protein Rab-9A, Cytochrome P450 (2D6, 2C9, 2C19, 1A1, 1A2, IB1, 3A4), TyrosyI-DNA phosphodiesterase I, MAP kinase ERK2, c-Jun N- terminal kinase (1, 2), Acetylcholinesterase, Estradiol 17-bet- dehydrogenase, Endoplasmic reticulum-associated amyloid beta-peptide-binding protein, Serine-protein kinase ATM, Dipeptidy1 peptidase IV, Glycogen synthase kinase- dehydrogenase 2, Aldehyde dehydrogenase 2, Aldehyde dehydrogenase 2, Aldehyde dehydrogenase 2, Aldehyde dehydrogenase 2, Aldehyde JO, Thrombin, TGF-beta receptor type II,	Example- Hesperetin: Replicase polyprotein lab (SARS coronavirus)	Example-Hespereini Beta-lactamase AmpC (Escherichia coli K-12)	Example- Hesperetin: SJDC	

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(<i>Table 2</i>) <i>cont</i> Phenolic compound	Food source	Biological & Pharmaceutical activities	Target class	Human Target	Virus Target	Bacteria Target	PDB Entries & 3D Protein complex crystal structure
Anthocyanidins (Cvanidin, Delphinidin, Malvidin, Pelargonidin, Peonidin, Capensinidin, Europinidin, Hirsutidin, Petunidin, Pulchellidin, Rosinidin)	Anthocyanidin present in fruits is cyanidin⊡ glucoside, Grapes, blueberry, red onions, oranges, and red wine. Blackcurrant, blackberry, and elderberry (only cyanide), present in epidermal tissues of fruits and flower	Cyanidin (anticarcinogenic activity [140], vasoprotective nature [141], anti- inflammatory int-obesity [143], anti-obesity [143], anti-abesity [143], anti-abetic activity [144]); Detphinidin (anti- angiogenic activity [147], anti- inflammatory activity [148], anti- difficut activity [147], anti- inflammatory activity [148], anti- difficut activity [149], anti- difficut activity [147], anti- inflammatory activity [153], anti-inflammatory activity [153], anti-septic effect [155], anti-septic effect [155], anti-tumour activity [156], antiseptic properties [159], anti-diabeta cactivity [158], antiseptic properties [159], anti-diabeta cactivity [161], anti- inflammatory activity [162], anti- tumour activity [161], anti- inflammatory activity [162], anti- tumour activity [163], anti- tumour activity [163], anti- tumour activity [164], activity [162], anti- tumour activity [163], anti- mutagenic activity [164], activity [164], anti-tumour activity [164], anti-tumour activity [164], anti-tumour activity [164], activity [164], activity [165], activity [164], activity [164], activity [165], activity [164], activity [164], activity [165], activity ample- Cyanidin: Enzyme, transcription factor	Example- Cyanidin: Lymphocyte differentiation antigen CD38, Death- associated protein kinase 1, Cyclooxygenase-1, Dipeptidyl peptidase IV, Cyclooxygenase-2, Glyoxalase I, Thrombin	Not found	Not found	Example- Cyanidin: 60CH	
Isoflavones (<u>Daidzein</u> , Genistin, Glycitein)	Soybeans and soy products are almost the sole dietary source of isoflavones. It is also in small amounts in chickpeas.	Daidzein (antihrombotic and anti-allergie activity [165], anticancer activity [166], anti- arthritogenic and cardioprotective action [167], anti- osteoprorsis activity [169], anti-diabetic activity [168], anti- osteoprorsis activity [172], estrogenic anti-hypoxia activity [173]).	Example-Daidzin: Enzyme, unclassified protein, epigenetic regulator, ion channel, olther cytosolic protein, transcription factor, transporter, membrane receptor, other nuclear protein structural protein	Example- Daidzein: Cerebrosids-sulfatase, Carbonic anhydrase (J, II, IV, VII, XII), Ras-related protein Rab-9A, Bloom syndrome protein, Cytochrome P450 (20b, C2O, 2019, 1A2, 19A4), 20b, C2O, 2019, 1A2, 19A4), 20b, 20b, 2019, 2019, 1A2, 19A4, 2019	Example- Daidzin: Neuramindase (Influenza A virus), Replicase polyprotein Iab (SARS coronavirus)	Example- Daidzein: Urease subunit alpha/Urease subunit beta (Helicobacter pylori tstain ATCC 00392 / 26695), Oleandomycin glycosyltransferase (Streptomyces antibioticus)	No PDB ID found Molecular interactions found with human genes: TRPC5 (transient receptor potential cation channel, subfamily C, member 5), ESRRB (estrogen related receptor beta), IBSP (integrin binding sialoprotein) PIK3CG (Phoephandi/hinositi 4,5-bisphosphate) 3-kinase catalytic subunit gamma isoform), ESRRA (estrogen- related receptor alpha), LIF (interleukin 6 family cytokine FOS (fos proto-oncogene, ap- transcription factor subunit)

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Phenolic compound	Food source	Biological & Pharmaceutical activities	Target class	Human Target	Virus Target	Bacteria Target	PDB Entries & 3D Protein complex crystal structure
Stilbenes (<u>Reveratrol</u> , Pinosylvin, Pterostilbene)	Found in berries, grapes, plums and pine nuts, peanuts, spinach, red wine, and red cabbage.	Resveratrol (antioxidant activity (174), anticancer activity [175], neuroprotective activity [176], anti- cardioprotective activity [177], anti- diabetic effect [178], non-al-coholic fatty (180), anti- anti-allergic activity (180), anti- inflammatory activity [181], immunomodulity [182]).	Example- Resveratrol: Enzyme, itranscription factor, unclassified protein, opigenetic regulator, transporter, ion channel, other cytosolic protein, secreted protein, others	Example- Resveratrol: Cerebroside-sulfatase, FAD- linked sulfydryl oxidase ALR, Carbonic anhydrase (I, IV, VI, VII, VA, VB, IX, XII, XIV), Lymphocyte differentiation antigen CD38, DNA polymerase kappa, DNA polymerase kappa, DNA phosphodiesterase 1, Breast cancer type 1 susceptibility protein, MAP kinase FRK2, Phosphodiesterase 5, MAD kinase p38 alpha, Beta- secretase 1, Epidermal growth factor receptor erbB1, Leukocyte elastase, Monoamine oxidase B, Ber/Abl fusion protein, etc.	Example- Resveratroi: Large T antigen (Simian virus 40)	Example- Resveratrol: ATP-dependent CIp protease proteolytic subunit (Bacillus subtilis strain 168), Beta- lactamase AmpC (Escherichia coli K-12)	Example-Resveration: 40ER 1S60, 5CR1, 4QOH, ICGZ, 4QOJ, 5JS4, 5U90, IDVS, 1U0W, 3CKL, 4Q93, 3MNQ, 4PP6, 2IIZ, 5JSTS, SNZL, 4DPN, 6JEM, 4HDA, 2VDX, 4JAZ, IZIF, 5BTR, 2L98 Crystal Structure of the Complex of Phospholipase A2 with Resveration 1 ± 20 Å Resolution (PDB: 4QER)
Stilhenol (<u>Piccataanol</u> Astringm)	Found in grapes, fruit, white tea and red wines	Picetannol (anti- inflammatory activity [183], anti- cancer activity [184], anti-atheregenic activity [185], anti- oxidant, neuroprotective and anti-allergic activity [186], amticariogene activity [187], anti- platele aggregation activity [188], Astringi (antioxidant activity [189], anti- inflammatory activity [190]).	Example- Piceatannol: Enzyme, inn, epigenetic regulator, ion channel, other cytosolic protein, cytosolic protein, transcription factor	Example- Piceatamol: PIK3CG (Phosphatidylinositol 4,5-hisphosphate 3-kinase catalytic subunit gamma isoform), CYP1A2 (Cytochrome P450 1A2), ATP synthase subunit alpha, mitochondrial,	Not found	Not found	Example-Picetannol: 5097, 4HD8, 2011
Lignan (Medioresinol, Matairesinol, Lariciresinol, Lariciresinol, Secoisdariciresinol, Hydroxymatairesinol, Syringaresinol, Sesamin)	Flaxseed is the richest source Buckwheat, sesame seed, rye, and wheat	Lignans (anti- tumour activity [191], anti-mitotic activity [192], anti-mitotic activity [192], antic insecticidal activity [194], antioxidant activity [195], antidiabetic activity [196], antiallergic activity [198], hypolipidemic activity [198], hypolipidemic activity [198], esterogenic activity [190], (200)	Example- Pinoresinol: unclassified protein, transcription factor	Example- Pinoresinol: Peroxisome proliferator- activated receptor (delta, alpha, gamma); Antineuroinflammatory activity (in mouse BV2 cells), Cytotoxicity against human PANCI cells,	Not found	Not found	Not found

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Phenolic compound	Food source	Biological & Pharmaceutical activities	Target class	Human Target	Virus Target	Bacteria Target	PDB Entries & 3D Protein complex crystal structure
Tamins'tamoids (Condensed tamin: Catechin, Epicatechin; Hydrolyzable tamins-Gallie acid; Ellagitamins: Punicalagin, Casuarictin)	Lentils, pear, grapes, peaches, plums, mangosteens, pears, red and white wine and apple juice Strawberries, blackberries, raspberries, walnuts, pecans, pomegranate bark, leaf and the fruit husk	Antiuleer activity [201], antimicrobial activity [202], anti- viral activity [202], anti- viral activity [203], histamine release inhibition [205], cytotoxic effect, anti-diabetic activities [206], anti-diabetic activities [207], anti-obesity effect [208], protecting effects on bone marrow hematopoietic stem cell [209], wound healing [210], inhibition of skin- tumor promotion [211].	Example- Catechin: Enzyme, unclassified protein, epigenetic regulator, ion channel, objective cytosolic protein, transcription factor	Example- Catechin: Carbonic anhydrase (I, III, IIV, VI, VII, IX, XII, XIIV, VA, VB), Lymphocyte differentiation antigen CD38, ATP-dependent DNA helicase QI, Ribouelcase pancratic, DNA polymerase icapa antigenetic antigenetic antigenetic phosphodiesterase 1, IAPP kinase ERK2, DNA-(apurinic or apyrimidinic site) jusae, Salivara ylaha-amylase, Ubiquitin carboxyl-terminal hydrolase 1, Intestinal alkalime phosphatase, Aldehyde dehydrogenase IAI, Arachidonate 15–, 15-hydroxyporstaglandin dehydrogenase [NAP-], UDP-glucuronosyltransforase, I-1, Alkaline phosphatase, 1-1, Alkaline phosphatase hydrose A2, Serum paraoxonase/arylesterase I, Makine phosphatase kidney isoform, mitochondrial	Example- Catechin: Neuramindase (Influenza A virus), Large T antigen (Simian virus 40)	Example- Catechin: 4- phosphopanteheinyl transferase ffp (Bacillus subtilis), Urease subunit alpha/Urease subunit bat (Helicobacter pylori strain ATCC 700392 / 26695)	Example: Catechin: 3152, 4C94, 4C91 Crystal Structure of the Strawberry Pathogenesis- Related 10 (PR-10) Fra a 3 protein in complex with Catechin (PDB: 4C94)

CONCLUSION

Oxidative stress and accumulation of free radicals play an essential role for the development of life threatening conditions such as cancer, cardiovascular complications, and neurodegeneration. The quest for efficient prevention has become the first concern for clinical science. Polyphenols have been considered as alternative therapeutics and have shown to be effective in various treatments, especially in cancer, neurodegeneration, and cardiovascular complications. It has already been well demonstrated that phenolics are able to exert protective effects on the heart, brain, tumor cells, and other organs. Their effects can be due to antioxidant properties, and they also interact with the basic cellular mechanisms. Polyphenols, such as curcumin, resveratrol, quercetin, and many others, have been shown to promote apoptosis induction in different types of cancers (e.g., skin, breast, lung, colon, prostate, melanoma, or leukemia). They also act as suppressing agents and inhibit the formation of tumors from initiated cells. However, the exact mechanisms of actions are not fully understood to date, require further consideration. Polyphenols are the potential candidates of natural origin in pharmaceutical sectors to promote human health, prevent and cure various diseases. However, intense research and profiling are required for the characterization of each compound on the basis of its mechanism of action.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The authors declared that there is no conflict of interest.

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REFERENCES

- Whiting DA. Natural phenolic compounds 1900-2000: a bird's eye view of a century's chemistry. Nat Prod Rep 2001; 18(6): 583-606.
 [http://dx.doi.org/10.1039/b003686m] [PMID: 11820759]
- [2] Oksana S, Marian B, Mahendra R, Bo SH. Plant phenolic compounds for food, pharmaceutical and cosmetics production. J Med Plants Res 2012; 6(13): 2526-39.
- [3] Harborne J. Secondary Plant Products, Bell, EA and Charlwood, BW [Eds], Enciclopedia of Plant Physiology, New Series, Berlin: Springer-Verlag 1980; 8.
- [4] Aoki T, Akashi T, Ayabe S-i. Flavonoids of leguminous plants: structure, biological activity, and biosynthesis. J Plant Res 2000; 113(4): 475.
 [http://dx.doi.org/10.1007/PL00013958]
- [5] Laura A, Alvarez-Parrilla E, Gonzalez-Aguilar GA. Fruit and vegetable phytochemicals: chemistry, nutritional value and stability. John Wiley & Sons 2009.
- [6] Yunusa AK, Rohin MAK, Bakar CAA. Free radical scavenging activity of polyphenols. J Chem Pharm Res 2015; 7(3): 1975-80.
- [7] Aquilano K, Baldelli S, Rotilio G, Ciriolo MR. Role of nitric oxide synthases in Parkinson's disease: a review on the antioxidant and anti-inflammatory activity of polyphenols. Neurochem Res 2008; 33(12): 2416-26.
 [http://dx.doi.org/10.1007/s11064-008-9697-6] [PMID: 18415676]
- [8] Mandel S, Youdim MB. Catechin polyphenols: neurodegeneration and neuroprotection in neurodegenerative diseases. Free Radic Biol Med 2004; 37(3): 304-17. [http://dx.doi.org/10.1016/j.freeradbiomed.2004.04.012] [PMID: 15223064]
- [9] Apostolou A, Stagos D, Galitsiou E, et al. Assessment of polyphenolic content, antioxidant activity, protection against ROS-induced DNA damage and anticancer activity of Vitis vinifera stem extracts. Food Chem Toxicol 2013; 61: 60-8. [http://dx.doi.org/10.1016/j.fct.2013.01.029] [PMID: 23380202]
- [10] Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: an overview. The Scientific World Journal 2013.
 [http://dx.doi.org/10.1155/2013/162750]
- [11] Ahmed SI, Hayat MQ, Tahir M, et al. Pharmacologically active flavonoids from the anticancer, antioxidant and antimicrobial extracts of Cassia angustifolia Vahl. BMC Complement Altern Med 2016; 16(1): 460. [http://dx.doi.org/10.1186/s12906-016-1443-z] [PMID: 27835979]
- [12] Valdés L, Cuervo A, Salazar N, Ruas-Madiedo P, Gueimonde M, González S. The relationship between phenolic compounds from diet and microbiota: impact on human health. Food Funct 2015; 6(8): 2424-39.

[http://dx.doi.org/10.1039/C5FO00322A] [PMID: 26068710]

- Kumar A, Khan F, Saikia D. Exploration of medicinal plants as sources of novel anticandidal drugs. Curr Top Med Chem 2019; 19(28): 2579-92.
 [http://dx.doi.org/10.2174/1568026619666191025155856] [PMID: 31654513]
- [14] Daglia M, Di Lorenzo A, Nabavi SF, Talas ZS, Nabavi SM. Polyphenols: well beyond the antioxidant capacity: gallic acid and related compounds as neuroprotective agents: you are what you eat! Curr Pharm Biotechnol 2014; 15(4): 362-72.
 [http://dx.doi.org/10.2174/138920101504140825120737] [PMID: 24938889]
- [15] Basli A, Belkacem N, Amrani I. Health benefits of phenolic compounds against cancers. Phenolic compounds–Biological activity London. UK: IntechOpen 2017; pp. 193-210.
- [16] Choi K-C, Lee Y-H, Jung MG, *et al.* Gallic acid suppresses lipopolysaccharide-induced nuclear factor-kappaB signaling by preventing RelA acetylation in A549 lung cancer cells. Mol Cancer Res 2009; 7(12): 2011-21.
 [http://dx.doi.org/10.1158/1541-7786.MCR-09-0239] [PMID: 19996305]
- [17] Shaik AH, Rasool SN, Vikram Kumar Reddy A, Abdul Kareem M, Saayi Krushna G, Lakshmi Devi K. Cardioprotective effect of HPLC standardized ethanolic extract of Terminalia pallida fruits against isoproterenol-induced myocardial infarction in albino rats. J Ethnopharmacol 2012; 141(1): 33-40. [http://dx.doi.org/10.1016/j.jep.2012.01.011] [PMID: 22366678]
- [18] Stanely Mainzen Prince P, Priscilla H, Devika PT. Gallic acid prevents lysosomal damage in isoproterenol induced cardiotoxicity in Wistar rats. Eur J Pharmacol 2009; 615(1-3): 139-43. [http://dx.doi.org/10.1016/j.ejphar.2009.05.003] [PMID: 19450577]
- [19] Badavi M, Sadeghi N, Dianat M, Samarbafzadeh A. Effects of gallic Acid and cyclosporine a on antioxidant capacity and cardiac markers of rat isolated heart after ischemia/reperfusion. Iran Red Crescent Med J 2014; 16(6)e16424 [6]. [http://dx.doi.org/10.5812/ircmj.16424] [PMID: 25068044]
- [20] Anand KK, Singh B, Saxena AK, Chandan BK, Gupta VN, Bhardwaj V. 3,4,5-Trihydroxy benzoic acid (gallic acid), the hepatoprotective principle in the fruits of Terminalia belerica-bioassay guided activity. Pharmacol Res 1997; 36(4): 315-21. [http://dx.doi.org/10.1006/phrs.1997.0236] [PMID: 9425622]
- [21] Nair GG, Nair CKK. Radioprotective effects of gallic acid in mice. BioMed research international 2013. [http://dx.doi.org/10.1155/2013/953079]
- [22] Kanbak G, Canbek M, Oğlakçı A, *et al.* Preventive role of gallic acid on alcohol dependent and cysteine protease-mediated pancreas injury. Mol Biol Rep 2012; 39(12): 10249-55. [http://dx.doi.org/10.1007/s11033-012-1901-8] [PMID: 23053933]
- [23] Yousuf MJ, Vellaichamy E. Protective activity of gallic acid against glyoxal -induced renal fibrosis in experimental rats. Toxicol Rep 2015; 2: 1246-54. [http://dx.doi.org/10.1016/j.toxrep.2015.07.007] [PMID: 28962467]
- [24] Hwang E, Park SY, Lee HJ, Lee TY, Sun ZW, Yi TH. Gallic acid regulates skin photoaging in UVBexposed fibroblast and hairless mice. Phytother Res 2014; 28(12): 1778-88. [http://dx.doi.org/10.1002/ptr.5198] [PMID: 25131997]
- [25] Nikbakht J, Hemmati AA, Arzi A, Mansouri MT, Rezaie A, Ghafourian M. Protective effect of gallic acid against bleomycin-induced pulmonary fibrosis in rats. Pharmacol Rep 2015; 67(6): 1061-7. [http://dx.doi.org/10.1016/j.pharep.2015.03.012] [PMID: 26481523]
- [26] Zhang Q, Chen W, Zhao J, Xi W. Functional constituents and antioxidant activities of eight Chinese native goji genotypes. Food Chem 2016; 200: 230-6. [http://dx.doi.org/10.1016/j.foodchem.2016.01.046] [PMID: 26830583]
- [27] Jaganath IB, Crozier A. Plant phenolics and human health: Biochemistry, nutrition and pharmacology.

Dietary flavonoids and phenolic compounds 2010; 1-39.

- [28] King A, Young G. Characteristics and occurrence of phenolic phytochemicals. J Am Diet Assoc 1999; 99(2): 213-8.
 [http://dx.doi.org/10.1016/S0002-8223(99)00051-6] [PMID: 9972191]
- [29] Naczk M, Shahidi F. Phenolics in cereals, fruits and vegetables: occurrence, extraction and analysis. J Pharm Biomed Anal 2006; 41(5): 1523-42. [http://dx.doi.org/10.1016/j.jpba.2006.04.002] [PMID: 16753277]
- [30] Damasceno SS, Dantas BB, Ribeiro-Filho J, Antônio M Araújo D, Galberto M da Costa J. Chemical properties of caffeic and ferulic acids in biological system: implications in cancer therapy. A review. Curr Pharm Des 2017; 23(20): 3015-23. [http://dx.doi.org/10.2174/1381612822666161208145508] [PMID: 27928956]
- [31] da Cunha FM, Duma D, Assreuy J, et al. Caffeic acid derivatives: in vitro and in vivo antiinflammatory properties. Free Radic Res 2004; 38(11): 1241-53. [http://dx.doi.org/10.1080/10715760400016139] [PMID: 15621702]
- [32] Sidoryk K, Jaromin A, Filipczak N, Cmoch P, Cybulski M. Synthesis and antioxidant activity of caffeic acid derivatives. Molecules 2018; 23(9): 2199. [http://dx.doi.org/10.3390/molecules23092199] [PMID: 30200272]
- [33] Magnani C, Isaac VLB, Correa MA, Salgado HRN. Caffeic acid: a review of its potential use in medications and cosmetics. Anal Methods 2014; 6(10): 3203-10. [http://dx.doi.org/10.1039/C3AY41807C]
- [34] Meng S, Cao J, Feng Q, Peng J, Hu Y. Roles of chlorogenic acid on regulating glucose and lipids metabolism: a review. Evidence-Based Complementary and Alternative Medicine 2013. [http://dx.doi.org/10.1155/2013/801457]
- [35] Kozuma K, Tsuchiya S, Kohori J, Hase T, Tokimitsu I. Antihypertensive effect of green coffee bean extract on mildly hypertensive subjects. Hypertens Res 2005; 28(9): 711-8. [http://dx.doi.org/10.1291/hypres.28.711] [PMID: 16419643]
- [36] Bao L, Li J, Zha D, *et al.* Chlorogenic acid prevents diabetic nephropathy by inhibiting oxidative stress and inflammation through modulation of the Nrf2/HO-1 and NF-κB pathways. Int Immunopharmacol 2018; 54: 245-53. [http://dx.doi.org/10.1016/j.intimp.2017.11.021] [PMID: 29161661]
- [37] Li G, Wang X, Xu Y, Zhang B, Xia X. Antimicrobial effect and mode of action of chlorogenic acid on Staphylococcus aureus. Eur Food Res Technol 2014; 238(4): 589-96. [http://dx.doi.org/10.1007/s00217-013-2140-5]
- [38] Rebai O, Belkhir M, Sanchez-Gomez MV, Matute C, Fattouch S, Amri M. Differential molecular targets for neuroprotective effect of chlorogenic acid and its related compounds against glutamate induced excitotoxicity and oxidative stress in rat cortical neurons. Neurochem Res 2017; 42(12): 3559-72.

[http://dx.doi.org/10.1007/s11064-017-2403-9] [PMID: 28948515]

- [39] Boo YC. p-Coumaric acid as an active ingredient in cosmetics: A review focusing on its antimelanogenic effects. Antioxidants 2019; 8(8): 275. [http://dx.doi.org/10.3390/antiox8080275] [PMID: 31382682]
- [40] Pragasam SJ, Venkatesan V, Rasool M. Immunomodulatory and anti-inflammatory effect of p-coumaric acid, a common dietary polyphenol on experimental inflammation in rats. Inflammation 2013; 36(1): 169-76.
 [http://dx.doi.org/10.1007/s10753-012-9532-8] [PMID: 22923003]
- [41] Luceri C, Giannini L, Lodovici M, et al. p-Coumaric acid, a common dietary phenol, inhibits platelet activity in vitro and in vivo. Br J Nutr 2007; 97(3): 458-63. [http://dx.doi.org/10.1017/S0007114507657882] [PMID: 17313706]

- [42] Amalan V, Vijayakumar N, Indumathi D, Ramakrishnan A. Antidiabetic and antihyperlipidemic activity of p-coumaric acid in diabetic rats, role of pancreatic GLUT 2: In vivo approach. Biomed Pharmacother 2016; 84: 230-6. [http://dx.doi.org/10.1016/j.biopha.2016.09.039] [PMID: 27662473]
- [43] Mazumder A, Raghavan K, Weinstein J, Kohn KW, Pommier Y. Inhibition of human immunodeficiency virus type-1 integrase by curcumin. Biochem Pharmacol 1995; 49(8): 1165-70. [http://dx.doi.org/10.1016/0006-2952(95)98514-A] [PMID: 7748198]
- [44] Anto RJ, George J, Babu KD, Rajasekharan K, Kuttan R. Antimutagenic and anticarcinogenic activity of natural and synthetic curcuminoids. Mutation Research/Genetic Toxicology 1996; 370(2): 127-31. [http://dx.doi.org/10.1016/0165-1218(96)00074-2]
- [45] Hu Y, Zhang J, Kong W, Zhao G, Yang M. Mechanisms of antifungal and anti-aflatoxigenic properties of essential oil derived from turmeric (Curcuma longa L.) on Aspergillus flavus. Food Chem 2017; 220: 1-8. [http://dx.doi.org/10.1016/j.foodchem.2016.09.179] [PMID: 27855875]
- [46] Jin S, Hong J-H, Jung S-H, Cho K-H. Turmeric and laurel aqueous extracts exhibit *in vitro* antiatherosclerotic activity and in vivo hypolipidemic effects in a zebrafish model. J Med Food 2011; 14(3): 247-56.
 [http://dx.doi.org/10.1089/jmf.2009.1389] [PMID: 21332404]
- [47] Shakeri A, Ward N, Panahi Y, Sahebkar A. Anti-angiogenic activity of curcumin in cancer therapy: a narrative review. Curr Vasc Pharmacol 2019; 17(3): 262-9. [http://dx.doi.org/10.2174/1570161116666180209113014] [PMID: 29424316]
- [48] Yang M, Wu Y, Li J, Zhou H, Wang X. Binding of curcumin with bovine serum albumin in the presence of t-carrageenan and implications on the stability and antioxidant activity of curcumin. J Agric Food Chem 2013; 61(29): 7150-5. [http://dx.doi.org/10.1021/jf401827x] [PMID: 23819626]
- Shukla PK, Khanna VK, Ali MM, Khan MY, Srimal RC. Anti-ischemic effect of curcumin in rat brain. Neurochem Res 2008; 33(6): 1036-43.
 [http://dx.doi.org/10.1007/s11064-007-9547-y] [PMID: 18204970]
- [50] Madhyastha R, Madhyastha H, Nakajima Y, Omura S, Maruyama M. Curcumin facilitates fibrinolysis and cellular migration during wound healing by modulating urokinase plasminogen activator expression. Pathophysiol Haemost Thromb 2010; 37(2-4): 59-66. [http://dx.doi.org/10.1159/000321375] [PMID: 21071923]
- [51] Girish C, Koner BC, Jayanthi S, Ramachandra Rao K, Rajesh B, Pradhan SC. Hepatoprotective activity of picroliv, curcumin and ellagic acid compared to silymarin on paracetamol induced liver toxicity in mice. Fundam Clin Pharmacol 2009; 23(6): 735-45. [http://dx.doi.org/10.1111/j.1472-8206.2009.00722.x] [PMID: 19656205]
- [52] Rao CV, Simi B, Reddy BS. Inhibition by dietary curcumin of azoxymethane-induced ornithine decarboxylase, tyrosine protein kinase, arachidonic acid metabolism and aberrant crypt foci formation in the rat colon. Carcinogenesis 1993; 14(11): 2219-25. [http://dx.doi.org/10.1093/carcin/14.11.2219] [PMID: 8242846]
- [53] Chai H, Yan S, Lin P, Lumsden AB, Yao Q, Chen C. Curcumin blocks HIV protease inhibitor ritonavir-induced vascular dysfunction in porcine coronary arteries. J Am Coll Surg 2005; 200(6): 820-30.
 [http://dx.doi.org/10.1016/j.jamcollsurg.2005.02.030] [PMID: 15922191]
- [54] Liu J-Y, Lin S-J, Lin J-K. Inhibitory effects of curcumin on protein kinase C activity induced by 12-tetradecanoyl-phorbol-13-acetate in NIH 3T3 cells. Carcinogenesis 1993; 14(5): 857-61. [http://dx.doi.org/10.1093/carcin/14.5.857] [PMID: 8504477]
- [55] Jnawali HN, Jeon D, Jeong M-C, et al. Antituberculosis activity of a naturally occurring flavonoid,

isorhamnetin. J Nat Prod 2016; 79(4): 961-9. [http://dx.doi.org/10.1021/acs.jnatprod.5b01033] [PMID: 26974691]

- [56] Abdal Dayem A, Choi HY, Kim YB, Cho S-G. Antiviral effect of methylated flavonol isorhamnetin against influenza. PLoS One 2015; 10(3)e0121610 [http://dx.doi.org/10.1371/journal.pone.0121610] [PMID: 25806943]
- [57] Lee YS, Lee S, Lee HS, Kim B-K, Ohuchi K, Shin KH. Inhibitory effects of isorhamnetin-3-Oβ-D-glucoside from Salicornia herbacea on rat lens aldose reductase and sorbitol accumulation in streptozotocin-induced diabetic rat tissues. Biol Pharm Bull 2005; 28(5): 916-8. [http://dx.doi.org/10.1248/bpb.28.916] [PMID: 15863906]
- [58] Kim D-W, Cho H-I, Kim K-M, et al. Isorhamnetin-3-O-galactoside protects against CCl4-induced hepatic injury in mice. Biomol Ther (Seoul) 2012; 20(4): 406-12. [http://dx.doi.org/10.4062/biomolther.2012.20.4.406] [PMID: 24009828]
- [59] Kim TH, Ku SK, Bae JS. Anti-inflammatory activities of isorhamnetin-3-O-galactoside against HMGB1-induced inflammatory responses in both HUVECs and CLP-induced septic mice. J Cell Biochem 2013; 114(2): 336-45. [http://dx.doi.org/10.1002/jcb.24361] [PMID: 22930571]
- [60] Ku S-K, Kim TH, Bae J-S. Anticoagulant activities of persicarin and isorhamnetin. Vascul Pharmacol 2013; 58(4): 272-9.
 [http://dx.doi.org/10.1016/j.vph.2013.01.005] [PMID: 23391847]
- [61] Huang L, He H, Liu Z, Liu D, Yin D, He M. Protective effects of isorhamnetin on cardiomyocytes against anoxia/reoxygenation-induced injury is mediated by SIRT1. J Cardiovasc Pharmacol 2016; 67 (6): 526-37.
 [http://dx.doi.org/10.1097/FJC.0000000000376] [PMID: 26859194]
- [62] Saud SM, Young MR, Jones-Hall YL, *et al.* Chemopreventive activity of plant flavonoid isorhamnetin in colorectal cancer is mediated by oncogenic Src and β-catenin. Cancer Res 2013; 73(17): 5473-84. [http://dx.doi.org/10.1158/0008-5472.CAN-13-0525] [PMID: 23824743]
- [63] Zhao J-J, Song J-Q, Pan S-Y, Wang K. Treatment with isorhamnetin protects the brain against ischemic injury in mice. Neurochem Res 2016; 41(8): 1939-48. [http://dx.doi.org/10.1007/s11064-016-1904-2] [PMID: 27161367]
- [64] Suchal K, Malik S, Khan SI, et al. Molecular pathways involved in the amelioration of myocardial injury in diabetic rats by kaempferol. Int J Mol Sci 2017; 18(5): 1001. [http://dx.doi.org/10.3390/ijms18051001] [PMID: 28505121]
- [65] Arif H, Sohail A, Farhan M, Rehman AA, Ahmad A, Hadi SM. Flavonoids-induced redox cycling of copper ions leads to generation of reactive oxygen species: A potential role in cancer chemoprevention. Int J Biol Macromol 2018; 106: 569-78. [http://dx.doi.org/10.1016/j.ijbiomac.2017.08.049] [PMID: 28834706]
- [66] Li F, Zhang B, Chen G, Fu X. The novel contributors of anti-diabetic potential in mulberry polyphenols revealed by UHPLC-HR-ESI-TOF-MS/MS. Food Res Int 2017; 100(Pt 1): 873-84. [http://dx.doi.org/10.1016/j.foodres.2017.06.052] [PMID: 28873762]
- [67] Wang L, Tu Y-C, Lian T-W, Hung J-T, Yen J-H, Wu M-J. Distinctive antioxidant and antiinflammatory effects of flavonols. J Agric Food Chem 2006; 54(26): 9798-804. [http://dx.doi.org/10.1021/jf0620719] [PMID: 17177504]
- [68] Semwal DK, Semwal RB, Combrinck S, Viljoen A. Myricetin: A dietary molecule with diverse biological activities. Nutrients 2016; 8(2): 90. [http://dx.doi.org/10.3390/nu8020090] [PMID: 26891321]
- [69] Jung SK, Lee KW, Kim HY, et al. Myricetin suppresses UVB-induced wrinkle formation and MMP-9 expression by inhibiting Raf. Biochem Pharmacol 2010; 79(10): 1455-61. [http://dx.doi.org/10.1016/j.bcp.2010.01.004] [PMID: 20093107]

- [70] Siegelin MD, Gaiser T, Habel A, Siegelin Y. Myricetin sensitizes malignant glioma cells to TRAIL-mediated apoptosis by down-regulation of the short isoform of FLIP and bcl-2. Cancer Lett 2009; 283 (2): 230-8.
 [http://dx.doi.org/10.1016/j.canlet.2009.04.002] [PMID: 19398149]
- [71] Zang BX, Jin M, Wu W, Chen WM, Piao YZ, Li JR. [Antagonistic effect of myricetin on platelet activing factor]. Yao Xue Xue Bao 2003; 38(11): 831-3. [PMID: 14991995]
- Borde P, Mohan M, Kasture S. Effect of myricetin on deoxycorticosterone acetate (DOCA)-sal- -hypertensive rats. Nat Prod Res 2011; 25(16): 1549-59. [http://dx.doi.org/10.1080/14786410903335190] [PMID: 21391110]
- [73] Kang BY, Kim SH, Cho D, Kim TS. Inhibition of interleukin-12 production in mouse macrophages via decreased nuclear factor-kappaB DNA binding activity by myricetin, a naturally occurring flavonoid. Arch Pharm Res 2005; 28(3): 274-9. [http://dx.doi.org/10.1007/BF02977791] [PMID: 15832812]
- [74] Grenier D, Chen H, Ben Lagha A, Fournier-Larente J, Morin M-P. Dual action of myricetin on Porphyromonas gingivalis and the inflammatory response of host cells: A promising therapeutic molecule for periodontal diseases. PLoS One 2015; 10(6)e0131758 [http://dx.doi.org/10.1371/journal.pone.0131758] [PMID: 26121135]
- [75] Medeiros KC, Figueiredo CA, Figueredo TB, et al. Anti-allergic effect of bee pollen phenolic extract and myricetin in ovalbumin-sensitized mice. J Ethnopharmacol 2008; 119(1): 41-6. [http://dx.doi.org/10.1016/j.jep.2008.05.036] [PMID: 18588965]
- [76] Hagenacker T, Hillebrand I, Wissmann A, Büsselberg D, Schäfers M. Anti-allodynic effect of the flavonoid myricetin in a rat model of neuropathic pain: Involvement of p38 and protein kinase C mediated modulation of Ca²⁺ channels. Eur J Pain 2010; 14(10): 992-8. [http://dx.doi.org/10.1016/j.ejpain.2010.04.005] [PMID: 20471878]
- [77] Matić S, Stanić S, Bogojević D, et al. Methanol extract from the stem of Cotinus coggygria Scop., and its major bioactive phytochemical constituent myricetin modulate pyrogallol-induced DNA damage and liver injury. Mutat Res 2013; 755(2): 81-9. [http://dx.doi.org/10.1016/j.mrgentox.2013.03.011] [PMID: 23830930]
- [78] Mohan M, Gupta S, Agnihotri S, Joshi S, Uppal A. Anticataract effect of topical quercetin and myricein in galactose cataracts. Med Sci Res 1988; 16: 685-6.
- [79] Ong KC, Khoo H-E. Insulinomimetic effects of myricetin on lipogenesis and glucose transport in rat adipocytes but not glucose transport translocation. Biochem Pharmacol 1996; 51(4): 423-9. [http://dx.doi.org/10.1016/0006-2952(95)02195-7] [PMID: 8619886]
- [80] Warren CA, Paulhill KJ, Davidson LA, *et al.* Quercetin may suppress rat aberrant crypt foci formation by suppressing inflammatory mediators that influence proliferation and apoptosis. J Nutr 2009; 139(1): 101-5.
 [http://dx.doi.org/10.3945/jn.108.096271] [PMID: 19056647]
- [81] Lekakis J, Rallidis LS, Andreadou I, *et al.* Polyphenolic compounds from red grapes acutely improve endothelial function in patients with coronary heart disease. Eur J Cardiovasc Prev Rehabil 2005; 12(6): 596-600.
 [http://dx.doi.org/10.1097/00149831-200512000-00013] [PMID: 16319551]
- [82] Choi GN, Kim JH, Kwak JH, Jeong C-H, Jeong HR, Lee U, *et al.* Effect of quercetin on learning and memory performance in ICR mice under neurotoxic trimethyltin exposure. Food Chem 2012; 132(2): 1019-24. [http://dx.doi.org/10.1016/j.foodchem.2011.11.089]
- [83] Akan Z, Garip AI. Antioxidants may protect cancer cells from apoptosis signals and enhance cell viability. Asian Pac J Cancer Prev 2013; 14(8): 4611-4.

Kumar et al.

[http://dx.doi.org/10.7314/APJCP.2013.14.8.4611] [PMID: 24083712]

- [84] Alarcón de la Lastra C, Martín MJ, Motilva V. Antiulcer and gastroprotective effects of quercetin: a gross and histologic study. Pharmacology 1994; 48(1): 56-62. [http://dx.doi.org/10.1159/000139162] [PMID: 8309988]
- [85] Ramos FA, Takaishi Y, Shirotori M, et al. Antibacterial and antioxidant activities of quercetin oxidation products from yellow onion (Allium cepa) skin. J Agric Food Chem 2006; 54(10): 3551-7. [http://dx.doi.org/10.1021/jf060251c] [PMID: 19127724]
- [86] Stephen-Cole L. Quercetin: a review of clinical applications. Clin Sci (Lond) 1998; 40: 234-8.
- [87] Santini A, Novellino E. Nutraceuticals: Beyond the diet before the drugs. Curr Bioact Compd 2014; 10(1): 1-12. [http://dx.doi.org/10.2174/157340721001140724145924]
- [88] Takagaki N, Sowa Y, Oki T, Nakanishi R, Yogosawa S, Sakai T. Apigenin induces cell cycle arrest and p21/WAF1 expression in a p53-independent pathway. Int J Oncol 2005; 26(1): 185-9. [http://dx.doi.org/10.3892/ijo.26.1.185] [PMID: 15586239]
- [89] Salehi B, Venditti A, Sharifi-Rad M, et al. The therapeutic potential of apigenin. Int J Mol Sci 2019; 20(6): 1305. [http://dx.doi.org/10.3390/ijms20061305] [PMID: 30875872]
- [90] de Font-Réaulx Rojas E, Dorazco-Barragan G. [Clinical stabilisation in neurodegenerative diseases: clinical study in phase II]. Rev Neurol 2010; 50(9): 520-8. [PMID: 20443170]
- [91] Zick SM, Wright BD, Sen A, Arnedt JT. Preliminary examination of the efficacy and safety of a standardized chamomile extract for chronic primary insomnia: a randomized placebo-controlled pilot study. BMC Complement Altern Med 2011; 11(1): 78. [http://dx.doi.org/10.1186/1472-6882-11-78] [PMID: 21939549]
- [92] Shoara R, Hashempur MH, Ashraf A, Salehi A, Dehshahri S, Habibagahi Z. Efficacy and safety of topical Matricaria chamomilla L. (chamomile) oil for knee osteoarthritis: A randomized controlled clinical trial. Complement Ther Clin Pract 2015; 21(3): 181-7. [http://dx.doi.org/10.1016/j.ctcp.2015.06.003] [PMID: 26256137]
- [93] Amsterdam JD, Shults J, Soeller I, Mao JJ, Rockwell K, Newberg AB. Chamomile (Matricaria recutita) may provide antidepressant activity in anxious, depressed humans: an exploratory study. Altern Ther Health Med 2012; 18(5): 44-9. [PMID: 22894890]
- [94] Wang Q-Q, Cheng N, Yi W-B, Peng S-M, Zou X-Q. Synthesis, nitric oxide release, and α-glucosidase inhibition of nitric oxide donating apigenin and chrysin derivatives. Bioorg Med Chem 2014; 22(5): 1515-21.
 [http://dx.doi.org/10.1016/j.bmc.2014.01.038] [PMID: 24508143]
- [95] Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol 2007; 39(1): 44-84. [http://dx.doi.org/10.1016/j.biocel.2006.07.001] [PMID: 16978905]
- [96] Finkel T. Radical medicine: treating ageing to cure disease. Nat Rev Mol Cell Biol 2005; 6(12): 971-6. [http://dx.doi.org/10.1038/nrm1763] [PMID: 16227974]
- [97] López-Lázaro M. Distribution and biological activities of the flavonoid luteolin. Mini Rev Med Chem 2009; 9(1): 31-59.
 [http://dx.doi.org/10.2174/138955709787001712] [PMID: 19149659]
- [98] Palko-Labuz A, Sroda-Pomianek K, Uryga A, Kostrzewa-Suslow E, Michalak K. Anticancer activity of baicalein and luteolin studied in colorectal adenocarcinoma LoVo cells and in drug-resistant LoVo/Dx cells. Biomed Pharmacother 2017; 88: 232-41. [http://dx.doi.org/10.1016/j.biopha.2017.01.053] [PMID: 28110189]

- [99] Eumkeb G, Siriwong S, Thumanu K. Synergistic activity of luteolin and amoxicillin combination against amoxicillin-resistant Escherichia coli and mode of action. J Photochem Photobiol B 2012; 117: 247-53.
 [http://dx.doi.org/10.1016/j.jphotobiol.2012.10.006] [PMID: 23159507]
- [100] Roy S, Mallick S, Chakraborty T, *et al.* Synthesis, characterisation and antioxidant activity of luteolinvanadium(II) complex. Food Chem 2015; 173: 1172-8. [http://dx.doi.org/10.1016/j.foodchem.2014.10.141] [PMID: 25466140]
- [101] Geetha T, Garg A, Chopra K, Pal Kaur I. Delineation of antimutagenic activity of catechin, epicatechin and green tea extract. Mutat Res 2004; 556(1-2): 65-74. [http://dx.doi.org/10.1016/j.mrfmmm.2004.07.003] [PMID: 15491633]
- [102] Meyer AS, Heinonen M, Frankel EN. Antioxidant interactions of catechin, cyanidin, caffeic acid, quercetin, and ellagic acid on human LDL oxidation. Food Chem 1998; 61(1-2): 71-5. [http://dx.doi.org/10.1016/S0308-8146(97)00100-3]
- [103] Jaffri JM, Mohamed S, Rohimi N, Ahmad IN, Noordin MM, Manap YA. Antihypertensive and cardiovascular effects of catechin-rich oil palm (Elaeis guineensis) leaf extract in nitric oxide-deficient rats. J Med Food 2011; 14(7-8): 775-83. [http://dx.doi.org/10.1089/jmf.2010.1170] [PMID: 21631357]
- [104] Kang W-S, Chung K-H, Chung J-H, et al. Antiplatelet activity of green tea catechins is mediated by inhibition of cytoplasmic calcium increase. J Cardiovasc Pharmacol 2001; 38(6): 875-84. [http://dx.doi.org/10.1097/00005344-200112000-00009] [PMID: 11707691]
- [105] Chen X-Q, Hu T, Han Y, et al. Preventive effects of catechins on cardiovascular disease. Molecules 2016; 21(12): 1759.
 [http://dx.doi.org/10.3390/molecules21121759] [PMID: 28009849]
- [106] Coşarcă S, Tanase C, Muntean DL. Therapeutic aspects of catechin and its derivatives-an update. Acta Biologica Marisiensis 2019; 2(1): 21-9. [http://dx.doi.org/10.2478/abmj-2019-0003]
- [107] Ghudhaib KK. Effect of Alcoholic Catechin Extract on Hyperglycemia, Hyperlipidemia and Liver Functions in Alloxan Diabetic Mice. Baghdad Science Journal 2014; 11(3): 1192-200. [http://dx.doi.org/10.21123/bsj.11.3.1192-1200]
- [108] Fujimura Y, Umeda D, Yamada K, Tachibana H. The impact of the 67kDa laminin receptor on both cell-surface binding and anti-allergic action of tea catechins. Arch Biochem Biophys 2008; 476(2): 133-8.
 [http://dx.doi.org/10.1016/j.abb.2008.03.002] [PMID: 18358230]
- [109] Friedman M. Overview of antibacterial, antitoxin, antiviral, and antifungal activities of tea flavonoids and teas. Mol Nutr Food Res 2007; 51(1): 116-34.
 [http://dx.doi.org/10.1002/mnfr.200600173] [PMID: 17195249]
- [110] Park JH, Shon HS, Sung HK, Liu Y, Kim JY, Sung EK, et al. Anti-tumor Effect of Green Tea Catechin on Cancer Cell Lines. Korean Journal of Anatomy 2000; 33(4): 447-58.
- [111] Wilcox LJ, Borradaile NM, Huff MW. Antiatherogenic properties of naringenin, a citrus flavonoid. Cardiovasc Drug Rev 1999; 17(2): 160-78. [http://dx.doi.org/10.1111/j.1527-3466.1999.tb00011.x]
- [112] Jiannong W, Junjie J, Yanming X, et al. Effect of naringenin in Qianggu capsule on population pharmacokinetics in Chinese women with primary osteoporosis. J Tradit Chin Med 2015; 35(2): 141-53.
 - [http://dx.doi.org/10.1016/S0254-6272(15)30021-2] [PMID: 25975046]
- [113] Nyane NA, Tlaila TB, Malefane TG, Ndwandwe DE, Owira PMO. Metformin-like antidiabetic, cardio-protective and non-glycemic effects of naringenin: Molecular and pharmacological insights. Eur J Pharmacol 2017; 803: 103-11.

[http://dx.doi.org/10.1016/j.ejphar.2017.03.042] [PMID: 28322845]

- [114] Ramadan DT, Ali MA, Yahya SM, El-Sayed WM. Correlation between Antioxidant/Antimutagenic and Antiproliferative Activity of Some Phytochemicals. Anti-Cancer Agents in Medicinal Chemistry [Formerly Current Medicinal Chemistry-Anti-Cancer Agents] 2019; 19(12): 1481-90.
- [115] Ribeiro IA, Rocha J, Sepodes B, Mota-Filipe H, Ribeiro MH. Effect of naringin enzymatic hydrolysis towards naringenin on the anti-inflammatory activity of both compounds. J Mol Catal, B Enzym 2008; 52: 13-8.

[http://dx.doi.org/10.1016/j.molcatb.2007.10.011]

- [116] Lee M-H, Yoon S, Moon J-O. The flavonoid naringenin inhibits dimethylnitrosamine-induced liver damage in rats. Biol Pharm Bull 2004; 27(1): 72-6. [http://dx.doi.org/10.1248/bpb.27.72] [PMID: 14709902]
- [117] Tsai S-J, Huang C-S, Mong M-C, Kam W-Y, Huang H-Y, Yin M-C. Anti-inflammatory and antifibrotic effects of naringenin in diabetic mice. J Agric Food Chem 2012; 60(1): 514-21. [http://dx.doi.org/10.1021/jf203259h] [PMID: 22117528]
- [118] Han S-S, Lee C-K, Kim Y-S. Antimicrobial effects of naringenin alone and in combination with related flavonoids. Yakhak Hoeji 1992; 36(5): 407-11.
- [119] Ahmed OM, Hassan MA, Abdel-Twab SM, Abdel Azeem MN. Navel orange peel hydroethanolic extract, naringin and naringenin have anti-diabetic potentials in type 2 diabetic rats. Biomed Pharmacother 2017; 94: 197-205. [http://dx.doi.org/10.1016/j.biopha.2017.07.094] [PMID: 28759757]
- [120] Santos KFR. Oliveira TTd, Nagem TJ, Pinto AdS, Oliveira MGdA. Hypolipidaemic effects of naringenin, rutin, nicotinic acid and their associations 1999.
- [121] Evranos-Aksoz B, Ucar G, Tas ST, et al. New human monoamine oxidase A inhibitors with potential anti-depressant activity: design, synthesis, biological screening and evaluation of pharmacological activity. Comb Chem High Throughput Screen 2017; 20(6): 461-73. [http://dx.doi.org/10.2174/1386207320666170504113158] [PMID: 28474547]
- [122] Bansal T, Jaggi M, Khar RK, Talegaonkar S. Status of flavonols as P-glycoprotein inhibitors in cancer chemotherapy. Curr Cancer Ther Rev 2009; 5(2): 89-99. [http://dx.doi.org/10.2174/157339409788166742]
- [123] Lee JK. Anti-inflammatory effects of eriodictyol in lipopolysaccharide-stimulated raw 264.7 murine macrophages. Arch Pharm Res 2011; 34(4): 671-9. [http://dx.doi.org/10.1007/s12272-011-0418-3] [PMID: 21544733]
- [124] Habtemariam S, Dagne E. Comparative antioxidant, prooxidant and cytotoxic activity of sigmoidin A and eriodictyol. Planta Med 2010; 76(6): 589-94. [http://dx.doi.org/10.1055/s-0029-1240604] [PMID: 19941260]
- [125] Xie Y, Ji R, Han M. Eriodictyol protects H9c2 cardiomyocytes against the injury induced by hypoxia/reoxygenation by improving the dysfunction of mitochondria. Exp Ther Med 2019; 17(1): 551-7.
 [PMID: 30651835]
- [126] Mokdad-Bzeouich I, Mustapha N, Sassi A, *et al.* Investigation of immunomodulatory and anti-inflammatory effects of eriodictyol through its cellular anti-oxidant activity. Cell Stress Chaperones 2016; 21(5): 773-81.
 [http://dx.doi.org/10.1007/s12192-016-0702-8] [PMID: 27250501]
- [127] Imen M-B, Chaabane F, Nadia M, Soumaya KJ, Kamel G, Leila C-G. Anti-melanogenesis and antigenotoxic activities of eriodictyol in murine melanoma (B16-F10) and primary human keratinocyte cells. Life Sci 2015; 135: 173-8. [http://dx.doi.org/10.1016/j.lfs.2015.06.022] [PMID: 26141996]
- [128] Li Y, Yang Z-Y, Wang M-F. Synthesis, characterization, DNA binding properties and antioxidant

activity of Ln(III) complexes with hesperetin-4-one-(benzoyl) hydrazone. Eur J Med Chem 2009; 44(11): 4585-95. [http://dx.doi.org/10.1016/j.ejmech.2009.06.027] [PMID: 19615791]

- [129] Trivedi PP, Kushwaha S, Tripathi DN, Jena GB. Cardioprotective effects of hesperetin against doxorubicin-induced oxidative stress and DNA damage in rat. Cardiovasc Toxicol 2011; 11(3): 215--. [http://dx.doi.org/10.1007/s12012-011-9114-2] [PMID: 21553131]
- [130] Orallo F, Álvarez E, Basaran H, Lugnier C. Comparative study of the vasorelaxant activity, superoxide-scavenging ability and cyclic nucleotide phosphodiesterase-inhibitory effects of hesperetin and hesperidin. Naunyn Schmiedebergs Arch Pharmacol 2004; 370(6): 452-63. [http://dx.doi.org/10.1007/s00210-004-0994-6] [PMID: 15599707]
- [131] Choi EJ, Ahn WS. Neuroprotective effects of chronic hesperetin administration in mice. Arch Pharm Res 2008; 31(11): 1457-62.
 [http://dx.doi.org/10.1007/s12272-001-2130-1] [PMID: 19023542]
- [132] Shimoda K, Hamada H. Production of hesperetin glycosides by Xanthomonas campestris and cyclodextrin glucanotransferase and their anti-allergic activities. Nutrients 2010; 2(2): 171-80. [http://dx.doi.org/10.3390/nu2020171] [PMID: 22254014]
- [133] Nagashio Y, Matsuura Y, Miyamoto J, Kometani T, Suzuki T, Tanabe S. Hesperidin inhibits development of atopic dermatitis-like skin lesions in NC/Nga mice by suppressing Th17 activity. J Funct Foods 2013; 5(4): 1633-41. [http://dx.doi.org/10.1016/j.jff.2013.07.005]
- [134] Denny BJ, West PW, Mathew TC. Antagonistic interactions between the flavonoids hesperetin and naringenin and β-lactam antibiotics against Staphylococcus aureus. Br J Biomed Sci 2008; 65(3): 145-7.
 [http://dx.doi.org/10.1080/09674845.2008.11732819] [PMID: 18986103]
 - [IIII]//dx.doi.org/10.1080/090/4645.2006.11/52619] [FMID. 16960105]
- [135] Roleira FM, Tavares-da-Silva EJ, Varela CL, et al. Plant derived and dietary phenolic antioxidants: anticancer properties. Food Chem 2015; 183: 235-58. [http://dx.doi.org/10.1016/j.foodchem.2015.03.039] [PMID: 25863633]
- [136] Choi H-J. In vitro antiviral activity of sakuranetin against human rhinovirus 3. Osong Public Health Res Perspect 2017; 8(6): 415-20. [http://dx.doi.org/10.24171/j.phrp.2017.8.6.09] [PMID: 29354400]
- [137] Miyazawa M, Kinoshita H, Okuno Y. Antimutagenic activity of sakuranetin from Prunus jamasakura. J Food Sci 2003; 68(1): 52-6. [http://dx.doi.org/10.1111/j.1365-2621.2003.tb14113.x]
- [138] Ribnicky DM, Poulev A, Kuhn PE, Logendra S, Zuberi A, Cefalu WT, et al. Bioavailability Assessment of an Extract of Artemisia dracunculus L with Antidiabetic Activities In vitro and In Vivo. Diabetes 2007; •••: 56.
- [139] Atkinson P, Blakeman J. Seasonal occurrence of an antimicrobial flavanone, sakuranetin, associated with glands on leaves of Ribes nigrum. New Phytol 1982; 92(1): 63-74. [http://dx.doi.org/10.1111/j.1469-8137.1982.tb03363.x]
- [140] Liang T, Guan R, Wang Z, Shen H, Xia Q, Liu M. Comparison of anticancer activity and antioxidant activity between cyanidin-3-O-glucoside liposomes and cyanidin-3-O-glucoside in Caco-2 cells *in vitro*. RSC Advances 2017; 7(59): 37359-68. [http://dx.doi.org/10.1039/C7RA06387C]
- [141] Galvano F, La Fauci L, Vitaglione P, Fogliano V, Vanella L, Felgines C. Bioavailability, antioxidant and biological properties of the natural free-radical scavengers cyanidin and related glycosides. Ann Ist Super Sanita 2007; 43(4): 382-93. [PMID: 18209272]
- [142] Amin H. The vascular and anti-inflammatory activity of cyanidin-3-glucoside and its metabolites in

human vascular endothelial cells. University of East Anglia 2015.

- [143] Kaume L, Gilbert WC, Brownmiller C, Howard LR, Devareddy L. Cyanidin 3-O-β-D-glucoside-rich blackberries modulate hepatic gene expression, and anti-obesity effects in ovariectomized rats. J Funct Foods 2012; 4(2): 480-8. [http://dx.doi.org/10.1016/j.jff.2012.02.008]
- [144] You Q, Chen F, Wang X, Luo PG, Jiang Y. Inhibitory effects of muscadine anthocyanins on αglucosidase and pancreatic lipase activities. J Agric Food Chem 2011; 59(17): 9506-11. [http://dx.doi.org/10.1021/jf201452v] [PMID: 21797278]
- [145] Rocha-Guzmán NE, Herzog A, González-Laredo RF, Ibarra-Pérez FJ, Zambrano-Galván G, Gallegos-Infante JA. Antioxidant and antimutagenic activity of phenolic compounds in three different colour groups of common bean cultivars. Food Chem 2007; 103(2): 521-7. [Phaseolus vulgaris]. [http://dx.doi.org/10.1016/j.foodchem.2006.08.021]
- [146] Lim W-C, Kim H, Kim Y-J, et al. Delphinidin inhibits BDNF-induced migration and invasion in SKOV3 ovarian cancer cells. Bioorg Med Chem Lett 2017; 27(23): 5337-43. [http://dx.doi.org/10.1016/j.bmcl.2017.09.024] [PMID: 29122484]
- [147] Duluc L, Jacques C, Soleti R, Andriantsitohaina R, Simard G. Delphinidin inhibits VEGF inducedmitochondrial biogenesis and Akt activation in endothelial cells. Int J Biochem Cell Biol 2014; 53: 9-14.
 [http://dx.doi.org/10.1016/j.biocel.2014.03.030] [PMID: 24792670]
- [148] Sogo T, Terahara N, Hisanaga A, et al. Anti-inflammatory activity and molecular mechanism of delphinidin 3-sambubioside, a Hibiscus anthocyanin. Biofactors 2015; 41(1): 58-65. [http://dx.doi.org/10.1002/biof.1201] [PMID: 25728636]
- [149] Domitrović R, Jakovac H. Antifibrotic activity of anthocyanidin delphinidin in carbon tetrachlorideinduced hepatotoxicity in mice. Toxicology 2010; 272(1-3): 1-10. [http://dx.doi.org/10.1016/j.tox.2010.03.016] [PMID: 20371262]
- [150] Ravindra PV, Narayan MS. Antioxidant activity of the anthocyanin from carrot (Daucus carota) callus culture. Int J Food Sci Nutr 2003; 54(5): 349-55. [http://dx.doi.org/10.1080/09637480120092134] [PMID: 12907406]
- [151] Xu H, Zhang J, Huang H, Liu L, Sun Y. Malvidin induced anticancer activity in human colorectal HCT-116 cancer cells involves apoptosis, G2/M cell cycle arrest and upregulation of p21WAFI. Int J Clin Exp Med 2018; 11(3): 1734-41.
- [152] Saulite L, Jekabsons K, Klavins M, Muceniece R, Riekstina U. Effects of malvidin, cyanidin and delphinidin on human adipose mesenchymal stem cell differentiation into adipocytes, chondrocytes and osteocytes. Phytomedicine 2019; 53: 86-95. [http://dx.doi.org/10.1016/j.phymed.2018.09.029] [PMID: 30668416]
- [153] Bognar E, Sarszegi Z, Szabo A, et al. Antioxidant and anti-inflammatory effects in RAW264.7 macrophages of malvidin, a major red wine polyphenol. PLoS One 2013; 8(6)e65355 [http://dx.doi.org/10.1371/journal.pone.0065355] [PMID: 23755222]
- [154] Jeong S, Ku S-K, Bae J-S. Anti-inflammatory effects of pelargonidin on TGFBIp-induced responses. Can J Physiol Pharmacol 2017; 95(4): 372-81. [http://dx.doi.org/10.1139/cjpp-2016-0322] [PMID: 28060523]
- [155] Min G, Ku S-K, Park MS, Park T-J, Lee H-S, Bae J-S. Anti-septic effects of pelargonidin on HMGB1induced responses *in vitro* and in vivo. Arch Pharm Res 2016; 39(12): 1726-38. [http://dx.doi.org/10.1007/s12272-016-0834-5] [PMID: 27778275]
- [156] Chen Y, Wang S, Geng B, Yi Z. Pelargonidin induces antitumor effects in human osteosarcoma cells via autophagy induction, loss of mitochondrial membrane potential, G2/M cell cycle arrest and downregulation of PI3K/AKT signalling pathway. J BUON 2018; 23(3): 735-40. [PMID: 30003744]

- [157] Guo L, Otgonbayar D, Cui Z, Park JH, Kang JS, Kang NJ, et al. Pelargonidin From Strawberry Reduces Adipogenesis by Inhibition of PPAR-γ Pathway Signaling in 3T3-L1 Cells.
- [158] Ku S-K, Yoon E-K, Lee W, Kwon S, Lee T, Bae J-S. Antithrombotic and antiplatelet activities of pelargonidin in vivo and *in vitro*. Arch Pharm Res 2016; 39(3): 398-408. [http://dx.doi.org/10.1007/s12272-016-0708-x] [PMID: 26762345]
- [159] Lee I-C, Bae J-S. Pelargonidin protects against renal injury in a mouse model of sepsis. J Med Food 2019; 22(1): 57-61.
 [http://dx.doi.org/10.1089/jmf.2018.4230] [PMID: 30160593]
- [160] Roy M, Sen S, Chakraborti AS. Action of pelargonidin on hyperglycemia and oxidative damage in diabetic rats: implication for glycation-induced hemoglobin modification. Life Sci 2008; 82(21-22): 1102-10.
 [102-10.

[http://dx.doi.org/10.1016/j.lfs.2008.03.011] [PMID: 18440560]

- [161] Rahman M, Sabir AA, Mukta JA, *et al.* Plant probiotic bacteria Bacillus and Paraburkholderia improve growth, yield and content of antioxidants in strawberry fruit. Sci Rep 2018; 8(1): 2504. [http://dx.doi.org/10.1038/s41598-018-20235-1] [PMID: 29410436]
- [162] Sari DRT, Cairns JRK, Safitri A, Fatchiyah F. Virtual Prediction of the Delphinidin-3-O-glucoside and Peonidin-3-O-glucoside as Anti-inflammatory of TNF-α Signaling. Acta Inform Med 2019; 27(3): 152-7.
 [http://dx.doi.org/10.5455/aim.2019.27.152-157] [PMID: 31762569]
- [163] Ho M-L, Chen P-N, Chu S-C, et al. Peonidin 3-glucoside inhibits lung cancer metastasis by downregulation of proteinases activities and MAPK pathway. Nutr Cancer 2010; 62(4): 505-16. [http://dx.doi.org/10.1080/01635580903441261] [PMID: 20432172]
- [164] Yoshimoto M, Okuno S, Yamaguchi M, Yamakawa O. Antimutagenicity of deacylated anthocyanins in purple-fleshed sweetpotato. Biosci Biotechnol Biochem 2001; 65(7): 1652-5. [http://dx.doi.org/10.1271/bbb.65.1652] [PMID: 11515552]
- [165] Choo M-K, Park E-K, Yoon H-K, Kim D-H. Antithrombotic and antiallergic activities of daidzein, a metabolite of puerarin and daidzin produced by human intestinal microflora. Biol Pharm Bull 2002; 25(10): 1328-32.
 [http://dx.doi.org/10.1248/bpb.25.1328] [PMID: 12392089]
- [166] Hua F, Li CH, Chen XG, Liu XP. Daidzein exerts anticancer activity towards SKOV3 human ovarian cancer cells by inducing apoptosis and cell cycle arrest, and inhibiting the Raf/MEK/ERK cascade. Int J Mol Med 2018; 41(6): 3485-92. [http://dx.doi.org/10.3892/ijmm.2018.3531] [PMID: 29512690]
- [167] Ahmad S, Alam K, Hossain MM, et al. Anti-arthritogenic and cardioprotective action of hesperidin and daidzein in collagen-induced rheumatoid arthritis. Mol Cell Biochem 2016; 423(1-2): 115-27. [http://dx.doi.org/10.1007/s11010-016-2830-y] [PMID: 27704466]
- [168] Dwiecki K, Neunert G, Polewski P, Polewski K. Antioxidant activity of daidzein, a natural antioxidant, and its spectroscopic properties in organic solvents and phosphatidylcholine liposomes. J Photochem Photobiol B 2009; 96(3): 242-8. [http://dx.doi.org/10.1016/j.jphotobiol.2009.06.012] [PMID: 19648024]
- [169] Fujioka M, Uehara M, Wu J, et al. Equol, a metabolite of daidzein, inhibits bone loss in ovariectomized mice. J Nutr 2004; 134(10): 2623-7.
 [http://dx.doi.org/10.1093/jn/134.10.2623] [PMID: 15465757]
- [170] Ae Park S, Choi M-S, Cho S-Y, et al. Genistein and daidzein modulate hepatic glucose and lipid regulating enzyme activities in C57BL/KsJ-db/db mice. Life Sci 2006; 79(12): 1207-13. [http://dx.doi.org/10.1016/j.lfs.2006.03.022] [PMID: 16647724]
- [171] Živanović J, Jarić I, Ajdžanović V, et al. Daidzein upregulates anti-aging protein Klotho and NaPi 2a

cotransporter in a rat model of the andropause. Ann Anat 2019; 221: 27-37. [http://dx.doi.org/10.1016/j.aanat.2018.08.001] [PMID: 30240906]

- [172] ZENG J, HUANG Z-h, QIU F, YAO X-s, YE H-y. The anti-hypoxia activity of daidzein [J]. The Chinese Journal of Modern Applied Pharmacy 2004; 6.
- [173] Lehmann L, Esch HL, Wagner J, Rohnstock L, Metzler M. Estrogenic and genotoxic potential of equol and two hydroxylated metabolites of Daidzein in cultured human Ishikawa cells. Toxicol Lett 2005; 158(1): 72-86. [http://dx.doi.org/10.1016/j.toxlet.2005.02.011] [PMID: 15993745]
- [174] Oh WY, Shahidi F. Antioxidant activity of resveratrol ester derivatives in food and biological model systems. Food Chem 2018; 261: 267-73. [http://dx.doi.org/10.1016/j.foodchem.2018.03.085] [PMID: 29739593]
- [175] Karthikeyan S, Prasad NR, Ganamani A, Balamurugan E. Anticancer activity of resveratrol-loaded gelatin nanoparticles on NCI-H460 non-small cell lung cancer cells. Biomedicine & Preventive Nutrition 2013; 3(1): 64-73. [http://dx.doi.org/10.1016/j.bionut.2012.10.009]
- [176] Andrade S, Ramalho MJ, Pereira MDC, Loureiro JA. Pereira MdC, Loureiro JA. Resveratrol brain delivery for neurological disorders prevention and treatment. Front Pharmacol 2018; 9: 1261. [http://dx.doi.org/10.3389/fphar.2018.01261] [PMID: 30524273]
- [177] Wu JM, Hsieh TC. Resveratrol: a cardioprotective substance. Ann N Y Acad Sci 2011; 1215(1): 16--. [http://dx.doi.org/10.1111/j.1749-6632.2010.05854.x] [PMID: 21261637]
- [178] Hausenblas HA, Schoulda JA, Smoliga JM. Resveratrol treatment as an adjunct to pharmacological management in type 2 diabetes mellitus--systematic review and meta-analysis. Mol Nutr Food Res 2015; 59(1): 147-59. [http://dx.doi.org/10.1002/mnfr.201400173] [PMID: 25138371]
- [179] Li L, Hai J, Li Z, et al. Resveratrol modulates autophagy and NF-κB activity in a murine model for treating non-alcoholic fatty liver disease. Food Chem Toxicol 2014; 63: 166-73. [http://dx.doi.org/10.1016/j.fct.2013.08.036] [PMID: 23978414]
- [180] Matsuda H, Tewtrakul S, Morikawa T, Yoshikawa M. Anti-allergic activity of stilbenes from Korean rhubarb (Rheum undulatum L.): structure requirements for inhibition of antigen-induced degranulation and their effects on the release of TNF-α and IL-4 in RBL-2H3 cells. Bioorg Med Chem 2004; 12(18): 4871-6.
 [http://dx.doi.org/10.1016/j.bmc.2004.07.007] [PMID: 15336266]
- [181] Zhou ZX, Mou SF, Chen XQ, Gong LL, Ge WS. Anti-inflammatory activity of resveratrol prevents inflammation by inhibiting NF□κB in animal models of acute pharyngitis. Mol Med Rep 2018; 17(1): 1269-74.
 [PMID: 29115472]
- [182] Gao X, Xu YX, Janakiraman N, Chapman RA, Gautam SC. Immunomodulatory activity of resveratrol: suppression of lymphocyte proliferation, development of cell-mediated cytotoxicity, and cytokine production. Biochem Pharmacol 2001; 62(9): 1299-308. [http://dx.doi.org/10.1016/S0006-2952(01)00775-4] [PMID: 11705464]
- [183] Yamamoto T, Li Y, Hanafusa Y, *et al.* Piceatannol exhibits anti-inflammatory effects on macrophages interacting with adipocytes. Food Sci Nutr 2016; 5(1): 76-85. [http://dx.doi.org/10.1002/fsn3.366] [PMID: 28070318]
- [184] Kuo PL, Hsu YL. The grape and wine constituent piceatannol inhibits proliferation of human bladder cancer cells via blocking cell cycle progression and inducing Fas/membrane bound Fas ligandmediated apoptotic pathway. Mol Nutr Food Res 2008; 52(4): 408-18. [http://dx.doi.org/10.1002/mnfr.200700252] [PMID: 18381677]
- [185] Piotrowska H, Kucinska M, Murias M. Biological activity of piceatannol: leaving the shadow of

resveratrol. Mutat Res 2012; 750(1): 60-82. [http://dx.doi.org/10.1016/j.mrrev.2011.11.001] [PMID: 22108298]

- [186] Sato D, Shimizu N, Shimizu Y, *et al.* Synthesis of glycosides of resveratrol, pterostilbene, and piceatannol, and their anti-oxidant, anti-allergic, and neuroprotective activities. Biosci Biotechnol Biochem 2014; 78(7): 1123-8.
 [http://dx.doi.org/10.1080/09168451.2014.921551] [PMID: 25229845]
- [187] Park Y, Lee M-H. Anticariogenic activity of piceatannol isolated fromCallistemon citrinus fruit against Streptococcus mutans. 2008; 21(6): 431-.
- [188] Ko SK, Lee SM, Whang WK. Anti-platelet aggregation activity of stilbene derivatives from Rheum undulatum. Arch Pharm Res 1999; 22(4): 401-3. [http://dx.doi.org/10.1007/BF02979065] [PMID: 10489881]
- [189] Mérillon J-M, Fauconneau B, Teguo PW, Barrier L, Vercauteren J, Huguet F. Antioxidant activity of the stilbene astringin, newly extracted from Vitis vinifera cell cultures. Clin Chem 1997; 43(6 Pt 1): 1092-3.
 [http://dx.doi.org/10.1093/clinchem/43.6.1092] [PMID: 9191572]
- [190] Ashok PK, Upadhyaya K. Tannins are astringent. J Pharmacogn Phytochem 2012; 1(3): 45-50.
- [191] Luo J, Hu Y, Kong W, Yang M. Evaluation and structure-activity relationship analysis of a new series of arylnaphthalene lignans as potential anti-tumor agents. PLoS One 2014; 9(3)e93516 [http://dx.doi.org/10.1371/journal.pone.0093516] [PMID: 24675875]
- [192] Musey PI, Adlercreutz H, Gould KG, et al. Effect of diet on lignans and isoflavonoid phytoestrogens in chimpanzees. Life Sci 1995; 57(7): 655-64. [http://dx.doi.org/10.1016/0024-3205(95)00317-Y] [PMID: 7637537]
- [193] Liu S, Wei W, Shi K, Cao X, Zhou M, Liu Z. *In vitro* and in vivo anti-hepatitis B virus activities of the lignan niranthin isolated from Phyllanthus niruri L. J Ethnopharmacol 2014; 155(2): 1061-7. [http://dx.doi.org/10.1016/j.jep.2014.05.064] [PMID: 25009077]
- [194] Yamauchi S, Taniguchi E. Synthesis and insecticidal activity of lignan analogs. Biosci Biotechnol Biochem 1992; 56(3): 412-7. [II].
 [http://dx.doi.org/10.1271/bbb.56.412] [PMID: 27320990]
- [195] Choi S-R, Kim C-S, Kim J-Y, You D-H, Kim J-M, Kim Y-S, *et al.* Changes of antioxidant activity and lignan contents in Schisandra chinensis by harvesting times. Hanguk Yakyong Changmul Hakhoe Chi 2011; 19(6): 414-20. [http://dx.doi.org/10.7783/KJMCS.2011.19.6.414]
- [196] Xu Z, Ju J, Wang K, Gu C, Feng Y. Evaluation of hypoglycemic activity of total lignans from Fructus Arctii in the spontaneously diabetic Goto-Kakizaki rats. J Ethnopharmacol 2014; 151(1): 548-55. [http://dx.doi.org/10.1016/j.jep.2013.11.021] [PMID: 24269245]
- [197] Choi HG, Choi YH, Kim JH, et al. A new neolignan and lignans from the stems of Lindera obtusiloba Blume and their anti-allergic inflammatory effects. Arch Pharm Res 2014; 37(4): 467-72. [http://dx.doi.org/10.1007/s12272-013-0239-7] [PMID: 24014307]
- [198] Zanwar AA, Hegde MV, Bodhankar SL. Cardioprotective activity of flax lignan concentrate extracted from seeds of Linum usitatissimum in isoprenalin induced myocardial necrosis in rats. Interdiscip Toxicol 2011; 4(2): 90-7. [http://dx.doi.org/10.2478/v10102-011-0016-8] [PMID: 21753905]
- [199] Kuroda T, Kondo K, Iwasaki T, Ohtani A, Takashima K. Synthesis and hypolipidemic activity of diesters of arylnaphthalene lignan and their heteroaromatic analogs. Chem Pharm Bull (Tokyo) 1997; 45(4): 678-84.
 [http://dx.doi.org/10.1248/cpb.45.678] [PMID: 9145504]
- [200] Kiyama R. Biological effects induced by estrogenic activity of lignans. Trends Food Sci Technol 2016; 54: 186-96.

[http://dx.doi.org/10.1016/j.tifs.2016.06.007]

- [201] de-Faria FM, Almeida ACA, Luiz-Ferreira A, et al. Mechanisms of action underlying the gastric antiulcer activity of the Rhizophora mangle L. J Ethnopharmacol 2012; 139(1): 234-43. [http://dx.doi.org/10.1016/j.jep.2011.11.007] [PMID: 22100564]
- [202] Shohayeb M, Abdel-Hameed E, Bazaid S. Antimicrobial activity of tannins and extracts of different parts of Conocarpus erectus L. Int J Pharm Bio Sci 2013; 3(2): 544-53.
- [203] Orlowski P, Tomaszewska E, Gniadek M, et al. Tannic acid modified silver nanoparticles show antiviral activity in herpes simplex virus type 2 infection. PLoS One 2014; 9(8)e104113 [http://dx.doi.org/10.1371/journal.pone.0104113] [PMID: 25117537]
- [204] Hu X, Wang H, Lv X, et al. Cardioprotective effects of tannic acid on isoproterenol induced myocardial injury in rats: Further insight into 'french paradox'. Phytother Res 2015; 29(9): 1295-303. [http://dx.doi.org/10.1002/ptr.5376] [PMID: 25989747]
- [205] Kanoh R, Hatano T, Ito H, Yoshida T, Akagi M. Effects of tannins and related polyphenols on superoxide-induced histamine release from rat peritoneal mast cells. Phytomedicine 2000; 7(4): 297-302.

[http://dx.doi.org/10.1016/S0944-7113(00)80047-1] [PMID: 10969723]

- [206] Barrajón-Catalán E, Fernández-Arroyo S, Saura D, et al. Cistaceae aqueous extracts containing ellagitannins show antioxidant and antimicrobial capacity, and cytotoxic activity against human cancer cells. Food Chem Toxicol 2010; 48(8-9): 2273-82. [http://dx.doi.org/10.1016/j.fct.2010.05.060] [PMID: 20510328]
- [207] Kunyanga CN, Imungi JK, Okoth M, Momanyi C, Biesalski HK, Vadivel V. Antioxidant and antidiabetic properties of condensed tannins in acetonic extract of selected raw and processed indigenous food ingredients from Kenya. J Food Sci 2011; 76(4): C560-7. [http://dx.doi.org/10.1111/j.1750-3841.2011.02116.x] [PMID: 22417336]
- [208] Hui LFTJS, Du Lijun XDLH. An Evaluation of Anti-Obesity and Anti-hyperlipidemia of Pomegranate Tannins Using Hierarchy Program. World Science and Technology-Modernization of Traditional Chinese Medicine and Materia Medica 2007; p. 4. J
- [209] Xiong Y-a, Yu Q-n, Zou J-b, He Y-h, Zhang S-j, Xu R-c, et al. Protective effects of tannins in Sanguisorbae Radix on myelosuppression mice. Chin Herb Med 2014; 6(3): 222-7. [http://dx.doi.org/10.1016/S1674-6384(14)60032-0]
- [210] Li K, Diao Y, Zhang H, et al. Tannin extracts from immature fruits of Terminalia chebula Fructus Retz. promote cutaneous wound healing in rats. BMC Complement Altern Med 2011; 11(1): 86. [http://dx.doi.org/10.1186/1472-6882-11-86] [PMID: 21982053]
- [211] Yoshizawa S, Horiuchi T, Fujiki H, Yoshida T, Okuda T, Sugimura T. Antitumor promoting activity of [-] epigallocatechin gallate, the main constituent of "Tannin" in green tea. Phytother Res 1987; 1(1): 44-7.

[http://dx.doi.org/10.1002/ptr.2650010110]

Structure-Activity Relationship of Flavonoids: Recent Updates

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Abstract: The biological, physicochemical, and environmental properties of substances are anticipated by utilizing structure-activity relationships (SARs). A SAR is a (qualitative) correlation between a chemical substructure and the potential for one of the chemicals in the substructure to have a particular biological property or effect. We are familiar with the term SAR which is a powerful concept in the discovery of any active pharmaceutical ingredient with both qualitative and quantitative associations that relate to the chemical structure and biological activity of any chemical compound. Due to their safety and medicinal efficacy, plant-derived functional foods are of great interest. In this chapter, the different types of biologically active compounds, their chemistry and SAR, and the different biologically active compounds from daily dietary supplements, foods, and fruits, which contain polyphenolic compounds, have been discussed. SAR of flavonoids like anthocyanidins, which is the principal component of the majority of fruits, vegetables, and flower petals, flavonols, usually called catechins, and isoflavones, are also discussed. Other polyphenolic compounds like tannins with their subtypes like hydrolysable and non-hydrolysable tannins are well covered.

Keywords: Flavonoids, Polyphenols, Stilbenes, Structure-activity relationships, Tannins.

INTRODUCTION

Herbal or medicinal plants have gained exceptional importance around the world. Plant products have been noticed for nutrition, cosmetics preparations, diagnostic agents, and mitigating diseases of human beings. Therefore, a plethora of studies have been done on different plant species, and their effects have been investigat-

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ed. Due to their safety and medicinal efficacy, plant-derived functional foods are of great interest.

Many researchers have already conducted studies on secondary metabolites of plants because of their importance as dietary supplements. The first scientist who discovered the presence of secondary metabolites in plants was Kossel. Among secondary metabolites, 'Polyphenols' are diverse in nature and found in large varieties of plants with antioxidant, anti-inflammatory, and antimicrobial properties. They have gained importance as they have evolved in growth and reproduction, and they provide resistance to plants against pathogens and predators. Moreover, they protect crops from plague and preharvest seed germination [1, 2]. They are polyhydroxylated phytochemicals with common structures. Polyphenols are the secondary metabolites derived from the two main synthetic pathways: shikimic acid and acetate pathway. Chemical structures of natural polyphenols vary from basic phenolic acids to strongly polymerized compounds like tannins. Three main subclasses of phytochemicals that add abundant micronutrients to the diet are phenolic acids (derivatives of cinnamic acid and benzoic acid), flavonoids, and stilbenoids (stilbenes). Phenolic compounds are the most complex classes of chemicals present in the plants. More than 8000 compounds are assumed to have been isolated [3, 4]. They are widely distributed in fruits like berries, apricots, cherries, apples, grapes, and pears, vegetables such as onion, garlic, carrot, tomato, cabbage, and celery, beverages like chocolates, wine, tea, and coffee, and are consumed as dietary supplements [5]. Structural diversity of different polyphenolic compounds has been reviewed in this chapter. The different types of phenolic compounds, their food sources, and their biogenetic pathways have been summarized in Table 1.

Type of Phenolic Compound	Chemistry	Structure	Food Source	Biological Activity	Biogenetic Pathway
Phenolic acids	A phenolic ring and an organic carboxylic acid feature are present in this class of polyphenolic compounds (C6-C1 skeleton) [6].	ОН	Generally found in Horse grams, Mushrooms, and dry fruits [7].	Antimicrobial and antioxidant activity [8].	They are derived from L-tyrosine or L- phenylalanine through the Shikimate pathway [8].
Flavonoids This is the most common class, which has a general structural backbone		roots, branches, bulbs, tea, and wine are all high in	Reported to have antioxidant, antihypertensive activity and is used in many cardiovascular disorders [9].	They are synthesized by the phenylpropanoid pathway [10].	

Table 1. Food sources and biogenetic pathway of some phenolic compounds.
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Flavonoids

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Type of Phenolic Compound	Chemistry	Structure	Food Source	Biological Activity	Biogenetic Pathway
Furanocoumarins	Furanocoumarins have a furan ring connected to a coumarin ring in their chemical structure. The furan ring can be fused in many ways to produce a variety of isomers [11].		Mainly present in citrus fruits like oranges, lemons, mandarins, grapefruits, and limes [12].	known for	phenylalanine through the shikimate pathway
Lignans	Two phenylpropane units are linked by a C-C bond between the central atoms of the respective side chains (position 8 or β), also known as the β - β ' bond [16].		Vegetables, grain, nuts, beans, and beverages like tea, coffee, and wine are all rich in this nutrient. Flaxseed contains the most significant amounts of dietary lignans as secoisolariciresinol diglucoside [17].	antihypertensive, antiviral, estrogenic, and insecticidal properties have been documented	They are synthesized by the phenylpropanoid pathway [19].
Stilbenes	Stilbenes are phenylpropanoids with a 1,2- diphenylethylene backbone and belong to a small group of phenylpropanoids [19].		metabolite found in grapevine, berries, and	various biological effects, including neuroprotection, cardioprotection, depigmentation,	critical link between primary metabolism and secondary metabolic pathways such as phenylpropanoid, flavonoid, and stilbenoid
Hydrolysable Tannins	They are made up of polyhydric alcohol with hydroxyl groups that are partly or wholly esterified with gallic or hexahydroxy diphenic and having long chains of gallic acid coming from the central glucose core [23].	HO + O + O + O + O + O + O + O + O + O +	They are found in leguminous seeds, cereals, and, most importantly, in many fruits and vegetables [24, 25].	antimicrobial, cardioprotective, and anti-cancer properties, in addition to antioxidant and free radical scavenging properties. They also tend to have a	A UDP- glucosyltransferase (UGT) action forms an ester bond between gallic acid and glucose to produce β β-glucogallin is converted to pentagalloylglucose, which is then converted to gallotannin by several acyltransferases [27, 28].

The different polyphenols have been summarized with their chemistry, structure, biological sources, their biological activity, and their biogenetic pathway.

PHENOLIC ACIDS

There are two kinds of phenolic acids found in polyphenols: hydroxybenzoic acid

and hydroxycinnamic acid. Salicylic acid (Fig. 1), a hydroxybenzoic acid derived from the bark of willow trees (Salix spp.) [29], was the earliest compound identified to have therapeutic activity. It was chewed in herbal medicine to provide pain and inflammation relief [30].



Fig. (1). Salicylic acid.

Salicylic acid is biosynthesized in plants by the shikimic acid pathway, which is a typical intermediate for a number of branched pathways that produce tyrosine, tryptophan, phenylalanine, and a variety of other aromatic compounds. The structure of salicylic acid is:

If we consider SAR, the presence of hydroxyl group at ortho position is important for the pharmacological activity. Shifting of the hydroxyl group at meta or para position abolishes the activity. Substitution of the ring decreases the activity of salicylic acid [31, 32]. The side effects related to the polyphenolic compound are associated with the presence of the carboxylic acid group. Salicylic acid is used in medicinal preparation to cure dandruff, seborrhea, acne, and insect bites [33].

If we consider hydroxy-cinnamic acid (Fig. 2), which is more common when compared to hydroxyl-benzoic acid, it mainly consists of ferulic acid and caffeic acid [34]. Caffeic acid is predominantly found in different fruits like plum, blueberries, cherries, apples, and kiwi [35]. The structure is shown below:

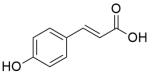


Fig. (2). Hydroxycinnamic acid.

If we consider SAR, hydroxy-cinnamic acid is a highly potent antioxidative agent. The structural modification can be done for more potency [36]. The availability of an unsaturated bond on the side chain is essential for their biological activity. Modifications of the aromatic ring, such as changes in position and number of hydroxyl groups and the addition of an electron-withdrawing group, as well as modifications of the carboxylic group, are essential structural features for antioxidant activity [37, 38].

FLAVONOIDS

Flavonoids have been found in over 8000 different varieties and many of them. are responsible for the colours of fruits and flowers. Humans receive them from the consumption of seeds, tea, wine, berries, dark chocolate, wheat, roots, leaves, stems, and flowers. From the literature, it has been found that these are the largest group of polyphenols, which include several thousands of low molecular weight phenolic compounds, belonging to 7 major subgroups – flavanols, flavones, flavanols, flavanones, flavanonols, anthocyanidins, and isoflavonoids (Tables 2 & 3) Among these, flavones, flavanols, and flavanones are the most abundant naturally occurring flavonoids. These molecules are characterized by the presence of flavan nucleus and are known as C6–C3–C6 phenolics [39]. Flavonoids, like carotenoids, are responsible for the coloration of fruits, vegetables, and herbs [40]. The flavonoid parent compound (Fig. 3) is made up of a fifteen-carbon skeleton that includes two benzene rings and a heterocyclic pyrane ring (C). They are divided into flavones (e.g., apigenin, flavone, and luteolin), flavonols (e.g., myricetin, quercetin, kaempferol, and fisetin), and flavanones (e.g., hesperetin, flavanone and naringenin) [41]. The aglycone is the flavonoid's basic parent structure [42]. An α -pyrone (flavonols and flavanones) or its dihydroderivative (flavonols and flavanones) is a six-membered ring structure condensed with the benzene ring. Flavonoids (2-position) and isoflavonoids (3-position) are distinguished by the position of the benzenoid substituent. Flavonols differ from flavanones in having a hydroxyl group at 3-position and a C2–C3 double bond [43]. Flavonoids are hydroxylated in positions 3, 5, 7, 2, 3', 4', and 5'. In nature, methyl ethers and acetyl esters of the alcohol group are known [44].

S. no.	Types of Flavonoids	Chemical Structures
1.	FLAVONES a. Apiogenin	HO HO O

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(Table 2) cont	T-mar of Flammer 1	Charries 1 States at 199
S. no.	Types of Flavonoids	Chemical Structures
	b. Luteolin	HO OH OH HO OH
2.	FLAVONOLS a. Quercetin b. Kaempferol	HO HO O O O O O O O H O H O H
3.	ISOFLAVONOLS a. Genistein b. Daidzein	HO O OH HO OH O OH
4.	CHALCONES	HO O HO O O OH
5.	ANTHOCYANINS	HO O O OH
6.	FLAVANONES	

Flavonoids

S. no.	Types of Flavonoids	Chemical Structures
7.	FLAVONONOLS	ö

Table 3. Different types of flavonoid have been summarized with their chemistry	and biological activity.

S.no.	Sub-Classes of Flavonoids	Chemistry	Food Sources	Biological Activity
1.	Isoflavonoids	They are also called as phytoestrogens. They have 3- phenylchroman backbone [52].	especially red clover, soybeans, and other legumes, are the	nonsteroidal polyphenolic metabolites found in
2.	Flavones	Flavones have a double bond between C-2 and C-3 and a non- saturated 3-C chain [56].	in leaves, flowers, and fruits as glucosides. Celery, parsley, chamomile, red peppers, ginkgo biloba and mint, are	cardiovascular and neurological disorders have been shown by
3.	Flavonols	They have a double bond between positions 2 and 3 of the C ring, as well as a ketonic group at position 4. A hydroxyl group is present in position 5 of the A ring in the majority of flavones found in vegetables and fruits [59].	bananas, brussels sprouts, cabbage, sprouts, green beans, kale, endive, spinach,	

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S.no.	Sub-Classes of Flavonoids	Chemistry	Food Sources	Biological Activity
4.	Chalcones	acetophenone. An aliphatic three- carbon chain connects two aromatic rings in chalcones.	most common in citrus fruits, apples, and vegetables such as tomatoes, green sprouts, potatoes, and various plants	antitubercular, anti- inflammatory, antioxidant, anti-leishmanial activity are present in chalcones
5.	Anthocyanidins	This type of flavonoid consists of polyhydroxy derivatives of 2- phenylbenzophyryllium [65]. A three-carbon chain connects the two aromatic rings A and B with the phenolic group, forming an oxygenated heterocyclic ring (C ring) [66].	blueberries, cherries, chokeberries, black and red currants, grapes, strawberries, pomegranate, elderberries, and other fruits and vegetables have	agent, in neurological and coronary disorders, inflammation, diabetes, and bacterial infections. The chelating property of phenolic compounds is
6.	Flavanones	Flavanone is made up of two aromatic rings joined by a carbon bridge, which often forms a heterocyclic ring [69].	parsley, rutin, olives, spinaches, and other foods contain	cardiovascular health,
7.	Flavonols	These are dihydroflavonols with saturated C3 fragment which make them odorless [72].		

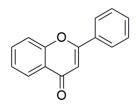


Fig. (3). Parent structure of flavonoid.

Flavonoids

Structure Activity Relationship

Substitution at the A or B ring in the flavonoid structure increases phyto-estrogen activity. The existence of the catechol moiety is the most significant factor for high antioxidant activity. In this case, the presence of a C2-C3 double bond is of no significance [45]. In the absence of catechol, the C2-C3 double bond, along with C3-OH and C4 carbonyl, is crucial for antioxidant action [46]. If structure is considered, the total number of OH groups and position of OH at 2nd carbon is very important. The flavonoid is inactive when there is no OH or just one OH present. Isoflavones having OH groups at position 4' on ring B and presence of OH group at 6 and 7 on ring A showed an increase in inhibitory property [47]. Generally, C2 and C3 double bond is very necessary for the antioxidant activity. The different sub-classes of flavonoids can be formed by the substitution or the presence of carbon on ring C on which B ring is attached [48]. The degree of unsaturation and oxidation of C ring is also used to classify different sub-classes of flavonoids [49]. If structure-activity relationship is considered, the molecular form and the capacity of hydrogen atom donation to free radicals is responsible for the antioxidant activity. The presence of phenolic hydrogen and stability of phenoxy free radical and the substitution of phenolic hydroxyl group are responsible for the activity. The presence of 3,4-dihydroxy in case of quercetin, luteolin and catechin makes them highly effective antioxidants [50, 51].

FURANOCOUMARINS

Secondary metabolites found in citrus plants contain furanocoumarins—the fusion of the furan ring with the α -benzopyrone forms the structure. Furan moiety can be in either a 3,2 - or 2,3 -arrangement at the c, f, g, or h bonds of the coumarin; most naturally occurring derivatives belong to the psoralen, allopsoralen, and angelicin group [75]. Coumarin and its derivatives act as anticoagulants and are taken orally. They are insoluble in water; however, the 4-hydroxy substitution gives the molecule weakly acidic properties, making it water-soluble in mildly alkaline environments.

The different most commonly occurring furanocoumarins are:

a. **\alpha-Benzopyrone:** It is found in all aspects of plants, herbs, citrus, spices, the fruits, roots, branches, and leaves. The general structure is given in Fig. (4) [76, 77].



Fig. (4). α-Benzopyrone.

b. **Psoralen:** Psoralen is the parent molecule in a family of furanocoumarins (Fig. **5**). It is found in plants like *Psoralea corylifolia* and *Ficus salicifolia* and acts as a plant metabolite. It is structurally related to coumarin by the addition of a fused furan ring and derivative of 9-methoxy-7*H*-furo [3,2-g]chromen-7-one tricyclic ring structures. Cross-linkers, biotinylation complexes, and nucleic acid probes all contain them as photoreactive groups. Psoralens have been used as a photochemotherapy agent for the treatment of psoriasis and vitiligo for many years [78 - 80].

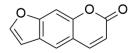
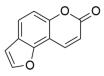


Fig. (5). Psoralen.

c. Allopsoralen: It is a novel tetracyclic coumarin derivative produced by the condensation of a fourth cyclohexenylic (5-7) or benzenic (8-10) ring at the furan side with a methoxy (5 and 8) or hydroxy (6 and 9) at ten position (Fig. 6) [81].





d. **Angelicin:** Angelicin is the parent compound of naturally occurring angular furanocoumarins. It is made up of benzapyra-2-one fused to a furan moiety at 7,8-position [82]. Plants in the Apiaceae and Fabaceae families, such as *Bituminaria bituminosa*, contain lots of angelicin (Fig. 7). Angelicin derivatives are used for the treatment of cancer and psoriasis [83] and are also reported to have a tranquillo sedative and anticonvulsant activity [84].

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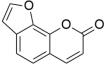


Fig. (7). Angelicin.

LIGNANS

Lignans are the polyphenolic compounds having steroid-like chemical structures, also known as phytoestrogens (Fig. 8). Heart attack, menopausal complications, osteoporosis, and breast cancer are decreased by lignans, according to a study [85]. Lignans are nothing but phenylpropanoid dimers consisting of two phenyl propane units (C6 and C3) linked by their central C8 carbon and mainly synthesized by phenylpropanoid pathway [86]. Lignans are present in a wide range of foods, including nuts, beans, grains, vegetables, and beverages, including coffee, tea, and wine. Flaxseed has the highest concentrations of dietary lignans [87]. It is well known that ligning and lignans both originate from the C6-C3 units, which indicates that these metabolites are biosynthesized through phenylpropanoid pathway. If the structure of lignans is considered, they contain a basic two or more phenylpropanoid units [88] and the monomers which form lignans. Classical lignans are the molecules that have a molecular linkage between positions β - β' (also referred to as an 8-8'). If the major structural units are linked in some other ways (non β - β ' linkage), the compounds are known as "neolignans" [89]. There is oxidation at C9 and C9' carbon of the most of the natural lignans. The different lignans are formed by the incorporation of oxygen into the parent structure and on the cyclization process [90].

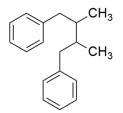


Fig. (8). Lignan.

Lignans have a wide range of structural variations due to the presence of phenoxy free radicals coupling property which is responsible for anti-oxidative action. Lignans also possess antiinflammatory, immunosuppressive activity, and hepatoprotective activity that has been reported in different reviews [91, 92].

Flavonoids

Dietary consumption of lignan-rich foods can prevent a few types of cancers due to the presence of bioactive properties (*e.g.*, breast cancer and colon cancer in post-menopausal women). In the case of chronic lifestyle-related disorders, some researches suggest that lignan use is linked to a lower risk of cardiovascular complications [93].

STILBENES

Stilbenes are non-flavonoids that have health-promoting properties. They are phytoalexins, which are produced in plant tissues in response to fungal attack or abiotic stress such as UV irradiation [94]. Stilbenes are polyphenols made up of two phenyl compounds connected by a two-carbon methylene bridge (Fig. 9) [95]. As per studies, stilbenes are the most common in cowberry, blueberry, lingon berry, and acai berry. Resveratrol is the most well-known stilbene (trans-3,4,5trihydroxystilbene) and found in small quantities in grapes, peanuts, wine, and cranberries. Antioxidative, anticarcinogenic, phytoestrogenic, and cardioprotective activity have all been identified [96]. If structure-activity relationship is considered, the two aromatic rings joined by ethylene moiety present two diastereoisomeric forms, E-1,2-diphenylethylene (trans-configuration) and Z-1,--diphenylethylene (cis-configuration). From the parent structure, several structures are developed by the substitution of substituted hydroxyl-, alkoxy- and glycoside [97]. The structure of stilbene is given here from which the different modifications are conducted for change in the activities. Stilbenes are phenylpropanoids with a 1,2-diphenylethylene backbone. They are the mainly derivatives of the monomeric unit trans-resveratrol (trans-R, 3, 40, 5trihydroxystilbene), although other type of stilbene structures are also found in plants [98]. The most common stilbene, resveratrol, has been shown to have bacteriostatic, estrogenic, and fungistatic properties, prevents tumors in men and is used as growth stimulants in the breeding of animals. Resveratrol is a phytoncide that alters the qualitative and quantitative structure of micro flora in the gastrointestinal tract, similar to antibiotic growth stimulants did previously [99].

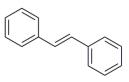


Fig. (9). Stilbenes.

Flavonoids

TANNINS

These are the phenolic compounds with astringent properties. They are mainly abundant in coniferous trees and in flowering plant families. Tannins are present in the stems, wood, bark, leaves, and fruit of several plants, mostly in oak species, sumac and myrobalan. They also found in galls, pathological growths caused by insect attacks [100, 101].

Classification of Tannins on the basis of structure:

There is no carboxylic group in tannins but is weakly acidic in nature because of the multiplicity of phenolic hydroxyls. The hydroxyl group increases the solublity in water. All regulatory authorities classify it as a nonhazardous substance.

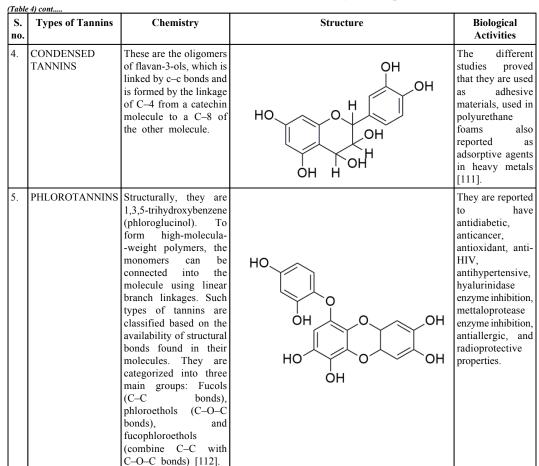
Hydrolysable tannins: In such types of tannins, there is partial or complete esterification of the hydroxyl group by gallic acid [102].

Condensed tannins: These types of tannins are formed by the condensation of phenolic compounds, abundantly found in grapes, pomegranates etc., and have high molecular weights and potential therapeutic activity [103]. They are also known as proanthocyanins. Tannins are non-crystalline substances sparingly soluble in water [104].

Tannins have two to three phenolic hydroxyl groups on a relatively large phenyl ring responsible for these characteristics. Previously, tannins were classified as pyrogallol and catechol type (or catechin) tannins [105]. Two classes were renamed hydrolyzable tannins and condensed tannins as tannin chemistry progressed. The alteration of structure and composition is vital for the pharmacological activity of any target molecule. The previous evidence proved that tannins are high molecular weight compounds due to multiple functional groups in their structure, such as hydroxyls, which provides strong and stable cross-linked association with other molecules, such as carbohydrates or proteins [106]. The tannins are further classified on the basis of chemical composition, which has been given in the tabular form (Table 4):

S. no.	Types of Tannins	Chemistry	Structure	Biological Activities
1.	GALLOTANNINS	Gallotannins are galloyl-based polymers that bind to polyol-, catechin-, or triterpenoid-units, A core molecule, such as glucose, is surrounded by gallic acid units in gallotannins [107].		Different pieces of evidence proved that due to the presence of short one or more polygalloyl chains, they possess antioxidant activity, anti- inflammatory and are used for the treatment of Alzheimer's disease [108].
2.	ELLAGITANNINS	Ellagitannins are polyphenols, and they are characterized by the presence of one or more hexahydroxydiphenoyl moieties, <i>i.e.</i> , esterified to a sugar molecule, generally glucose [109]	но о о о он	Ellagitannins have proved to be highly effective as an antioxidant, anti-adipogenic activity, and high potential cancer prevention [109].
3.	COMPLEX TANNINS	They are formed by the combination of ellagitannin and gallotannin and joined through a glycosidic bond to a flavan-3-ol. Eugenigrandin A and Acutissimin A are complex tannins [110].		Acutissimin A, which is the most common example of ellagitannin, is found to have topoisomerase inhibiting activity [110].

Table 4. Types of tannins with their chemistry and biological activities.



CONCLUSION

Flavonoids

In this chapter, all the most abundant polyphenolic compounds with their structure-activity relationship have been discussed. As we are all aware that the molecular structures are directly related to biological activities, the activities can be modified by the structural modifications of covered natural polyphenols. The analysis of the biological effects of a chemical depends on its molecular structure. The researches have proved that polyphenols are the most promising naturally derived chemical compounds found mainly in fruits, beverages, vegetables, and cereals. The different polyphenolic compounds have been summarized in this chapter with their structures, biological sources and their structural modifications. Polyphenols are the most promising natural metabolites, which being propitious compounds for some common health problems of man and possible certain effects of aging are highly celebrated. Different reviews and data have been collected to summarize the different polyphenolic compounds. The study was aimed to

provide a detailed overview of polyphenolic compounds like phenolic acids, flavonoids, furanocoumarins, stilbenes and tannins content, their structure and relationship along with their biological activity. These compounds are used industrially in the development of various materials, fabrics, food, or medicine. Further, deeper research is needed to explore the chemistry of polyphenolic compounds.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The authors declared that there is no conflict of interest, financial or otherwise.

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REFERENCES

- [1] Pittelli A, Gollücke B, *et al.* Chapter 2 Polyphenols as Supplements in Foods and Beverages: Recent Discoveries and Health Benefits, an Update.Polyphenols: Mechanisms of Action in Human Health and Disease. 2018; 2: pp. 11-8.
- [2] Kasprzak-Drozd K, Oniszczuk T, Stasiak M, Oniszczuk A. Beneficial Effects of Phenolic Compounds on Gut Microbiota and Metabolic Syndrome. Int J Mol Sci 2021; 22(7): 3715. [http://dx.doi.org/10.3390/ijms22073715] [PMID: 33918284]
- Polyphenols: food sources, properties and applications a review. Int J Food Sci Technol 2009; 44: 2512-8.
 [http://dx.doi.org/10.1111/j.1365-2621.2009.02077.x]
- [4] Pandey KB, Rizvi SI. Plant polyphenols as dietary antioxidants in human health and disease. Oxid Med Cell Longev 2009; 2(5): 270-8. [http://dx.doi.org/10.4161/oxim.2.5.9498] [PMID: 20716914]
- [5] Mini-Review A. Front Nutr 2018; 5: 87. Hannah Cory and JosiemerMattei et al.The Role Of Polyphenols In Human Health And Food Systems: [http://dx.doi.org/10.3389/fnut.2018.00087] [PMID: 30298133]
- [6] Bioactivity of phenolic acids: metabolites versus parent compounds: a review. Food Chemistry 2015; 173: 501-13.
- [7] Phenolic Compounds: Functional Properties, Impact ocessing and Bioavailability. 2017. Web of Science
- [8] Sendamangalam V, Choi OK. YoungwooSeo, Don-Shik Kim, Antimicrobial and Antioxidant Activities of Polyphenols against Streptococcus mutans. Free Radic Antioxid 2011; 1(3): 48-55. [http://dx.doi.org/10.5530/ax.2011.3.7]
- [9] P.F. Fox. Significance and applications of phenolic compounds in the production and quality of milk and dairy products: a review. Int Dairy J 2001; 11(3): 103-20.

Flavonoids

[http://dx.doi.org/10.1016/S0958-6946(01)00033-4]

- [10] Polyphenols and Polyphenol-Derived Compounds From Plants and Contact Dermatitis.Polyphenols. Prevention and Treatment of Human Disease 2018; 2: 349-84.
- [11] Chemistry and health effects of furanocoumarins in grapefruit. Journal of Food and Drug Analysis 2017; 25(1): 71-83.
- [12] Bruni R, Barreca D, Protti M, et al. Botanical Sources, Chemistry, Analysis, and Biological Activity of Furanocoumarins of Pharmaceutical Interest. Molecules 2019; 24(11): 2163. [http://dx.doi.org/10.3390/molecules24112163] [PMID: 31181737]
- [13] Cui Q, Du R, Liu M, Rong L. Lignans and Their Derivatives from Plants as Antivirals. Molecules 2020; 25(1): 183.
 [http://dx.doi.org/10.3390/molecules25010183] [PMID: 31906391]
- [14] Wilson R. Cunhaand Jairo Kenupp Bastos et alLignans: Chemical and Biological Properties. Phytochemicals – A Global Perspective of Their Role in Nutrition and Health 2012.
- [15] Suzuki S, Umezawa T, *et al.* Biosynthesis of lignans and norlignans. J Wood Sci 2007; 53(1435): 273-84.
 [http://dx.doi.org/10.1007/s10086-007-0892-x]
- [16] Bastos JK, Albuquerque S, Silva ML. Evaluation of the trypanocidal activity of lignans isolated from the leaves of Zanthoxylum naranjillo. Planta Med 1999; 65(6): 541-4.
- [http://dx.doi.org/10.1055/s-1999-14012] [PMID: 10483375]
- [17] Ovesná Z, Horváthová-Kozics K. Structure-activity relationship of trans-resveratrol and its analogues. Neoplasma 2005; 52(6): 450-5.
 [PMID: 16284688]
- [18] Giacomini E, Rupiani S, Guidotti L, Recanatini M, Roberti M. The Use of Stilbene Scaffold in Medicinal Chemistry and Multi- Target Drug Design. Curr Med Chem 2016; 23(23): 2439-89. [http://dx.doi.org/10.2174/0929867323666160517121629] [PMID: 27183980]
- [19] Rivière C, Pawlus AD, Mérillon JM. Natural stilbenoids: distribution in the plant kingdom and chemotaxonomic interest in Vitaceae. Nat Prod Rep 2012; 29(11): 1317-33.
 [http://dx.doi.org/10.1039/c2np20049j] [PMID: 23014926]
- [20] Murray RH, Mendez J, Brown SA. The Natural Coumarins: Occurrence, Chemistry and Biochemistry. Chichester, UK: Johns Wiley & Sons 1982.
- [21] Lila MA. Anthocyanins and Human Health: An *In Vitro* Investigative Approach. J Biomed Biotechnol 2004; 2004(5): 306-13.
 [http://dx.doi.org/10.1155/S111072430440401X] [PMID: 15577194]
- [22] Khoo HE, Azlan A, Tang ST, Lim SM. Anthocyanidins and anthocyanins: colored pigments as food, pharmaceutical ingredients, and the potential health benefits. Food Nutr Res 2017; 61(1): 1361779. [http://dx.doi.org/10.1080/16546628.2017.1361779] [PMID: 28970777]
- [23] Fraga-Corral M, García-Oliveira P, Pereira AG, et al. Technological Application of Tannin-Based Extracts. Molecules 2020; 25(3): 614. [http://dx.doi.org/10.3390/molecules25030614] [PMID: 32019231]
- [24] Smeriglio A, Barreca D, Bellocco E, Trombetta D. Proanthocyanidins and hydrolysable tannins: occurrence, dietary intake and pharmacological effects. Br J Pharmacol 2017; 174(11): 1244-62. [http://dx.doi.org/10.1111/bph.13630] [PMID: 27646690]
- [25] Okuda T. Novel aspects of tannins. Renewed concept and structure-activity relationships. Curr Org Chem 1999; 3: 609-22.
- [26] Serrano J. Mol Nutr Food Res 2009; 53: 310-29. [http://dx.doi.org/10.1002/mnfr.200900039]

- [27] Kraus TEC, Dahlgren RA, Zasoski RJ. Tannins in nutrient dynamics of forest ecosystems-a review. Plant Soil 2003; 256: 41-6. [http://dx.doi.org/10.1023/A:1026206511084]
- [28] Hardingand Liang-Jiao Xue SA, *et al.* Condensed tannin biosynthesis and polymerization synergistically condition carbon use, defense, sink strength and growth in Populus. Tree Physiol 2013; 00: 1-12.
- [29] Anantharaju PG, Gowda PC, Vimalambike MG, Madhunapantula SV. An overview on the role of dietary phenolics for the treatment of cancers. Nutr J 2016; 15(1): 99. [http://dx.doi.org/10.1186/s12937-016-0217-2] [PMID: 27903278]
- [30] Saibabu V, Fatima Z, et al. Therapeutic Potential of Dietary Phenolic Acids. Advancements in Pharmacological and Pharmaceutical Sciences. 2015; pp. 1-10. [http://dx.doi.org/10.1155/2015/823539]
- [31] Tuck KL, Tan HW, Hayball PJ. Synthesis of tritium-labeled hydroxytyrosol, a phenolic compound found in olive Oil. J Agric Food Chem 2000; 48(9): 4087-90. [http://dx.doi.org/10.1021/jf0004681] [PMID: 10995319]
- [32] Scalbert A, Manach C, Morand C, Rémésy C, Jiménez L. Dietary polyphenols and the prevention of diseases. Crit Rev Food Sci Nutr 2005; 45(4): 287-306. [http://dx.doi.org/10.1080/1040869059096] [PMID: 16047496]
- [33] Hussein I. Abdel-ShafyAnd Mona S.M. Mansour. Polyphenols: Properties, Occurrence, Content in Food and Potential Effects. Environ Sci & Engg 2017; 6: 232-63.
- [34] Rebecca J, Robbins J. Phenolic Acids in Foods: An Overview of Analytical Methodology Agric. Food Chem 2003; 51(10): 2866-87.
 [http://dx.doi.org/10.1021/jf026182t]
- [35] Alam MA, Subhan N, Hossain H, et al. Hydroxycinnamic acid derivatives: a potential class of natural compounds for the management of lipid metabolism and obesity. Nutr Metab (Lond) 2016; 13(27): 27. [http://dx.doi.org/10.1186/s12986-016-0080-3] [PMID: 27069498]
- [36] Hydroxycinnamic Acids and Their Derivatives: Cosmeceutical Significance, Challenges and Future Perspectives, a Review. Molecules 2017; 22(2): 281-96.
- [37] Tomas-Barberan FA, Clifford MN. Dietary hydroxybenzoic acid derivatives and their possible role in health protection. J Sci Food Agric 2000; 80: 1024-32.
- [38] Walter S. Schroeder, Linda Ghobrial and Pritesh J. Gandhi.Possible mechanisms of drug-induced aspirin and clopidogrel resistance. J Thromb Thrombolysis 2006; 22: 139-50. [http://dx.doi.org/10.1007/s11239-006-8670-y]
- [39] Pereira DM, Valentao P, Pereira JA, Andrade PB. Phenolics: From chemistry to biology. Molecules 2009; 14: 2202-11.
 [http://dx.doi.org/10.3390/molecules14062202]
- [40] Seo YK, Kim SJ, Boo YC, Baek JH, Lee SH, Koh JS. Effects of p-coumaric acid on erythema and pigmentation of human skin exposed to ultraviolet radiation. Clin Exp Dermatol 2011; 36(3): 260-6. [http://dx.doi.org/10.1111/j.1365-2230.2010.03983.x] [PMID: 21198798]
- [41] Hollman PCH, Katan MB. Dietary flavonoids: intake, health effects and bioavailability. Food Chem Toxicol 1999; 37(9-10): 937-42.
 [http://dx.doi.org/10.1016/S0278-6915(99)00079-4] [PMID: 10541448]
- [42] Matsumoto T, Kaneko A, Koseki J, Matsubara Y, Aiba S, Yamasaki K. Pharmacokinetic Study of Bioactive Flavonoids in the Traditional Japanese Medicine Keigairengyoto Exerting Antibacterial Effects against Staphylococcus aureus. Int J Mol Sci 2018; 19(2): 328. [http://dx.doi.org/10.3390/ijms19020328] [PMID: 29360768]
- [43] Wang H, Murphy PA. Isoflavone content in commercial soybean foods. J Agric Food Chem 1994; 42:

1666-73. [http://dx.doi.org/10.1021/jf00044a016]

- [44] Jacob Vaya, Tavori Hagai, Soliman Khatib. Structure-Activity Relationships of Flavonoids. Current Organic Chemistry 2011; 15: 2641-57.
- [45] PrithvirajKarak.Biological Activities of Flavonoids: An Overview. Int J Pharm Sci Res 2019; 10(4): 1567-74.
- [46] Yankep E, et al. TheMillettia of Cameroon. O-Geranylated isoflavones and a 3-phenylcoumarin from Millettiagriffoniana. Phytochemistry 1998; 49: 2521. [http://dx.doi.org/10.1016/S0031-9422(98)00392-6]
- [47] Oyvind M. Andersen and Kenneth R. Markham. Flavonoids Chemistry, Biochemistry and Applications from Taylor and Francis 2006; pp. 1-1199.
- [48] Leutert T, von Arx E. Preparative medium-pressure liquid chromatography. Journal o chromatography 1984; 292: 333.
- [49] Adell J, Barbera O, Marco JA. Flavonoid glycosides from Anthyllis sericea. Phytochemistry 1988; 27: 2967.
 [http://dx.doi.org/10.1016/0031-9422(88)80698-8]

- [50] Zogg GC, Nyiredy Sz, Sticher O. Operating conditions in preparative medium pressure liquid chromatography (MPLC). II. Influence of solvent strength and flow rate of the mobile phase, capacity and dimensions of the column. J Liq Chromatogr 1989; 12: 2049.
- [51] Zhou B-N, Blasko G, Cordell GA. Iternanthin, a C-glycosylated flavonoid from Alternanthera philoxeroides. Phytochemistry 1988; 27: 3633. [http://dx.doi.org/10.1016/0031-9422(88)80781-7]
- [52] Victor R. Preedy and ShermaZibadi.Polyphenols. Mechanisms of Action in Human Health and Disease from Academic Press 2018; 2: 70.
- [53] Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. Polyphenols: food sources and bioavailability. Am J Clin Nutr 2004; 79(5): 727-47. [http://dx.doi.org/10.1093/ajcn/79.5.727] [PMID: 15113710]
- [54] Bioactive flavonoids in medicinal plants: Structure, activity and biological fateAsian. J Pharm Sci 2018; 13(1): 12-23.
- [55] Tungmunnithum D, Thongboonyou A, Pholboon A, Yangsabai A. Flavonoids and Other Phenolic Compounds from Medicinal Plants for Pharmaceutical and Medical Aspects: An Overview. Medicines (Basel) 2018; 5(3): 1-16. [http://dx.doi.org/10.3390/medicines5030093] [PMID: 30149600]
- [56] Heim KE, Tagliaferro AR, Bobilya DJ. Flavonoid antioxidants: chemistry, metabolism and structureactivity relationships. J Nutr Biochem 2002; 13(10): 572-84. [http://dx.doi.org/10.1016/S0955-2863(02)00208-5] [PMID: 12550068]
- [57] Alzand KI, Mohamed MA. Flavonoids: Chemistry, Biochemistry and Antioxidant activity. J Pharm Res 2012; 5(8): 4013-40.
- [58] Weston LA, Mathesius U. Flavonoids: their structure, biosynthesis and role in the rhizosphere, including allelopathy. J Chem Ecol 2013; 39(2): 283-97. [http://dx.doi.org/10.1007/s10886-013-0248-5] [PMID: 23397456]
- [59] Brusselmans K, Vrolix R, Verhoeven G, Swinnen JV. Induction of cancer cell apoptosis by flavonoids is associated with their ability to inhibit fatty acid synthase activity. J Biol Chem 2005; 280(7): 5636-45.
 [http://dx.doi.org/10.1074/jbc.M408177200] [PMID: 15533929]
- [60] Kumar S, Pandey AK. Chemistry and Biological Activities of Flavonoids: An Overview. The Scientific World Journal. Hindawi Publishing Corporation. Scientific World Journal 2013; •••: 162750:1-.

- [61] Erlejman AG, Verstraeten SV, Fraga CG, Oteiza PI. The interaction of flavonoids with membranes: potential determinant of flavonoid antioxidant effects. Free Radic Res 2004; 38(12): 1311-20. [http://dx.doi.org/10.1080/10715760400016105] [PMID: 15763955]
- [62] Agati G, Azzarello E, Pollastri S, Tattini M. Flavonoids as antioxidants in plants: location and functional significance. Plant Sci 2012; 196: 67-76. [http://dx.doi.org/10.1016/j.plantsci.2012.07.014] [PMID: 23017900]
- [63] Andersen ØM, Markham KR. Flavonoids: Chemistry, Biochemistry and Applications. Boca Raton: CRC Press 2006.
- [64] Zandi K, Teoh BT, Sam SS, Wong PF, Mustafa MR, Abubakar S. Antiviral activity of four types of bioflavonoid against dengue virus type-2. Virol J 2011; 8: 560. [http://dx.doi.org/10.1186/1743-422X-8-560] [PMID: 22201648]
- [65] Wang A, Zhang F, Huang L, et al. New progress in biocatalysis and biotransformation of flavonoids. J Med Plants Res 2010; 4(10): 847-56.
- [66] Agati G, Azzarello E, Pollastri S, Tattini M. Flavonoids as antioxidants in plants: location and functional significance. Plant Sci 2012; 196: 67-76. [http://dx.doi.org/10.1016/j.plantsci.2012.07.014] [PMID: 23017900]
- [67] Huang Z, Fang F, Wang J, Wong CW. Structural activity relationship of flavonoids with estrogenrelated receptor gamma. FEBS Lett 2010; 584(1): 22-6. [http://dx.doi.org/10.1016/j.febslet.2009.11.026] [PMID: 19914244]
- [68] Ross JA, Kasum CM. Dietary flavonoids: bioavailability, metabolic effects, and safety. Annu Rev Nutr 2002; 22: 19-34. [http://dx.doi.org/10.1146/annurev.nutr.22.111401.144957] [PMID: 12055336]
- [69] Pietta PG. Flavonoids as antioxidants. J Nat Prod 2000; 63(7): 1035-42. [http://dx.doi.org/10.1021/np9904509] [PMID: 10924197]
- [70] Ofman DJ, et al. Flavonoid profiles of New Zealand kauri and other species of Agathis. Phytochemistry 1995; 38: 1223.
 [http://dx.doi.org/10.1016/0031-9422(94)00783-P]
- [71] Bors W, Heller W, Michel C, Saran M. Flavonoids as antioxidants: determination of radicalscavenging efficiencies. Methods Enzymol 1990; 186: 343-55. [http://dx.doi.org/10.1016/0076-6879(90)86128-I] [PMID: 2172711]
- [72] Szabados-Furjesi P, Pajtas D, Barta A, *et al.* Synthesis, *in Vitro* Biological Evaluation, and Oxidative Transformation of New Flavonol Derivatives: The Possible Role of the Phenyl-N,N-Dimethylamino Group. Molecules 2018; 23(12): 3161.
 [http://dx.doi.org/10.3390/molecules23123161] [PMID: 30513682]
- [73] Mishra A, Kumar S, Bhargava A, Sharma B, Pandey AK. Studies on in vitro antioxidant and antistaphylococcal activities of some important medicinal plants. Cell Mol Biol 2011; 57(1): 16-25. [PMID: 21366958]
- [74] Pandey AK, Mishra AK, Mishra A, Kumar S, Chandra A. Therapeutic potential of C. zeylanicum extracts: an antifungal and antioxidant perspective. Int J Biol Med Res 2010; 1: 228-33.
- [75] Wang Y, Chen S, Yu O. Metabolic engineering of flavonoids in plants and microorganisms. Appl Microbiol Biotechnol 2011; 91(4): 949-56.
 [http://dx.doi.org/10.1007/s00253-011-3449-2] [PMID: 21732240]
- [76] Owen RW, Haubner R, Hull WE, *et al.* Isolation and structure elucidation of the major individual polyphenols in carob fibre. Food Chem Toxicol 2003; 41(12): 1727-38.
 [http://dx.doi.org/10.1016/S0278-6915(03)00200-X] [PMID: 14563398]
- [77] El-Sawy ER, Abdelwahab AB, Kirsch G. Synthetic Routes to Coumarin(Benzopyrone)-Fused Five-Membered Aromatic Heterocycles Built on the α-Pyrone Moiety. Part 1: Five-Membered Aromatic

Rings with One Heteroatom. Molecules 2021; 26(2): 483. [http://dx.doi.org/10.3390/molecules26020483] [PMID: 33477568]

- [78] Myers RB, Parker M, Grizzle WE. synthesis of stilbene coumarin hybrid compounds & compounds has been reported as anticancer activity. J Cancer Res Clin Oncol 1994; 12: 11-23. [http://dx.doi.org/10.1007/BF01377115]
- [79] Lacy A, O'Kennedy R. Studies on coumarins and coumarin-related compounds to determine their therapeutic role in the treatment of cancer. Curr Pharm Des 2004; 10(30): 3797-811. [http://dx.doi.org/10.2174/1381612043382693] [PMID: 15579072]
- [80] Dalla Via L, Mammi S, Uriarte E, et al. New furan side tetracyclic allopsoralen derivatives: synthesis and photobiological evaluation. J Med Chem 2006; 49(14): 4317-26. [http://dx.doi.org/10.1021/jm058032q] [PMID: 16821791]
- [81] Hung W-L, Suh JH, Wang Y. Chemistry and health effects of furanocoumarins in grapefruit. Yao Wu Shi Pin Fen Xi 2016; •••: 1-13.
 [PMID: 28911545]
- [82] Lourdes Santana et al. Coumarins. An Important Class of Phytochemicals. Phytochemicals. Isolation, Characterisation and Role in Human Health 2015; pp. 1-28.
- [83] Kontogiorgis C. HadjipavlouLitina DJ.Biological Evaluation of Several Coumarin Derivatives Designed as Possible Anti-inflammatory/Antioxidant Agents. Journal of Coumarins. An Important Class of Phytochemicals. Enzyme Inhibition and Medicinal Chemistry 2003; 18(1): 63-9. [http://dx.doi.org/10.1080/1475636031000069291]
- [84] Fylaktakidou KC, Hadjipavlou-Litina DJ, Litinas KE, Nicolaides DN. Natural and synthetic coumarin derivatives with anti-inflammatory/ antioxidant activities. Curr Pharm Des 2004; 10(30): 3813-33. [http://dx.doi.org/10.2174/1381612043382710] [PMID: 15579073]
- [85] Peng XM, Damu GLV, Zhou C. Current developments of coumarin compounds in medicinal chemistry. Curr Pharm Des 2013; 19(21): 3884-930. [http://dx.doi.org/10.2174/1381612811319210013] [PMID: 23438968]
- [86] Liang S, Shen Y-H, Tian J-M, Wu Z-J, Jin H-Z, Zhang W-D, et al. Three New Dicou□ marins from Daphne feddei. HCA 2009; 92(1): 133-8. [http://dx.doi.org/10.1002/hlca.200800232]
- [87] Sugahara T, Kakinuma Y, et al. The Structure-Activity Relationships of Flaxseed Lignan, Secoisolariciresinol. Interdisciplinary Studies on Environmental Chemistry—Biological Responses to Chemical Pollutants 2008; pp. 263-8.
- [88] Wilson R. Cunha and Jairo Kenupp Bastos et alLignans: Chemical and Biological Properties. Phytochemicals – A Global Perspective of Their Role in Nutrition and Health 2012; pp. 213-33.
- [89] Greenberg M, Dodds M, Tian M. Naturally occurring phenolic antibacterial compounds show effectiveness against oral bacteria by a quantitative structure-activity relationship study. J Agric Food Chem 2008; 56(23): 11151-6. [http://dx.doi.org/10.1021/jf8020859] [PMID: 19007234]
- [90] Charlton JL. Antiviral activity of lignans. J Nat Prod 1998; 61(11): 1447-51. [http://dx.doi.org/10.1021/np9801362] [PMID: 9834179]
- [91] Adlercreutz H. Lignans and human health. Crit Rev Clin Lab Sci 2007; 44(5-6): 483-525.
 [http://dx.doi.org/10.1080/10408360701612942] [PMID: 17943494]
- [92] McRae DW, Towers NGH. Biological activities of lignans. Phytochemistry 1984; 23(6): 1207-20. [http://dx.doi.org/10.1016/S0031-9422(00)80428-8]
- [93] Pan JY, Chen SL, Yang MH, Wu J, Sinkkonen J, Zou K. An update on lignans: natural products and synthesis. Nat Prod Rep 2009; 26(10): 1251-92.
 [http://dx.doi.org/10.1039/b910940d] [PMID: 19779640]

- [94] Chong J, Poutaraud A, Hugueney P. Metabolism and roles of stilbenes in plants. Plant Sci 2004; 177(3): 143-55.
 [http://dx.doi.org/10.1016/j.plantsci.2009.05.012]
- [95] Chou Ya-Chun, Chi-Tang Ho. Curr Pharmacol Rep 2018; 4: 202-9. [http://dx.doi.org/10.1007/s40495-018-0134-5]
- [96] Alonso F, Riente P, Yus M. Wittig-type olefination of alcohols promoted by nickel nanoparticles: synthesis of polymethoxylated and polyhydroxylated stilbenes. Eur J Org Chem 2009; 34: 6034-42. [http://dx.doi.org/10.1002/ejoc.200900951]
- [97] Reinisalo M, Karlund A, Koskela A, Kaarniranta K, Karjalainen RO. Polyphenol Stilbenes: Molecular Mechanisms of Defence against Oxidative Stress and Aging-Related Diseases. Oxidative Medicine and Cellular Longevity. 2015.
- [98] Arnaud Courtois. JosepValls. Tristan Richard. Ste'phanieKrisa. A review of dietary stilbenes: sources and bioavailability. Phytochemistry Journal Elsevier 2018; 17: 1007-29.
- [99] Frombaum M, Le Clanche S, Bonnefont-Rousselot D, Borderie D. Antioxidant effects of resveratrol and other stilbene derivatives on oxidative stress and *NO bioavailability: Potential benefits to cardiovascular diseases. Biochimie 2012; 94(2): 269-76. [http://dx.doi.org/10.1016/j.biochi.2011.11.001] [PMID: 22133615]
- [100] Arbenz A, Avérous L. Chemical modification of tannins to elaborate aromatic biobased macromolecular architectures. Green Chem 2015; 17(5): 2626-46. [http://dx.doi.org/10.1039/C5GC00282F]
- [101] Tannins are Astringent. J Pharmacogn Phytochem 2012; 1(3): 45-50.
- [102] Funatogawa K, Hayashi S, Shimomura H, et al. Antibacterial activity of hydrolyzable tannins derived from medicinal plants against Helicobacter pylori. Microbiol Immunol 2004; 48(4): 251-61. [http://dx.doi.org/10.1111/j.1348-0421.2004.tb03521.x] [PMID: 15107535]
- [103] Pizzi A. Tannins: Prospectives and Actual Industrial Applications. Biomolecules 2019; 9(8): 1-30. [http://dx.doi.org/10.3390/biom9080344] [PMID: 31387308]
- [104] Roux DG, Ferreira D, Hundt HKL, Malan E. Structure, stereochemistry and reactivity of condensed tannins as basis for their extended industrial application. J Appl Polym Sci 1975; 28: 335-53.
- [105] Okuda T, Mori K, Hatano T. Relationship of the structures of tannins to the binding activities with hemoglobin and methylene blue. Chem Pharm Bull (Tokyo) 1985; 33(4): 1424-33. [http://dx.doi.org/10.1248/cpb.33.1424] [PMID: 4042219]
- [106] Buziashvili Sh, Komissarenko NF, Kovalev IP, Gordienko VG, Kolesnikov DG, et al. The structure of gallotannins. Chem Nat Compd 1973; 9: 752-5. [http://dx.doi.org/10.1007/BF00565801]
- [107] Tahiri Sylla and StiphaneQuideau.Gallotannins and Tannic Acid: First Chemical Syntheses and In Vitro Inhibitory Activity on Alzheimer's Amyloid b-Peptide Aggregation. Angew Chem Int Ed 2015; 54: 8217-21. [http://dx.doi.org/10.1002/anie.201411606]
- [108] Okuda T, Yoshida T, Hatano T. Correlation of oxidative transformations of hydrolyzable tannins and plant evolution. Phytochemistry 2000; 55(6): 513-29. [http://dx.doi.org/10.1016/S0031-9422(00)00232-6] [PMID: 11130661]
- [109] Landete JM. Ellagitannins, ellagic acid and their derived metabolites: A review about source, metabolism, functions and health. Food Res Int 2011; 44: 1150-60. [http://dx.doi.org/10.1016/j.foodres.2011.04.027]
- [110] Quideau S, Jourdes M, Saucier C, Glories Y, Pardon P, Baudry C. DNA topoisomerase inhibitor acutissimin a and other flavano-ellagitannins in red wine. Angew Chem Int Ed 2003; 42(48): 6012-4. [http://dx.doi.org/10.1002/anie.200352089] [PMID: 14679557]

Flavonoids

- [111] Ohara S. Chemistry and Utilization of Condensed Tannins from Tree Barks. Jpn Agric Res Q 1998; 28(1): 70-8.
- [112] Sonani RR, Rastogi RP, Madamwar D. Natural Antioxidants From Algae: A Therapeutic Perspective. Algal Green Chemistry. Recent Progress in Biotechnology 2017; pp. 91-120. [http://dx.doi.org/10.1016/B978-0-444-63784-0.00005-9]

Biologically Active Compounds and Structure-Activity Relationship

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Abstract: Naturally occurring compounds are found to be the most prominent and effective biological active compounds against various diseases. The majority of drugs approved between 1983 to 1994 are derived from natural products. Still today, the majority of pharmaceutical laboratories are hoping to get new drug candidates from natural resources. The traditional method of drug discovery from naturally occurring compounds has been upgraded by using advanced computer-based drug discovery.

In drug discovery, the initial efforts are to know the relationship between the biological activity of natural compounds and their chemical structures. To be precise, the method of structure-activity relationship aims to recognize the basic structural component responsible for biological activity.

The computational modeling drug discovery using various tools plays a major role in identifying the lead compounds. In this method, three major ways are utilized to understand the structure-activity relationship.

The foremost one is the Quantitative Structure-Activity Relationship (QSAR). In this method, the relationship was established using regression techniques between the 'Predictor Variable (X)' with the potency of the 'Response Variable (Y)'. The predictor variables are molecular descriptors, while the response variables represent the biological activities of the molecules against the selected diseases. If the response variable represents the chemical property, in that case, the model is called as Quantitative Structure-Property Relationship (QSPR).

The second method is called "Inhibition Studies". In this process, the designed chemical entity is docked to the targeted enzyme using docking software. The basic principle of this method is the executive competitive inhibition between the natural inhibitor and the designed chemical entity. The law of thermodynamic is used to understand the best-docked chemical entity by obtaining the value of binding energy (ΔG kcal/mole) due to the complex formation between the chemical moiety and target enzyme.

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The third approach is very advanced and more accurate. It is called "The drug discovery using Artificial Neural Network". This is the recent technique adapted by major international pharmaceutical research laboratories. In this method, the neural network is designed and trained to identify the potent chemical compound against a particular disease. The designing of the network can be achieved using the chemical properties of a neuron, and output is related to the biological activity.

This chapter discussed all three methods in detail, along with examples. It also provides the practical procedure to use available computational tools.

The final aim of this chapter is not only to provide the theoretical background of drug discovery using structure-activity relationships but also to provide practical methods.

INTRODUCTION

Naturally occurring compounds are found to be the most prominent and effective biological active compounds against various diseases. Medicines originated from plants and other living organisms were initially used in the crude form [1, 2]. With the progress of the science and isolation techniques, the active components were isolated from natural sources and used as a drug. In the early 19th century, morphine was extracted from opium. Further advancement of techniques provided the method to isolate cocaine, codeine, digitoxin, and quinine [2, 3].

It is well documented that the drugs originated from natural products gives useful information about the possible process for the drug development. It is also reported that most drugs approved between 1983 to 1994 are derived from natural products. The importance of the drug from natural products is still at the highest rate even today, as 11% of the 252 essential drugs are originated from flowering plants. It is also reported that out of 175 cancer drugs, 85 are directly or indirectly originated from natural products.

Still today, the majority of pharmaceutical laboratories hope to get new drug candidates from natural resources. The traditional method of drug discovery from naturally occurring compounds has been upgraded by using advanced computer-based drug discovery. The major techniques are QSAR, In-Silico Docking, Molecular Dynamics, High-throughput Screening, *etc.*

The main philosophy behind computational drug discovery is to identify the structure-activity relationship between a lead compound and a target enzyme. The lead compound, also called a small molecule, has to fit into an enzyme to inhibit it.

The advanced computational chemistry and high-throughput screening help to identify the lead compound in a very short period. This also helps in getting drugs in the market quite early.

THE ORIGIN OF QSAR& QSPR

QSAR stands for Quantitative Structure-Activity Relationship and QSPR stands for Quantitative Structure-Property Relationship. These are predictive methods and uses statistical methodology to evaluate the drug-likeness of the small molecules against a particular target (generally enzyme). These methods are initially put forth by Hansch [4, 5] and then Free [6].

QSPR refers to the physical properties of the molecule, whereas, in the case of QSAR, the molecular properties are related to the biological activity of molecules using the regression analysis.

The mathematics of QSAR models is the relationship of free energy as depicted in the Hammett equation. This equation defines the relationship between dissociation constant and electronic properties of acid or bases [7 - 9].

The equation is defined as;

$$\log \frac{K}{K_o} = \rho \log \frac{K}{K_o}$$
(1)

In equation 1, a set of substituted aromatic acids are represented by K and K'. Unsubstituted acids are represented by K_{θ} and K_{θ}' . ρ is the slope of the best fit line for a graph fitted to observed constant values. logK/K0 describes the substituent.

Initially, Hansch tried to formulate the QSAR models based on the Hammett parameter. This model did not provide acceptable results. Then, he accommodates other parameters such as molecular size, lipophilicity, *etc*.

The main aim of the QSAR methodology is to understand the relationship between observed properties and structural features of the molecule. In this case, one lead compound is selected, and by substituting on this leads compound, numerous molecules are grouped. This group of compounds having a common base structure is called a set of molecules. Also, the properties of these molecules are set together. By considering a set of molecules (having various substituents and properties), a predictive model is developed using statistics. This derived model (an equation) is used to predict the possible biological activity of newly designed or synthesized molecules.

The main strategy is to convert or represent the structure of the molecular descriptors. The descriptors are nothing but the representation of molecular structure in numerical form.

The descriptors can be from simple physical properties like molecular weight or complex forms from partition coefficient, reflective index, *etc*.

Using a defined set of descriptors; a QSAR model can be built. In the QSAR model, the relationship is defined using the regression analysis between the descriptors and the biological activity.

It was Hansch who developed the first QSAR model using a linear relationship between the descriptors and biological activity. Linear models are commonly used. Nowadays, nonlinear regression and algorithmic techniques are also used to define the relationship between descriptors and molecular activities.

The recent advancement in the field of Artificial Intelligence, particularly pattern recognition and machine learning also been used to develop QSAR models [9 - 11]. The development in the QSAR modeling is achieved with the develop ment in the fields of computational algorithms, mathematics, and statistics.

COMPUTATIONAL CHEMISTRY METHODS FOR DRUG DISCOVERY

Computational chemistry, the recent developing science, helps in exploring and evaluating molecular properties. The computational techniques are generally used when it is difficult to evaluate the molecular properties in laboratories. The techniques are also used in collaboration with the experimental data to confirm the possible mathematical modeling designed for the exploration of molecular properties [11].

The computational chemistry includes Molecular modeling, Computer-aided molecular design, Chemical database, and organic synthesis design. The scope is also extended to the Computer-aided drug design.

Molecular modeling is to calculate the properties of molecules in-silico. Several properties like UV Spectra, IR spectra, dipole moments, the heat of formation, *etc.* can be calculated.

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Computational chemistry, in a broad sense, can be categorized into Classical mechanics and Quantum mechanics. Molecular mechanics is a method derived from classical mechanics, where it uses the laws of classical physics. The laws are applied to molecular nuclei, where it doesn't consider the electrons.

In the case of Quantum mechanics, the Schrödinger equation is applied to describe the molecule with an explicit treatment of electronic structure. This method is divided into two classes: Ab initio and semi-empirical. Fig. (1) explains the branches of computational chemistry methods [10, 11].

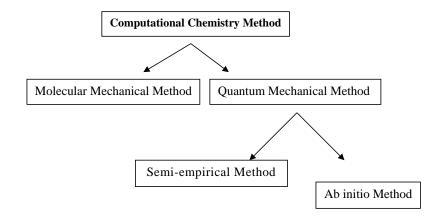


Fig. (1). Computational Chemistry Method [11].

A. Molecular mechanics

Molecular mechanics is a theoretical method to calculate the various properties of molecules, such as dipole moment, the heat of formation, *etc.* In this method, the atoms are assumed as spheres and bonds are like springs. The deformation of the spring is related to the stretch, bend, and twist of the bond as per Hooke's law. The non-bonded atoms interact using various forces like Van der Waals forces of attraction, the static force of attraction, and repulsion forces. The total potential energy is calculated using the following equation.

Potential Energy = Bending Energy + Torsion energy +Stretching Energy +Non-Bonded Interactionenergy.

In the case of structure-based drug discovery, the initial step is to design a 3-D model of a small molecule. Numbers of software are available in the market and

the majority of them use Molecular mechanics for finalizing the 3-D geometry of a molecule. The detailed theory of molecular mechanics is beyond the scope of this book.

B. Ab initio

Ab initio is a theoretical technique, which uses purely theoretical data for evaluating the properties of the system under consideration.

The ab initio calculations are executed using the computational programs to find the exact solution. It starts with the approximations with some magnitude and then using iterative techniques the approximations magnitude finally reaches a very small value. It is observed that for the small molecules, this technique gives good qualitative results with a very small number of iterations. The ab initio calculation uses huge computer memory and time, but due to advancements in the field of computer processors and memory chips, the calculations are faster and cheaper.

C. Semiempirical methods

The semiempirical method is the fastest method to evaluate the potential energy of the molecule. In this method, certain information is approximated or omitted and the error generated due to omission is corrected with parameterization. The parameters are used in curve fitting to bring the obtained results very near to the experimental data.

Semiempirical is the fastest tool to evaluate molecular properties, but sometimes it shows erratic results. It all depends on the selection of parameters of respective molecules. If the selected parameters from the database are of the same molecule which is under study, then the result may be very near to the acceptable limit, else, the obtained results may be of poor quality.

Semiempirical is widely used in organic chemistry for the calculation of geometry optimization or molecular properties. It also provides very good results for organic molecules as there are very few elements used in organic molecules and their parameterization are established with higher accuracy.

STRUCTURE-BASED DRUG DESIGN

Structure-based drug discovery is the new technique. The major problem in classical drug discovery is of time. This hurdle is removed by introducing computer-based drug discovery. Advanced computer programs are used to

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understand how small a molecule is interacting with the active site of the target protein. Before learning the concept and procedure of structure-based drug design, it is wise to understand the meaning of certain keywords which are commonly used in the literature. Table 1 shows the list of certain keywords commonly used in the literature regarding structure-based drug design.

Target Molecule	Generally the Biomolecule like DNA, Protein, or Specific Enzyme.
Active site	The pocket of an enzyme where the reaction is taking place. Generally, the lead compound interacts with an enzyme in this pocket.
3-D structure of an enzyme	Digital 3-D structure of enzyme, protein, or DNA. The structure is identified using X- Ray diffraction or NMR techniques. The structure is available from various databanks. 3-D protein structures are deposited at http://www.rcsb.org. Also possible to get it downloaded in various digital formats. A commonly used file format is PDB.
The ligand or Small molecule or drug molecule	A molecule that inhibits the target (DNA, Protein, Enzyme).
Complex	When a small molecule or ligand interacts with the target and making complex. It is also called a ligand-receptor complex or drug-receptor complex.
Lead Compound	It is a chemical compound that shows the biological or drug-likeness property. It initiates the development of new drugs by designing the new chemical compound by substituting the functional groups in the lead molecules.

Table 1. Basic terms used in structure-based drug	discovery [10, 11].
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Biomolecules like DNA, proteins, or enzymes interact with small molecules to carry the functional processes. The functional site of interaction is an active site of the target compound. To study forces responsible for docking into a target site by mimicking the natural ligand is the aim of structure-based drug design [10]. Based on conformations of the ligand and their interaction with the target compound, new chemical compounds can be designed, which may have a better binding ability with the target compound.

All the drug design techniques aim to visualize the designed drug docked into the target macromolecule, also to analyze the forces involved in their interactions. Visualization and analysis can reveal the minute differences in the binding between the two successive members of series of drug compounds and the conformational changes induced in the target molecule as a result of binding [12].

3-D structure of the functional site of the receptor compound can be explored and the topographies of complementary surfaces can be studied to determine the affinity of chemical molecules. Every drug is target-specific and specificity depends upon the physiochemical properties of the drug and the receptor molecule. The drug-receptor complex is the result of intermolecular forces exerted by the drug molecule.

Based on the architecture of the active site of the enzyme and the study of physiochemical interactions that stabilize the complex, a new ligand with better binding affinity and shape is designed.

Structure-based drug design has already identified several drugs, which are now in the stages of clinical trials. Some of the worth mentioning inhibitors are thymidylate synthetase as an anti-tuberculosis inhibitor, thrombin-based anti-coagulants, Ace inhibitors, HIV protease inhibitors, *etc.*

A. In-silico drug discover

In-silico drug design is the technique, where receptor biomolecule is inhibited with small molecule using a computer-based model system. Inhibitor-based design depends on the architecture of the active site of the target biomolecule.

In this technique, the 3-D structure of the receptor is used as a target, and a library of small molecules is subjected to inhibit the target using computer software [11]. The score function is calculated using semi-empirical or *ab initio* equations. The score function provides information about the best-suited molecules to inhibit the target compound.

The success of the in-silico drug design is due to the crystallography and NMR techniques, which can elucidate the unknown geometry of the target receptor. Also, several computer-based software which helps in inhibiting small molecules to the target receptor. Commonly used drug discovery computer programs (software) are listed in Table 2.

Name	Description	
AutoDock	Ligand – Protein docking.	[12,13]
Schrodinger	Ligand-receptor docking.	[14,15]
GOLD	Protein-ligand docking.	[16,17]
BioSuite	Genome analyzing and sequence analyzing	[18,19]
Maestro	Molecular modeling analysis	[20,21]
ArgusLab	rgusLab Molecular docking calculations and molecular modeling package	
GRAMM	RAMM Protein-protein docking and protein-ligand docking	
SYBYL-X Suite	Molecular modeling and ligand based design	[26,27]

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(Table 4) cont					
Name	Description	References			
Sanjeevini	Predict protein-ligand binding affinity				
PASS	Create and analysis of SAR models	[30,31]			

B. Stages Involved in Structure-based Drug Design

The structure-based drug design is a procedural method. The procedure depends on the software used for the docking. Following are the common steps that need to be executed [32].

The first step is to identify the site of action of the drug on the receptor (biomolecule). The details of the active site like domain structure, secondary and tertiary structural arrangements, and loop architecture give the constitution of the active site. Similarly, the ionic, hydrophobic, and solvent interactions are also understood.3-D structures of biomolecules are available from the online database. A few among them are Protein databank (PDB) [32 - 34], Structural classification of protein (SCOP), Swiss-model, ModBase, *etc*.

From these databases, the 3-D crystal structure of the biomolecule can be downloaded. Generally, this file is in PDB format (electronic file extension). This file may contain certain information which is required to be corrected before executing the next step. The general procedure is to remove all water molecules and hetero-atoms from this file.

Once the PDB file is corrected, it is advisable to sort out the active site. This is the location or motif where drug molecules will interact. The easiest way is to find out the natural inhibitor in this 3-D file. The location of the natural inhibitor is treated as the active site of the target biomolecule. This task can be executed with the help of drug discovery software. The advanced software automatically searches this location, whereas, in the case of amateur software, a user has to sort it out [35].

Fig. (2) shows the 3-D structure of Cyclin Dependent Kinase (CDK4) (PDB Code: 1GII) having 1-(5-oxo-2,3,5,9b-tetrahydro-1h-pyrrolo[2,1- a]isoindol-9-yl)- 3-pyridin-2-yl-urea as a natural inhibitor [36].

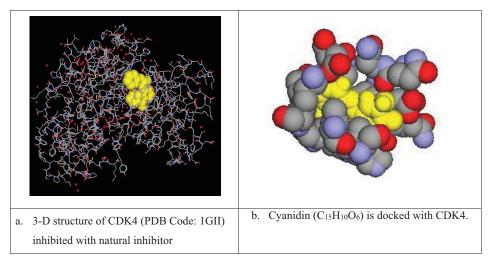


Fig. (2). CDK4 enzyme is docked with Cyanidin (Naturally occurring compound) [36].

The second step is to identify a potent pharmacophore or a lead compound that can bind to the active site of the target molecules in competition with a biologically available protein inhibitor (Natural inhibitor). A lead compound can be selected by screening available libraries of compounds or designing compounds using the software. The market is flooded with numerous software helping in the design of new chemical compounds. A few among them are ChemDraw, ChemSketch, *etc.* Fig. (3) shows the 2-D and 3-D structure of Cyanidin designed using ChemDraw.

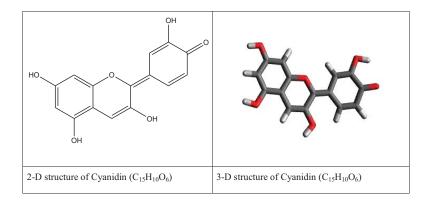


Fig. (3). 2-D and 3-D structure designed using ChemDraw and Chem3D software [36].

The third step is the process of docking of ligand into the active site of a target compound. For this purpose number of software are available. Table 2 shows the list of software that can be used for docking.

The docking is a process where the ligand is subjected to interact with the target biomolecule. The computer algorithm finds the best possible confirmation where the complex of ligand and biomolecule has minimum binding energy.

The essential condition for docking is that the selected ligand should possess the appropriate geometry to fit into the active site with minimal steric interactions. Favorable interactions of the chemical groups between the ligand and the active site of the biomolecule should be achieved [37, 38].

All the docking procedures carry out systematic searches in rotational and translational space and evaluate the electrostatic, hydrophobic, and hydrogen binding energy terms to achieve the best fit of the ligand. The procedures optimally position the ligand into the binding site and update the energy terms. The output of the docking is the binding energy (ΔG) of the ligand-receptor complex. The ligand showing the minimum binding energy is selected to proceed further [38].

To calculate the binding energy, the algorithm uses force field equations and parameters. The obtained binding energy (ΔG) is the total intermolecular interactions, including Van der Waals interaction, H- bonding interaction, electrostatic interaction, and internal static energy of the ligand. The equation is represented in equation number 2 [39 - 41].

```
\Delta G_{\text{bind}} = \Delta G_{\text{vdw}} + \Delta G_{\text{hydrophobic}} + \Delta G_{\text{H-bond}} + \Delta G_{\text{H-bond(chg)}} + \Delta G_{\text{deformation}} + \Delta G_0 - --(2)
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The docking result of Cyanidin as ligand with CDK4 as a receptor is shown in Fig. (4). Cyanidin is inhibiting CDK4 in the place of a natural inhibitor. This docking was performed using Autodock 4.0.

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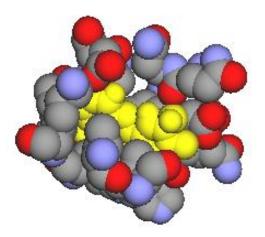


Fig. (4). Cyanidin $(C_{15}H_{10}O_6)$ is docked with CDK4 [36].

Table **3** shows the docking results of a few naturally occurring phenol compounds with CDK4 (PDB). The docking was performed using Autodock 4.0 [41]. Non-GA docking results are obtained using non-genetical algorithm whereas the GA docking results are obtained using the genetical algorithm.

S. No.	Phytochemical Name	Binding Energy (ΔG) in Kcal/Mol	
		Non-GA Docking	GA Docking
1	Apigenin	-9.96421	-10.5197
2	Baicalein	-10.176	-9.87955
3	Caffeic acid	-9.1941	-9.41513
4	Catechin	-10.1547	-10.4344
5	Chrisyn	-10.4538	-10.4848
6	Cyanidin	-10.0583	-10.7608
7	Daidzein	-9.99772	-11.0058
8	Diosmin	*	-10.5989
9	Ellagic acid	*	-9.54558
10	Emodin	-10.6053	-11.0401
11	Epicatechin	*	-9.82822
12	Epigallocatechin	*	-9.37263
13	Ferulic acid	-8.24349	-8.17937

Table 3. List of the observed binding energy of phenol-based ligands with 1GII [42].

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(Table 3) cont.....

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S. No.	Phytochemical Name Galangin	Binding Energy (ΔG) in Kcal/Mol	
14		-14.5898	-9.37035
15	Genistein	-9.34919	-9.3656
16	Hesperidin	*	-9.32745
17	Hydroxytyrosol	-8.65557	-9.35591
18	Kaempferol	-9.69367	-9.26569
19	Luteolin	-9.74437	-9.85704
20	Myricetin	-9.21467	-10.7346
21	Naringenine	-9.96618	-10.5528
22	Phloretin	-10.9983	-10.65
23	Quarcetin	-9.89271	-8.6555
24	Resveratrol	-11.0243	-10.5651
25	Rosmoric acid	-12.497	-12.011
26	Rottlerin	-10.8003	-13.9986
27	Rutin	*	-9.87545
28	Xanthohumol	-11.3567	-9.81086

DRUG DISCOVERY USING ARTIFICIAL NEURAL NETWORK

An artificial neural network(ANN) is a recent tool used in predicting the biological activities of molecules. ANN uses the available information related to the subject under study and provides the trained network. This trained network is used for the prediction of molecular properties or activities against a selected object.

The working process of ANN is just like the biological nervous system. In a biological system, such as the brain, there is a network of neurons responsible for the flow of information. ANN has also utilized the artificial network of neurons to flow the information [43].

ANN learns by example. It cannot predict without any learning process. Hence the first step in ANN is to train the network with known data. This process is called Network Training.

The training is for the specific application. Once the network is trained, it can be used to predict the properties in the domain of the application for which it is trained.

ANN is utilized to derive meaning from complicated data. The complex trends and patterns can be detected by ANN [44].

Once ANN is trained for a particular application, it can be used to predict the possible results by giving a new situation of interest [45].

Following are a few advantages of ANN

- Adaptive learning: Ability to learn from the given set of data for training
- Self-organization: ANN in process of learning creates its organization or representation of the information.
- Real-time operation: The training of the network and further use of the network can be carried out using a desktop computer. It is also possible to design special electronic hardware for the specific ANN task.

The architecture of Neural Network

A typical neural network has the following parts;

- Input layer
- Hidden Layer
- Output
- Neuron
- Connector

Fig. (5) depicts the simplest neural network. This network contains two layers. The first one is the input layer having three neurons. The second layer is the hidden layer contains two neurons. The first layer is also called the input layer. This layer receives the input from the system and delivers it to the hidden layer. In a hidden layer, each neuron receives the information from all input neurons. These inputs are 'weighted'.

Generally, a pre-set threshold value is set, which is used as a reference parameter for the forwarding of information from one layer to another. If the weighted input crosses the threshold value, then the information is passing onto the next layer. The algorithm to pass the information from one layer to other using weighted values may be very simple or may use complex mathematics. It all depends, how users design and define the ANN?

If the weighted input is not crossing the threshold value of a neuron, then the neuron does not send information to the next layer. In mathematical terms, the neuron fires only if X1.W1+X2.W2+X3.W3>Pre-set threshold Value as depicted

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in Fig. (5). The neural network may be as simple as shown in Fig. (5) or may have a quite large number of neurons in each layer. A certain network may have more than one hidden layer and that depends on the complexities of the problem that needs to be solved [44, 45].

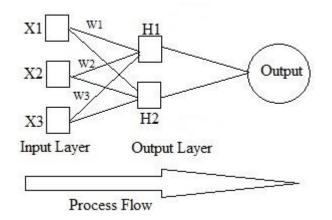


Fig. (5). A typical artificial neural network. [46].

While training the network, the final output is matched with the sample data. If it deviates then the weight of each neuron change using a specific function. This cycle continues until the network produces the desired output. The training of the network ends once the output matches with sample data with allowable standard deviation. The adaptive neural networks can be classified into two major categories depending on the way networks are being trained [47].

Supervised learning: In this method, the output is matched with the desired value. On comparing the obtained output with the desired output, the next step of learning decides. So between each cycle of learning, a watch process is set to understand whether the obtained output is matching with the desired output. The process of learning stops when the obtained output is matching with the desired output. Supervised learning includes error-correction learning, reinforcement learning, and stochastic learning.

Unsupervised learning: In the case of Unsupervised learning, the data are selforganized. The process of organization is based on local information.

In this case, the weight of each neuron is not changed. The point that needs to remember is that the trained network will work for the same domain of problem

Biologically Active Compounds

with the same type of input and expected output. Any deviation in the input parameter will require another trained network.

ANN experimental technique to map novel drug

The stepwise process of using ANN to map novel drugs is as follows:

- Select the list of existing drugs against a particular disease (For example, list of existing anti-cancer drugs)
- Prepare Molecular Electrostatic Potential of each drug using computer software
- Find out 12 points auto-correlation function
- Apply autocorrelation function to ANN
- Train the network
- Use this train network for the selection of unknown compounds as a novel drug against target disease.

In the case of the Auto-correlation function, molecular properties can be used for network training. The selection of molecular properties can be extracted using QSAR.

Another method is to use Kohonen neural network. This is a pattern recognition neural network. In this case, surface properties are used. For example, charge density can be used as an input pattern.

In this case, the pattern of the surface density of know drugs is supplied to the Kohonen network for training. Once the network is trained, the resultant pattern is used to identify the drug-likeness of the unknown molecule.

In the case of drug discovery, ANN can be used as a tool for predicting the novel drug. The simple process is explained here for anti-cancer drugs [47, 48].

- Select the list of anti-cancer drugs having certain common geometry. For example Alkylating antineoplastic agent.
- List the common properties along with biological activity against cancer cells.
- Apply these properties as a parameter to the backpropagation neural network and generate a trained network.
- Use this trained network for the selection of unknown compounds as an anticancer agent.

In the case of the Kohonen neural network, generate the MEP of each compound.

- Supply this MEP to the Kohonen neural network and generate the common pattern.
- Use this common pattern to screen a new anti-cancer agent (Fig. 6).

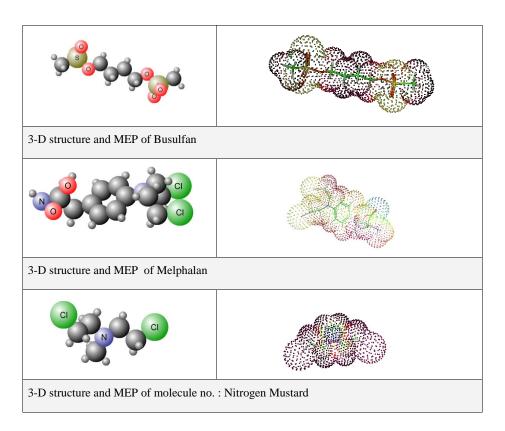


Fig. (6). 3-D structure and Molecular electrostatic potential of anti-cancer alkylating agents [46].

The Kohonen pattern of anti-cancer alkylating agents Busulfan, Melphalan, and Nitrogen Mustard were generated using SONNIA 4.2. Figure 7 shows the individual pattern of each molecule as well as an average pattern of all studied molecules. The average pattern can further be used to predict the anti-cancer activity in new molecules.

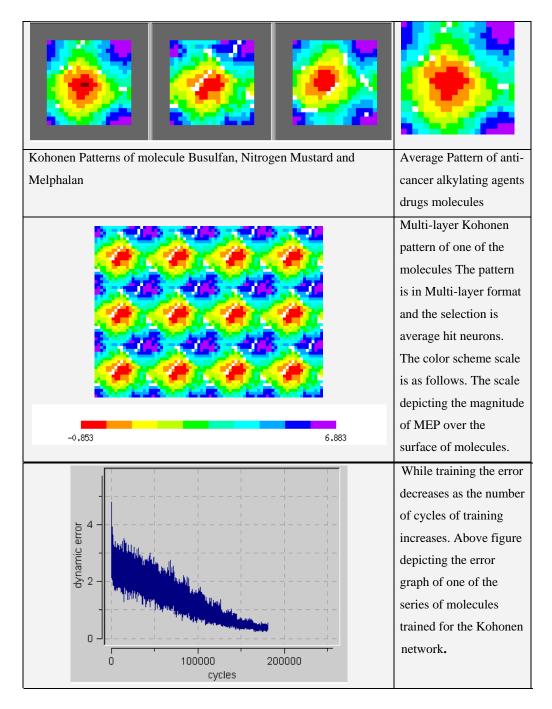


Fig. (7). Kohonen patter of the individual and average pattern obtained using Kohonen network [46].

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Numbers of computer software are available in the market to perform drug discovery using ANN. A few among them are SONNIA, Neural Designer, Darknet, Keras, *etc.*

Structure plays a vital role in acquiring the drug-likeness properties against a particular target. Knowledge of structure-function relationships provides the easiest path for designing novel drugs. For example, the majority of cancer drugs are originated from natural sources and their basic structure is of Di-terpene or Tri-terpene [49].

Flavonoids are very effective in preventing lipid peroxidation, whereas lipid peroxidation is caused by various diseases such as hepatotoxicity, inflation, atherosclerosis, diabetes, and aging [50].

Naturally occurring compounds play a vital role in managing human health. There is hardly any disease, which cannot be cured using the naturally occurring compound. There are still numerous compounds in nature, which are not explored for their drug-likeness properties. The structure-based drug discovery surely will help in providing unique drugs for each possible disease.

CONCLUSION

The drug discovery process has evolved with the progress of computer sciences. The technology of identifying the lead compound has matured and is widely used in a commercial establishment. The major techniques are Quantitative Structure-Activity Relationship (QSAR), Docking, MD simulation, and Artificial Neural Network.

Nature is the main source of obtaining novel drug molecules for various diseases. Numbers of new drug molecules are obtained from nature. This is possible due to the understanding of the structure-function relationship of the chemical compound obtained from nature.

The progress of computer algorithms helped in developing new state-of-art techniques to screen and identify novel drug molecules. The popular techniques are QSAR, Docking, MD Simulation, Artificial Neural Network (ANN), *etc*. These techniques are very much helpful in identifying the novel drug molecule in a very short time.

CONSENT FOR PUBLICATION

Not applicable.

Biologically Active Compounds

CONFLICT OF INTEREST

The authors declared that there is no conflict of interest.

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REFERENCES

- [1] Balick MJ, Cox PA. Plants, People, and Culture: the Science of Ethnobotany. New York, NY: Scientific American Library 1997.
- [2] Samuelsson G. Drugs of Natural Origin: a Textbook of Pharmacognosy. 5th Swedish Pharmaceutical Press Stockholm 2004.
- [3] Douglas Kinghorn A. Pharmacognosy in the 21st century. J Pharm Pharmacol 2001; 53(2): 135-48. [http://dx.doi.org/10.1211/0022357011775334] [PMID: 11273009]
- [4] Hansch C. A Quantitative Approach to Biochemical Structure-Activity Relationships. Acc Chem Res 1969; 2: 232-9.
 [http://dx.doi.org/10.1021/ar50020a002]
- [5] Hansch C, Fujita T. Analysis. A Method for the Correlation of Biological Activity and Chemical Structure. J Am Chem Soc 1964; 86: 1616-26.
 [http://dx.doi.org/10.1021/ja01062a035]
- [6] Free SMJ Jr, Wilson JW. A Mathematical Contribution to Structure-Activity Studies. J Med Chem 1964; 7: 395-9. [http://dx.doi.org/10.1021/jm00334a001] [PMID: 14221113]
- [7] Hammett LP. Some relations between reaction rates and equilibrium constants. Chem Rev 1935; 17(1): 125-36.

[http://dx.doi.org/10.1021/cr60056a010]

- [8] Hammett LP. The effect of structure upon the reactions of organic compounds. Benzene derivatives. J Am Chem Soc 1937; 59(1): 96-103.
 [http://dx.doi.org/10.1021/ja01280a022]
- Jing Yankang, *et al.* Deep Learning for Drug Design: an Artificial Intelligence Paradigm for Drug Discovery in the Big Data Era 2018. [http://dx.doi.org/10.1208/s12248-018-0210-0]
- [10] 10. The conceptual background and Development of Medicinal Chemistry by Alfred Burger, S.W. Dietrich, Med. Chem. Res., 2, 127-147(1992). (10).
- [11] 11. Annual Report on in Computational Chemistry, Volume 1, American Chemical Society, 2005.
- [12] http://www.scripps.edu/olson/forli/autodock_flex_rings.html
- [13] Rizvi SMD, Shakil S, Haneef M. A simple click by click protocol to perform docking: AutoDock 4.2 made easy for non-bioinformaticians. EXCLI J 2013; 12: 831-57.
 [PMID: 26648810]
- [14] http://www.schrodinger.com
- [15] Dineshkumar B, Kumar PV, Bhuvaneshwaran SP, Mitra A. Advanced drug designing software and their applications in medical research. Int J Pharm Pharm Sci 2010; 2: 16-8.
- [16] http://www.ccdc.cam.ac.uk/Solutions/GoldSuite/Pages/GOLD.aspx
- [17] Yuriev E, Agostino M, Ramsland PA. Challenges and advances in computational docking: 2009 in

review. J Mol Recognit 2011; 24(2): 149-64. [http://dx.doi.org/10.1002/jmr.1077] [PMID: 21360606]

- [18] http://www.serc.iisc.ernet.in/facilities/ComputingFacilities/software/biosuite.html
- [19] BioSuite: a comprehensive bioinformatics software package (A unique industry-academia collaboration). Curr Sci 2007; 92: 29-38.
- [20] https://www.schrodinger.com/Maestro
- [21] Sabitha K, Habeeb SKM. Molecular modeling and drug discovery of potential inhibitor for anticancer target gene Melk (Maternal Embryonic Leucine Zipper Kinase). Int Res J Pharmacy 2011; 2: 141-5.
- [22] http://www.arguslab.com/arguslab.com/ArgusLab.html
- [23] Mathew GE, George S, Shamnas M, Raj VBA. 2012.
- [24] http://vakser.bioinformatics.ku.edu/main/resources_gramm1.03.php
- [25] Kundrotas PJ, Vakser IA. Accuracy of protein-protein binding sites in high-throughput template-based modeling. PLOS Comput Biol 2010; 6(4): e1000727.
 [http://dx.doi.org/10.1371/journal.pcbi.1000727] [PMID: 20369011]
- [26] http://www.certara.com/products/molmod/sybyl-x
- [27] Dubey A, Kalra SS. Computational comparative modeling and visualization for HIV1 and HIV2 proteins via the software SYBYL-X. Int J Sci Res Publ 2013; 3: 2250-3153.
- [28] http://www.scfbio-iitd.res.in/sanjeevini/sanjeevini.jsp
- [29] Jayaram B, Singh T, Mukherjee G, Mathur A, Shekhar S, Shekhar V. Sanjeevini: a freely accessible web-server for target directed lead molecule discovery. BMC Bioinformatics 2012; 13 (Suppl. 17): S7. [http://dx.doi.org/10.1186/1471-2105-13-S17-S7] [PMID: 23282245]
- [30] http://www.genexplain.com/pass
- [31] Pramely R, Raj LS. Prediction of biological activity spectra of a few phytoconstituents of Azadirachta indicia A. Juss. J Biochem Technol 2012; 3: 375-9.
- [32] Jamkhandea PG, Ghantea MH, Ajgundeb BR. Software-based approaches for drug designing and development: A systematic review on commonly used software and its applications. Bull Fac Pharm Cairo Univ 2017; 55(2): 203-10. [http://dx.doi.org/10.1016/j.bfopcu.2017.10.001]
- [33] Mir S, Alhroub Y, Anyango S, et al. PDBe: towards reusable data delivery infrastructure at protein data bank in Europe. Nucleic Acids Res 2018; 46(D1): D486-92. [http://dx.doi.org/10.1093/nar/gkx1070] [PMID: 29126160]
- [34] Meng X-Y, Zhang H-X, Mezei M, Cui M. Molecular docking: a powerful approach for structure-based drug discovery. Curr Computeraided Drug Des 2011; 7(2): 146-57. [http://dx.doi.org/10.2174/157340911795677602] [PMID: 21534921]
- [35] Kirkpatrick P, Screening V. Gliding to Success. Nat Rev Drug Discov 2004; 3: 299. [http://dx.doi.org/10.1038/nrd1364]
- [36] Amita S. Critical studies of the effectiveness of natural products as an anti-cancer drug using the computer-based model system 2010.
- [37] Warren G L, Andrews CW, Capelli AM, et al. A Critical Assessment of docking programs and scoring functions, J Med Chem. 2006 Oct, 5; 49(20), 5912-31 2006.
- [38] Perola E, Walters WP, Charifson PS. A detailed comparison of current docking and scoring methods on systems of pharmaceutical relevance. Proteins 2004; 56(2): 235-49. [http://dx.doi.org/10.1002/prot.20088] [PMID: 15211508]
- [39] Muegge I, Martin YC. A general and fast scoring function for protein-ligand interactions: a simplified

Sunil H. Ganatra

potential approach. J Med Chem 1999; 42(5): 791-804. [http://dx.doi.org/10.1021/jm980536j] [PMID: 10072678]

- [40] Terp GE, Johansen BN, Christensen IT, Jørgensen FS. A new concept for multidimensional selection of ligand conformations (MultiSelect) and multidimensional scoring (MultiScore) of protein-ligand binding affinities. J Med Chem 2001; 44(14): 2333-43. [http://dx.doi.org/10.1021/jm0010901] [PMID: 11428927]
- [41] Morris GM, Huey R, Lindstrom W, et al. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. J Comput Chem 2009; 30(16): 2785-91. [http://dx.doi.org/10.1002/jcc.21256] [PMID: 19399780]
- [42] Ganatra SH, Suchak AS, Gurjar S. Inhibition studies of naturally occurring phenol-based compounds with cell cycle regulator enzyme using molecular modeling techniques. J Pharm Res 2012; 5(9): 4786-90.
- [43] Lipkowitz KB, Boyd DB. Reviews in Computational Chemistry" Volume 6, Willey Publication, 1995.
- Baskin II, Winkler D, Tetko IV. A renaissance of neural networks in drug discovery. Expert Opin Drug Discov 2016; 11(8): 785-95.
 [http://dx.doi.org/10.1080/17460441.2016.1201262] [PMID: 27295548]
- Silver D, Huang A, Maddison CJ, et al. Mastering the game of Go with deep neural networks and tree search. Nature 2016; 529(7587): 484-9.
 [http://dx.doi.org/10.1038/nature16961] [PMID: 26819042]
- [46] Ganatra SH. Studies and development of anti-cancer drug design using modeling techniques based on artificial neural network (ANN), Project Report. Research Award 2007; pp. 58-70.
- [47] Xu Y, Li X, Yao H, Lin K. Neural networks in drug discovery: current insights from medicinal chemists. Future Med Chem 2019; 11(14): 1669-72. [http://dx.doi.org/10.4155/fmc-2019-0118] [PMID: 31287735]
- [48] Merk D, Friedrich L, Grisoni F, Schneider G. De novo design of bioactive small molecules by artificial intelligence. Mol Inform 2018; 37(1-2): 1700153. [http://dx.doi.org/10.1002/minf.201700153] [PMID: 29319225]
- [49] Ganatra SH, Suchak AS. Inhibition studies of naturally occurring Terpene based compounds with cyclin-dependent kinase 2 enzyme, J Comput Sci Syst Biol, 2012;5(3),068-078.
- [50] Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. J Nutr Sci. 2016; 5: e47. Published 2016 Dec 29.
 [http://dx.doi.org/10.1017/jns.2016.41]

Turmeric Supplementation and Its Valued Clinical Connections

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Abstract: Turmeric (*Curcuma longa*) does not require any introduction for its benefits because it is an extremely renowned spice and cooking material of Indian kitchens. It has a valuable place in Ayurveda for its crude medicinal values. In India, it also has a sacred position in festivals, worships, and wedding ceremonies. For a long time, it is being used as an important ingredient in different Asian dishes and has a significant position in the cooking spices. Except for its valuable uses as a spice, it is known for its role in wound treatment, anti-inflammatory and anti-oxidant, pain relief, anti-cancer, and so on. It is being used for a long time with several expectations of its great health benefits, but there is still no concrete research that proves its heavy potency towards the treatment of any serious disease. Although it is not so potent individually for the treatment of any serious health issues, its supplemental values must be encouraged, and more research is essential to be done on it. This chapter concisely demonstrates the significance of turmeric in the treatment of various health issues and its role in food supplements.

Keywords: Anti-inflammatory, Antioxidant, Arthritis, Curcumin, Indigestion, Spice, Traditional medicine, Turmeric, Wound healing.

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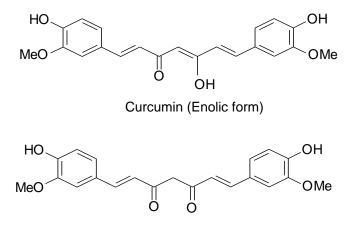
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INTRODUCTION

Turmeric does not require any special introduction. It is a renowned name in the field of traditional medication and Ayurveda. It plays an integral part in Indian kitchens for its use as a valuable spice. It is beneficial in curing various human problems and improving human health. Turmeric is famous in Ayurveda for its role in controlling and balancing the Vata, pitta, and kapha, and has useful effects on the blood and plasma of the circulatory system. It is highly helpful in the treatment of cough-related problems as well as in toxins removal. It is known for its good inflammatory properties, wound healing properties, antiseptic and antibacterial properties, arthritis problems, indigestion problems, beneficial for the (https://maharishiayurvedaindia.com/blogs/ayurvedabrains. and SO on knowledge-center/countless-benefits-of-turmeric) [1]. Turmeric is related to the Zingiberaceae (ginger family), generally known as *Curcuma longa*, and its roots are used in cooking [2]. It is a perennial, rhizomatous, and herbaceous plant native to the Indian subcontinent and Southeast Asia (https://en.wikipedia. org/wiki/Turmeric) [3].

Turmeric is used in the kitchen for coloring and flavoring the food by drying and grounding its rhizome in the form of yellow-orange powder. It is significantly being used in Ayurveda, Siddha medicine, Traditional Chinese Medicines, Unani [4], and other traditional/folk medications. Turmeric was also known as Indian saffron in Medieval Europe [5]. Curcumin, a biologically and pharmaceutically active molecule of turmeric, is responsible for its beautiful golden yellow color. It generally exists in keto and enol forms and is the main curcuminoid of turmeric (https://en.wikipedia.org/wiki/Curcumin) [3b]. Fig. (1) shows the chemical structure of keto and enol form of curcumin, a biologically active component of turmeric [3b].

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Curcumin (Keto form)

Fig. (1). Enol and Keto forms of the biologically active compound 'curcumin'.

Recent Research Updates

Although turmeric is known for its attractive color, its wide use in food for enhancing color and flavor, its nutraceutical virtues, and its use as traditional and folk medications, there are not many scientific proofs, which can prove its strong role in the cure of human problems. However, at <u>a</u> supplemental position, research strongly supports its clinical uses.

A good mechanistic review has been presented by Ahmad *et al.* (2020) [6] on biochemistry, safety, pharmacological activities, and clinical applications of turmeric. They have summarized the scientifically known studies on the aforesaid title. They reviewed the pharmaceutical roles of turmeric as its anti-inflammatory, anti-oxidant, anti-cancer, anti-mutagenic, anti-microbial, anti-obesity, hypolipidemic, cardio-protective, and neuro-protective effects and showed that due to these pharmaceutical roles of turmeric, it is a great spice for future research [6].

Rathore *et al.* (2020) [7] reviewed curcumin and its many benefits for health as well as its different biological activities such as anti-inflammatory, anti-oxidant, anti-arthritis, anti-cancer, wound-healing, anti-bacterial, anti-viral, in depression, anti-diabetic, anti-venom, anti-obesity, anti-asthmatic, and other. Observations of different clinical trials, various roles, and activities of curcumin have been discussed in this review [7].

An informative page on the title "Curcumin is the spice of life when delivered *via* tiny nanoparticles: Treatment for Alzheimer's and genital herpes" shows the

significance of curcumin (www.sciencedaily.com/releases/2020/03/20030513 2144.htm) [8].

Vitali *et al.* (2020) [9] presented a very informative work on curcumin with the title "Curcumin Can Decrease Tissue Inflammation and the Severity of HSV-2 (Herpes Simplex Virus Type 2) Infection in the Female Reproductive Mucosa". They investigated whether curcumin, encapsulated in nanoparticles and delivered by various *in vivo* routes, could minimize inflammation and prevent or reduce HSV-2 infection in the Female Genital Tract. Their result suggested that curcumin nanoparticle delivery in the vaginal tract could reduce local tissue inflammation [9].

Zahedipour *et al.* (2020) [10] wrote a review in order to show the effective role of curcumin in the cure of infections caused by COVID 19, which is acute respiratory syndrome coronavirus 2 (SARS-CoV-2) with considerable mortality worldwide. In this infection, symptoms produced are mostly related to a respiratory system such as the lungs. In this review, they have given an overview of the antiviral effects of curcumin, which can modulate a range of molecular targets, which is the reason behind its possible efficacy for the cure of this pandemic [10].

A review article written by Sharif-Rad <u>et al</u>. (2020) [11] deals with the role of turmeric and its main constituent 'curcumin' on health, considering their bioactive effects, safety profiles for food, pharmaceutical, and biotechnological applications. They have an in-depth discussion on curcumin for its food and biotechnological applications with its potential role in health and disease cure has a special emphasis on its biological activities like anti-inflammatory, anti-oxidant, anti-cancer, neuroprotective, hepatoprotective, and cardio-protective effects. They have also covered the multidimensional role of curcumin, agro-industrial procedures to offset its instability and low bioavailability, food attractiveness optimization, upcoming strategies for clinical application, and health concerns [11].

A review has been written by Stohs *et al.* (2020) [12] on highly bioavailable forms of curcumin and promising avenues for curcumin-based research and application in which they have discussed formulations designed to enhance the bioavailability, metabolism of curcumin, relationships between solubility and particle size relative to bioavailability, human pharmacokinetic studies involving formulated curcumin products, the widely used but inappropriate practice of hydrolyzing plasma samples for quantification of blood curcumin, current applications of curcumin and its metabolites, and promising directions for health maintenance and applications [12].

Shanmugarajan *et al.* (2020) [13] performed computational modelling, simulations, and ADMET studies to explore curcuminoids against novel SARS-CoV-2 targets. Bioactive ingredients (curcuminoids) present in *C. longa* are known for their different pharmacological properties, and fourteen curcuminoids were studied with the possibility of their role in SARS-CoV-2 inhibition. They performed a study on their *in-silico* properties towards SARS-CoV-2 target proteins by homology modelling, ADME, drug-likeness, toxicity predictions, docking molecular dynamics simulations, and MM-PBSA free energy estimation. This study shows that surface proteins are key drug target proteins of SARS-Co-2, and probably curcumin blocks essential biologically active drug target residues, thereby attenuating the viral infection.

Fadus *et al.* (2017) [14] wrote a review on tile "Curcumin: An age-old antiinflammatory and anti-neoplastic agent" in order to demonstrate the various pharmacological roles of curcumin, a biologically active compound of turmeric, which has a valued role in the cure of chronic conditions like **rheumatoid arthritis**, **inflammatory bowel disease**, Alzheimer's, and common malignancies like colon, lung, stomach, skin, and breast cancers. In this review, they showed the scientific information on the medicinal role of curcumin (the "curry spice") and identified the current gap [14]. A review on the mechanism of antiinflammatory effects of curcumin was written by Jacob *et al.* (2007) [15], in which they very finely highlighted the anti-inflammatory role of curcumin. The beneficial effect of curcumin in sepsis appears to be mediated by the upregulation of PPAR- γ (peroxisome proliferator-activated receptor- γ), leading to the suppression of pro-inflammatory cytokine, TNF- α expression, and release [15].

There are also a number of other research in the field of turmeric and curcumin and their clinical applications, but only a few recent reports have been summarized in this chapter in order to show the potency of turmeric and its constituent curcumin in the medicinal field. For more information and research updates on turmeric and its uses in the pharmaceutical application, the reader can also visit the works of Kim *et al.* (2020) [16] and Kocaadam & **Şanlier** (2017) [17]. There are other different articles that may be useful for readers [18 - 22]. A page showing ten important benefits of turmeric use can also be visited (https://www.healthline.com/nutrition/top-10-evidence-based-health-benefits-ofturmeric) [23].

CONCLUSION

In this way, the aforementioned studies on the pharmaceutical role of turmeric and its biologically active organic molecule 'curcumin' clearly demonstrate the great significance of turmeric in solving various human health issues. Although it is not a perfect medication, individually, its role as great supplemental food should be encouraged, and scientific research works should also be encouraged in order to search out its major valuable roles in treating the different types of health-related issues as well as in the purification of the human system for saving the human from various problems and enhancing their immunity.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The authors declared that there is no conflict of interest.

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REFERENCES

- [1] Aurveda M. 2020. https://maharishiayurvedaindia.com/blogs/ayurveda-knowledge-center/countlessbenefits-of-turmeric
- Priyadarsini KI. The chemistry of curcumin: from extraction to therapeutic agent. Molecules 2014; 19 (12): 20091-112.
 [http://dx.doi.org/10.2200/mslcmlcs1012200011 [DMID: 2547027(1)]

[http://dx.doi.org/10.3390/molecules191220091] [PMID: 25470276]

- [3] (a)Turmeric W. 2020.https://en.wikipedia.org/wiki/Turmeric (b) Curcumin W. 2020. https://en.wikipedia.org/wiki/Curcumin
- [4] Chattopadhyay I, Kaushik B, Uday B, Ranajit KB. Turmeric and curcumin: Biological actions and medicinal applications. Curr Sci 2004; 87(1): 44-53.
- [5] Ghillean Prance, Mark Nesbitt. The Cultural History of Plants. Routledge. 2005; p. 170.
- [6] Ahmad RS, Hussain MB, Sultan MT, et al. Biochemistry, Safety, Pharmacological Activities, and Clinical Applications of Turmeric: A Mechanistic Review. Evid-Based Complement Alternat Med. vol. 2020, Article ID 7656919, 14 pages. [http://dx.doi.org/10.1155/2020/7656919]
- [7] Rathore S, Mukim M, Sharma P, Devi S, Nagar JC, Khalid M. Curcumin: A Review for Health Benefits. Int J Res Rev 2020; 7(1): 273-90.
- [8] University of South Australia. Curcumin is the spice of life when delivered via tiny nanoparticles: Treatment for Alzheimer's and genital herpes. ScienceDaily 2020. www.sciencedaily.com/releases/ 2020/03/200305132144.htm
- [9] Vitali D, Bagri P, Wessels JM, et al. Curcumin Can Decrease Tissue Inflammation and the Severity of HSV-2 Infection in the Female Reproductive Mucosa. Int J Mol Sci 2020; 21(1): 337. [http://dx.doi.org/10.3390/ijms21010337] [PMID: 31947962]

- Zahedipour F, Hosseini SA, Sathyapalan T, *et al.* Potential effects of curcumin in the treatment of COVID-19 infection. Phytother Res 2020; 1-10. [http://dx.doi.org/10.1002/pt r.6738]
- Sharifi-Rad J, Rayess YE, Rizk AA, *et al.* Turmeric and its major compound curcumin on health: bioactive effects and safety profiles for food, pharmaceutical, biotechnological and medicinal applications. Front Pharmacol 2020; 11: 01021.
 [http://dx.doi.org/10.3389/fphar.2020.01021] [PMID: 33041781]
- [12] Stohs SJ, Chen O, Ray SD, Ji J, Bucci LR, Preuss HG. Highly bioavailable forms of curcumin and promising avenues for curcumin-based research and application: a review. Molecules 2020; 25(6): 1397.
 [1] Hard(1) doi: org(10.2200/product log/2020/12071/JDMDb. 2220/22721)

[http://dx.doi.org/10.3390/molecules25061397] [PMID: 32204372]

- [13] Shanmugarajan D, Prabitha P, Prashantha Kumar BR, Suresh B. Curcumin to inhibit binding of spike glycoprotein to ACE2 receptors: computational modelling, simulations, and ADMET studies to explore curcuminoids against novel SARS-CoV-2 targets. RSC Advances 2020; 10: 31385-99. [http://dx.doi.org/10.1039/D0RA03167D]
- [14] Fadus MC, Lau C, Bikhchandani J, Lynch HT. Curcumin: An age-old anti-inflammatory and antineoplastic agent. J Tradit Complement Med 2016; 7(3): 339-46. [http://dx.doi.org/10.1016/j.jtcme.2016.08.002] [PMID: 28725630]
- [15] Jacob A, Wu R, Zhou M, Wang P. Mechanism of the Anti-inflammatory Effect of Curcumin: PPARγActivation PPAR Research Volume 2007. Article ID 89369, 5 pages. [http://dx.doi.org/10.1155/2007/89369]
- [16] Kim H, Ban I, Choi Y, et al. Puffing of Turmeric (Curcuma longa L.) Enhances its Anti-Inflammatory Effects by Upregulating Macrophage Oxidative Phosphorylation. Antioxidants 2020; 9(10): 931. [http://dx.doi.org/10.3390/antiox9100931] [PMID: 33003300]
- Kocaadam B, Şanlier N. Curcumin, an active component of turmeric (*Curcuma longa*), and its effects on health. Crit Rev Food Sci Nutr 2017; 57(13): 2889-95.
 [http://dx.doi.org/10.1080/10408398.2015.1077195] [PMID: 26528921]
- [18] Lee G-H, Lee H-Y, Choi M-K, Chung H-W, Kim S-W, Chae H-J. Protective effect of *Curcuma longa* L. extract on CCl₄-induced acute hepatic stress. BMC Res Notes 2017; 10(1): 77. [http://dx.doi.org/10.1186/s13104-017-2409-z] [PMID: 28143589]
- [19] Khan H, Ullah H, Nabavi SM. Mechanistic insights of hepatoprotective effects of curcumin: Therapeutic updates and future prospects. Food Chem Toxicol 2019; 124: 182-91. [http://dx.doi.org/10.1016/j.fct.2018.12.002] [PMID: 30529260]
- [20] Farzaei MH, Zobeiri M, Parvizi F, et al. Curcumin in liver diseases: a systematic review of the cellular mechanisms of oxidative stress and clinical perspective. Nutrients 2018; 10(7): 855. [http://dx.doi.org/10.3390/nu10070855] [PMID: 29966389]
- [21] Kandezi N, Mohammadi M, Ghaffari M, Gholami M, Motaghinejad M, Safari S. Novel Insight to Neuroprotective Potential of Curcumin: A Mechanistic Review of Possible Involvement of Mitochondrial Biogenesis and PI3/Akt/ GSK3 or PI3/Akt/CREB/BDNF Signaling Pathways. Int J Mol Cell Med 2020; 9(1): 1-32. [http://dx.doi.org/10.22088/IJMCM.BUMS.9.1.1] [PMID: 32832482]
- [22] Blanton C, Gordon B. Effect of Morning vs. Evening Turmeric Consumption on Urine Oxidative Stress Biomarkers in Obese, Middle-Aged Adults: A Feasibility Study. Int J Environ Res Public Health 2020; 17(11): 4088. [http://dx.doi.org/10.3390/ijerph17114088] [PMID: 32521782]
- [23] Healthline, 10 Proven Health Benefits of Turmeric and Curcumin, written by Kris Gunnars. 2018. https://www.healthline.com/nutrition/top-10-evidence-based-health-benefits-of-turmeric

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